Elevated Temperature and CO2 Concentrations Affect Carbon Flux in Two Boreal Conifers

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A thesis submitted in partial fulfillment of the requirements for the Master of Science degree in Biology
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Abstract

Elevated temperatures and CO\textsubscript{2} alter carbon flux in two dominant boreal tree species \textit{Picea mariana} (black spruce) and \textit{Larix laricina} (tamarack). Seedlings were grown in three temperature treatments (ambient, ambient +4 °C, and ambient +8 °C) at either 400 ppm or 750 ppm CO\textsubscript{2}, to simulate climate conditions between now and the year 2100. Spruce acclimated to increasing temperature detractively; warming scenarios reduced spruce net carbon gain. Tamarack maintained comparable levels of net photosynthesis (\textit{A\textsubscript{net}}) across warming treatments and both species reduced respiration (\textit{R\textsubscript{dark}}) with increasing growth temperature. Elevated CO\textsubscript{2}-grown spruce suppressed \textit{A\textsubscript{net}} whereas \textit{A\textsubscript{net}} in tamarack was unresponsive to elevated CO\textsubscript{2}. Decreasing leaf N with warming explained reduced \textit{A\textsubscript{net}} and \textit{R\textsubscript{dark}} for both species; however, tamarack mitigated this by increasing stomatal conductance. Moderate warming benefited tamarack growth but hindered spruce; extreme warming hindered growth in both species. Reduced CO\textsubscript{2} uptake in these species with predicted warming may contribute to increased atmospheric CO\textsubscript{2} accumulation.

Keywords

Climate change, \textit{Picea mariana}, \textit{Larix laricina}, carbon assimilation, carbon flux, photosynthesis, respiration, acclimation, temperature, CO\textsubscript{2}.
Co-Authorship Statement

Experimental design was conceived with Dr. Danielle Way. I executed experiments, data analysis and interpretation, and the writing of this thesis. Editing was done by Drs. Danielle Way and Sheila Macfie. This project was part of a larger collaboration with Eric Dusenge.
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<th>Definition</th>
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<tbody>
<tr>
<td>AC</td>
<td>Ambient CO\textsubscript{2} concentration</td>
</tr>
<tr>
<td>\textit{A}_\textit{net}</td>
<td>Net CO\textsubscript{2} assimilation rate</td>
</tr>
<tr>
<td>\textit{A}_\textit{opt}</td>
<td>Maximum \textit{A}_\textit{net}</td>
</tr>
<tr>
<td>ADP</td>
<td>Adenosine diphosphate</td>
</tr>
<tr>
<td>ATP</td>
<td>Adenosine triphosphate</td>
</tr>
<tr>
<td>\textit{C}_\textsubscript{a}</td>
<td>Atmospheric CO\textsubscript{2} concentration</td>
</tr>
<tr>
<td>\textit{C}_\textsubscript{i}</td>
<td>Intercellular CO\textsubscript{2} concentration</td>
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<tr>
<td>CO\textsubscript{2}</td>
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<tr>
<td>E</td>
<td>Transpiration rate</td>
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<tr>
<td>EC</td>
<td>Elevated CO\textsubscript{2} concentration (750 ppm)</td>
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<tr>
<td>FADH\textsubscript{2}</td>
<td>Flavin adenine dinucleotide</td>
</tr>
<tr>
<td>FNR</td>
<td>Ferredoxin-NADP+ reductase</td>
</tr>
<tr>
<td>G3P</td>
<td>Glyceraldehyde 3-phosphate</td>
</tr>
<tr>
<td>g\textsubscript{s}</td>
<td>Stomatal conductance</td>
</tr>
<tr>
<td>H</td>
<td>Hydrogen</td>
</tr>
<tr>
<td>LMA</td>
<td>Leaf mass per unit area</td>
</tr>
<tr>
<td>N</td>
<td>Nitrogen</td>
</tr>
<tr>
<td>NAD</td>
<td>Nicotinamide adenine dinucleotide</td>
</tr>
<tr>
<td>NADP</td>
<td>Nicotinamide adenine dinucleotide phosphate</td>
</tr>
<tr>
<td>O\textsubscript{2}</td>
<td>Oxygen</td>
</tr>
<tr>
<td>P\textsubscript{i}</td>
<td>Inorganic phosphate</td>
</tr>
<tr>
<td>PSI</td>
<td>Photosystem I</td>
</tr>
<tr>
<td>PSII</td>
<td>Photosystem II</td>
</tr>
<tr>
<td>Symbol</td>
<td>Definition</td>
</tr>
<tr>
<td>--------</td>
<td>------------</td>
</tr>
<tr>
<td>$R_{\text{dark}}$</td>
<td>Dark respiration rate</td>
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<tr>
<td>$R_{25}$</td>
<td>$R_{\text{dark}}$ measured at 25 °C</td>
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<tr>
<td>Rubisco</td>
<td>Ribulose-1, 5-bisphosphate carboxylase/oxygenase</td>
</tr>
<tr>
<td>RuBP</td>
<td>Ribulose-1, 5-bisphosphate</td>
</tr>
<tr>
<td>T0</td>
<td>Ambient temperature</td>
</tr>
<tr>
<td>T4</td>
<td>T0 + 4 °C (moderate warming)</td>
</tr>
<tr>
<td>T8</td>
<td>T0 + 8 °C (extreme warming)</td>
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<tr>
<td>$T_m$</td>
<td>Measurement temperature</td>
</tr>
<tr>
<td>$T_{\text{max}}$</td>
<td>High temperature CO$_2$ compensation point</td>
</tr>
<tr>
<td>$T_{\text{opt}}$</td>
<td>Thermal optimum of $A_{\text{net}}$</td>
</tr>
<tr>
<td>VPD</td>
<td>Vapour pressure deficit</td>
</tr>
<tr>
<td>WUE</td>
<td>Water use efficiency</td>
</tr>
<tr>
<td>2PG</td>
<td>2-phosphoglycolate</td>
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Chapter 1: Introduction

1.1 Climate change

The widespread combustion of fossil fuels and land use change since the Industrial Revolution have caused atmospheric carbon dioxide (CO$_2$) concentrations to increase from 280 ppm in 1880 to 400 ppm in 2016 (IPCC, 2014). Combustion of fossil fuels, such as coal, natural gas and oil results in the production of CO$_2$, and autotrophic photosynthesis is the primary process by which CO$_2$ is removed from the atmosphere. While atmospheric CO$_2$ concentrations have only increased since the Industrial Revolution, urban and agricultural expansion have resulted in widespread deforestation around the world (Seto et al. 2012), reducing plant photosynthetic CO$_2$ uptake and long-term CO$_2$ sequestration. The accumulation of CO$_2$ in the atmosphere traps radiation emitted from the earth’s surface below the atmosphere, driving the increase in global surface temperature.

Since 1880, the global average surface temperature has increased by 0.65 °C and, if current CO$_2$ emission trends continue, it is predicted to climb a further 4 °C by the end of the 21$^{st}$ century (Ciais et al. 2014). By 2100, atmospheric CO$_2$ concentrations are predicted to reach between 550 and 750 ppm, depending on the emissions scenario used (IPCC, 2014). Even if worldwide greenhouse gas emissions ceased, global temperatures would still rise by up to 1.7 °C by the end of the 21$^{st}$ century – a legacy of previously-emitted CO$_2$ lingering in the atmosphere (IPCC, 2014). Warming is predicted to be greater at high latitudes such that northern regions, such as the Arctic and boreal forest,
may warm up to 8 °C by 2100, partially due to a diminishing albedo effect as arctic sea ice melts and causes a positive feedback on regional temperature (Curry et al. 1995).

1.2 The boreal forest
The boreal forest is the largest terrestrial biome on the planet, and is characterized by long, harsh winters, short summers, strong seasonal climatic variability, and low precipitation. Tree species in the Canadian boreal forest are chiefly conifers, such as black spruce (*Picea mariana*), jack pine (*Pinus banksiana*), and tamarack (*Larix laricina*); some broadleaved species such as trembling aspen (*Populus tremuloides*) and paper birch (*Betula papyrifera*) also inhabit the forest (Tjoelker et al. 1998). Dominance of the forest stands by conifer trees leads to acidic soils, as decomposing needles release defensive compounds such as phenols, preventing many other plant types from growing on the forest floor (Rice and Pancholy 1974, Kuiters and Denneman 1987). The boreal forest faces regular, large-scale disturbances from forest fires and insect outbreaks (Kasischke et al. 1995, Bogdanski 2008), and is home to some of North America’s most charismatic megafauna: wolves, caribou, grizzly bears, and lynxes. The extensive wetlands and peat bogs in the boreal forest are an integral part of both water cycling and filtration, as well as providing habitat to many species of waterfowl and other migratory birds (Bogdanski 2008).

In Canada, the boreal forest covers over half of the country’s landmass and accounts for 77% of the total forested area (Kurz and Apps 1999, Peng et al. 2011). 40 percent of the
nation’s total wood production comes from the boreal forest, totalling an annual revenue of CA$41 billion and nearly 130,000 jobs (Bogdanski 2008). Beyond forestry, the boreal sector encompasses billions’ worth of mining, agriculture, and hydroelectric energy, and sustains hundreds of thousands of jobs (Bogdanski 2008). The national-scale economics of the boreal forest are vast, but are perhaps eclipsed by the global significance of the forest in terms of atmospheric carbon dynamics. In Canada alone, the boreal forest stores 186 Gt of carbon in its living biomass, detritus, soils, and peat (Apps et al. 1993); it is estimated that 714 Gt of carbon are stored in the entire biome (Kasischke et al. 1995). Annually, the Canadian boreal drawdown of atmospheric CO₂ is approximately 95 Tg, making it a substantial carbon sink (Apps et al. 1993).

1.3 Black spruce and tamarack

Black spruce (Picea mariana) and tamarack (Larix laricina) are two dominant boreal conifer (cone-bearing) species; the former is evergreen, while the latter is a species of larch, and a deciduous conifer. Both species are gymnosperms from the Pinaceae (pine) family and are found throughout Canada and parts of Alaska – close relatives of these species inhabit the taiga of Eurasia (Gower and Richards 1990). Conifers have needle-shaped leaves and thrive in the harsh boreal environments where most broad-leaved tree species, excepting poplar and birch, cannot. High nutrient- and water-use efficiencies allow conifers to grow in boreal soils which are nutrient poor and often frozen or underlain by permafrost, limiting water uptake (Gower and Richards 1990, Givnish 2002). Nutrient limitation is partially caused by low rates of litter decomposition in cold temperatures (Kurz and Apps 1999, Robinson 2002). Both the evergreen and deciduous
leaf strategies allow for advantages and trade-offs which ultimately dictate species’ ability to survive and thrive in the harsh, cold climates of the boreal forest.

Evergreen needles have long lifespans and must withstand winter conditions which deciduous leaves never experience. By investing in structural compounds, such as thick cuticles, evergreen needles are able to endure wind shear, ice crystal abrasion, and snow accumulation, but this comes at the expense of developing more metabolic machinery (Givnish 2002, Van Ommen Kloke et al. 2012). Evergreen needles typically have a low nitrogen concentration, making them less appealing to herbivores (Mooney and Gulmon 1982). By investing in hardy, year-round needles, evergreens are able to gain carbon steadily throughout winter, albeit at a slower rate than during the summer months (Van Ommen Kloke et al. 2012). Photosynthesis ceases in evergreens when the ground freezes in winter and water lost from transpiration cannot be replenished (Troeng and Linder 1982). Water supplies the electrons to chlorophyll in the light-dependent reactions of photosynthesis (see Section 1.4.1) and frozen soil precludes water uptake by the roots, temporarily halting the electron transport chain until water becomes available once more.

In contrast to black spruce, photosynthetic carbon gain in tamarack is constrained by the limited growing season when needles are present. As such, tamarack need not invest as heavily in structural tissue as an evergreen species, and instead generates “cheaper” leaves which are able to photosynthesize at considerably higher rates than its sympatric evergreen counterparts (Gower and Richards 1990, Givnish 2002, Van Ommen Kloke et
al. 2012). By allocating less carbon to the structure of its needles, tamarack is able to build more leaves, allowing for fast growth rates; increased CO₂ diffusion through the comparatively thin needles of tamarack facilitates higher rates of photosynthesis, aided by high nitrogen investment per unit of leaf area. The deciduous strategy is consistent with opportunistic development during a shortened growing season: rapid, plastic response to environmental triggers, high rates of photosynthesis, and low leaf mass per unit area (Van Ommen Kloke et al. 2012). By comparison, the evergreen strategy is conservative, both in metabolic rates and in response to environmental stimuli. Annual foliage regeneration is a costly process both energetically and in nutrient requirement, so evergreens are the predominant tree type of the boreal forest.

While the highly seasonal boreal conditions may be thought to favour an evergreen strategy, tamarack aboveground net primary productivity (NPP; gross carbon uptake minus respiratory carbon losses) exceeds that of deciduous broadleaf species and is commensurate with evergreen conifers (Gower and Richards 1990, Wright et al. 2004). Larch species inhabit boreal forests around the world and are able to optimize growth during a short growing season, allowing them to compete successfully with evergreen conifers. Allocation to root mass is lower in tamaracks than evergreen conifers, allowing them to compete for light by growing upwards and developing a full foliage (Gower and Richards 1990). Tamaracks have similar nitrogen use efficiency to evergreen conifers, but foliar nitrogen concentrations are up to 50% higher in the deciduous species (Small, 1972, Gower and Richards, 1990, MacDonald and Lieffers, 1990), indicating a sizeable investment in metabolic enzymes such as Ribulose-1,5-bisphosphate
carboxylase/oxygenase (Rubisco), which would allow for increased photosynthetic capacity. Perhaps the most unique feature enabling tamarack to survive in the boreal forest is its high nitrogen recycling in senescing needles. Tamaracks can retract up to 25% more nitrogen from their needles before abscission than other boreal species, such as black spruce and paper birch, mitigating the high cost of the high leaf N and deciduous strategy (Chapin and Kedrowski 1983, Gower and Richards 1990, Givnish 2002).

1.4 Plant physiology

1.4.1 Photosynthesis

Photosynthesis is the process by which plants and other autotrophs create their own food – inorganic carbon dioxide (CO₂) is drawn from the air and fixed into sugars. This process is the foundation upon which all successive trophic layers are built; autotrophic carbon fixation is the only point in food webs where energy is stored in the bonds between carbon atoms in sugar molecules. Photosynthesis can be divided into two main parts: the light-dependent reactions of electron transport and the dark reactions of the Calvin-Benson cycle.

The light-dependent reactions begin in the thylakoid where light energy is harvested by pigment molecules such as chlorophyll. The pigment molecules absorb photons in the visible light spectrum; blue and red wavelengths are preferentially absorbed and yellow and green wavelengths are reflected, hence plants appear green in colour. The proteins of the light-dependent reactions are associated with the thylakoid membrane and begin with
Photosystem II (PSII) (Figure 1.1). As light strikes a leaf, pigment molecules, such as chlorophyll, in PSII harvest energy from photons and transfer it to a pair of electrons from a photo-dissociated water molecule. The electron transporter, plastoquinone, accepts the excited electrons from PSII as well as a pair of $H^+$ ions from the chloroplast stroma, both of which are transferred to the next major protein of the electron transport chain, cytochrome $b_{6}f$. Cytochrome $b_{6}f$ utilizes the energy from the excited electrons to pump the $H^+$ ions across the thylakoid membrane into the thylakoid lumen, creating a high $H^+$ concentration relative to the chloroplast stroma. The electrons in cytochrome $b_{6}f$ are returned to a low energy state after providing the energy to pump the $H^+$ ions, and are transferred to Photosystem I (PSI) via the transporter plastocyanin where the electrons are re-energized in the same manner as in PSII. The re-energized electrons are then passed on from PS I to ferredoxin, and lastly to ferredoxin NADP+ oxioreductase (FNR). The last protein in electron transport, FNR transfers the excited electrons to nicotinamide adenine dinucleotide phosphate (NADP$^+$) to create NADPH in the chloroplast stroma.
Figure 1.1 Schematic diagram of the enzymes involved in photosynthetic electron transport.

Electrons from a photo-dissociated water molecule are passed along a series of chloroplast membrane-associated enzymes: Photosystem II (1), plastoquinone (2), cytochrome b_{6}f (3), plastocyanin (4), Photosystem I (5), ferredoxin (6), and ferredoxin NADP+ oxioreductase (7). ATP synthase (8) generates adenosine triphosphate using a proton motive force. Red arrows represent the path of the excited electrons from a photo-dissociated water molecule and blue lines represent the points at which hydrogen ions cross the thylakoid membrane during photosynthetic electron transport. Black, dotted lines represent the path of light as it strikes the two Photosystems and solid, black, horizontal lines represent the thylakoid membrane.
NADPH is one of two major products of the light-dependent reactions and is used as a reductant in the subsequent dark reactions (the Calvin Cycle). The final protein in the electron transport chain (though itself not involved in the movement of electrons) is ATP synthase. The high H\(^+\) gradient in the thylakoid lumen is used to power the motor of ATP synthase, which binds a molecule of inorganic phosphate (P\(_i\)) to adenosine diphosphate to form adenosine triphosphate (ATP) in the chloroplast stroma. ATP is the other main product of the light-dependent reaction and is used to fuel the Calvin Cycle. As mentioned earlier, the electrons provided to the electron transport chain come from a photo-dissociated water molecule; after passing on the excited electrons, PS II becomes a strong oxidizing agent and breaks a water molecule apart into a pair of electrons and free oxygen gas (the latter being a by-product of photosynthesis).

ATP and NADPH produced in the light-dependent reactions are used in the Calvin cycle, which occurs in the chloroplast stroma. CO\(_2\) diffuses through stomata in the leaf surface, through the intercellular airspace of the leaf, into the mesophyll cells where photosynthesis occurs and into the chloroplast, where it is fixed by the enzyme Rubisco to the five-carbon molecule, Ribulose-1,5-bisphosphate (RuBP). The remaining steps in the Calvin-Benson cycle consist of a series of reduction reactions which consume ATP and NADPH from the light reactions to produce glyceraldehyde 3-phosphate (G3P – a three carbon molecule). G3P is used to make sugars, such as glucose, as well as lipids and polysaccharides, all of which are usable sources of energy for plant metabolic processes, and to regenerate RuBP. Three turns of the Calvin Cycle regenerate three
molecules of RuBP and produce one extra G3P that is used to make glucose, which itself is made of two G3P molecules.

While the enzyme Rubisco fixes CO\textsubscript{2} in photosynthesis, it is a dual function enzyme and the active site is able to fix O\textsubscript{2} in a process known as photorespiration, which emits CO\textsubscript{2} (Figure 1.2). In photorespiration, Rubisco fixes O\textsubscript{2} to RuBP to produce G3P (albeit in half the quantity as would normally be in a carboxylation reaction) as well as 2-phosphoglycolate (2PG). To convert 2PG into G3P costs a molecule of ATP and releases one molecule of CO\textsubscript{2}; therefore, photorespiration is both counterproductive to positive carbon assimilation in plants, and consumes ATP. Rubisco’s relative affinity to O\textsubscript{2} and CO\textsubscript{2} changes with temperature and will be elaborated on in section 1.5.1.1. While photorespiration reduces plant carbon gain, it has been shown to play a role in protecting plants from radiative stress (Wingler et al. 2000). Photosynthesis itself defines the process by which plants fix CO\textsubscript{2}; however, from a methodological standpoint, net CO\textsubscript{2} assimilation (A\textsubscript{net}) is the relevant process to measure. A\textsubscript{net} accounts for gross photosynthesis minus losses of CO\textsubscript{2} from photorespiration and mitochondrial respiration in the dark (R\textsubscript{dark}) or light (R\textsubscript{light}).
Rubisco fixes either CO$_2$ (photosynthesis) or O$_2$ (photorespiration). The process of photosynthesis involves fixing incoming CO$_2$ to ribulose-1, 5-bisphosphate (RuBP) resulting in the generation of glyceraldehyde 3-phosphate (G3P), and the production of glucose. In photorespiration, O$_2$ is fixed to RuBP to produce 2-phosphoglycolate (2PG); regenerating RuBP from 2PG does not result in sugar production, but emits CO$_2$ while still requiring the input of ATP and NADPH.
1.4.2 Respiration

The sugars produced in photosynthesis are the source of energy for the catabolic reactions of \( R_{\text{dark}} \). The disaccharide sucrose is the primary form of energy transported throughout the plant via the phloem to be later broken down in sink tissues. Whereas photosynthesis occurs in the chloroplasts, and mainly in leaves, respiration takes place in the mitochondria in all living plant tissue. Respiration occurs in four steps: glycolysis, pyruvate oxidation, the Krebs cycle (also called the citric acid cycle), and the electron transport chain. The stepwise combustion of a single glucose molecule through redox reactions results in the net production of at least 30 ATP molecules by the end of the respiratory process.

Glycolysis occurs in the cytoplasm and involves the halving of a glucose molecule into two three-carbon molecules of pyruvate; two NAD\(^+\) molecules are reduced to NADH in glycolysis and the electron acceptors and pyruvate are shuttled into the mitochondria. The ATP investment required to phosphorylate glucose can become a limiting factor in sustaining high rates of respiration (see section 1.5.2.2) (Atkin et al. 2000).

The processes of pyruvate oxidation and the Krebs cycle generate reducing agents in the forms of NADH and FADH\(_2\), as well as two molecules of ATP per glucose molecule, and produce \( \text{CO}_2 \) as a waste product. Pyruvate is oxidized to form acetyl-coenzyme A, which is attached to a pre-existing four-carbon molecule, oxaloacetate, in the Krebs cycle. As
with the regeneration of RuBP in the Calvin-Benson cycle, oxaloacetate is continually regenerated in the Krebs cycle so that the cycle is perpetuated.

The reducing agents produced in the Krebs cycle then power the electron transport chain (ETC) by donating their electrons to a series of proteins embedded in the inner mitochondrial membrane. The sequence of ETC proteins is as follows: Complex I (NADH dehydrogenase), Complex II (succinate dehydrogenase), Complex III (cytochrome bc₁ complex), and Complex IV (cytochrome c oxidase) – the final electron acceptor of the sequence is oxygen (O₂), which, when reduced, forms H₂O. The energy derived from electron flow across the chain is used to pump protons into the intermembrane space and create an electrochemical gradient across the inner mitochondrial membrane. The protons then move down the proton gradient through ATP synthase, converting large amounts of ADP to ATP through oxidative phosphorylation. This step is ultimately how the energy stored in glucose is converted into usable energy (in the form of ATP) for the plant.

Measurements of $A_{\text{net}}$ are derived from CO₂ taken up and O₂ emitted by the plant through photosynthesis, balanced with O₂ taken up and CO₂ emitted via respiration (and photorespiration, see above). In plants, respiration occurs both in the light and in darkness; however, from a methodological standpoint, it is difficult to measure $A_{\text{net}}$ and isolate the CO₂ emitted from $R_{\text{light}}$ from that emitted concurrently by photorespiration.
1.5 Climate change and plant metabolism

The increases in atmospheric CO\textsubscript{2} concentrations predicted for this century will have direct impacts on the ability of plants to take up CO\textsubscript{2} via photosynthesis. As well, because photosynthesis and respiration are both highly sensitive to temperature, it is crucial to examine how increases in global temperatures will influence plant carbon fluxes. Plant carbon balance is dictated by the amount of carbon taken in through photosynthesis minus carbon losses through respiration, photorespiration, root exudates, and emissions of volatile organic compounds. Climate-mediated changes in rates of photosynthesis and respiration will have substantial consequences for both individual leaf-level carbon balance and the overall productivity of plant populations and entire ecosystems.

Ecosystem productivity is the rate at which biomass is created in an ecosystem, and the foundation of biomass creation is laid by inorganic carbon being made available through autotrophic photosynthesis. It is well understood that autotrophic carbon balance defines ecosystem productivity, and therefore energy flow through higher trophic levels; however, it is less clear how autotrophic carbon balance, which is affected by global climate, then feeds back to influence global climate.

High temperature-induced reduction in A\textsubscript{net} by plants, either through decreased carbon uptake or by increased respiratory losses, can lead to a buildup of atmospheric CO\textsubscript{2}, leading to a positive feedback effect on further warming (Ainsworth and Rogers 2007, Slot and Kitajima 2015). Possible increases in A\textsubscript{net} resulting from enriched CO\textsubscript{2} in a
future world could offset warming-induced reductions in $A_{net}$ and reduce atmospheric CO$_2$ concentrations (Ainsworth and Long 2017). Earth system models, used to predict climate trajectories for the coming century, require an in-depth understanding of how carbon fluxes in different species and plant functional types vary in response to climate drivers (i.e. rising CO$_2$ and temperature) if these models are to make accurate forecasts.

1.5.1 Short-term plant responses to temperature

1.5.1.1 Photosynthesis

At low leaf temperatures, $A_{net}$ is low; as leaf temperature increases, $A_{net}$ increases until it reaches a thermal optimum ($T_{opt}$), then declines (Berry and Bjorkman, 1980) (Figure 1.3). The $T_{opt}$ of $A_{net}$ is highly variable among species and the nature of $A_{net}$ response to acute changes in temperature can differ among species. How $A_{net}$ responds to a short term (minutes to hours) increase in leaf temperature therefore depends on the degree of warming and how close the initial leaf temperature is to the plant’s $T_{opt}$ (Way and Yamori 2014, Yamori et al. 2014). A cold-limited plant—one operating at a temperature below its $T_{opt}$—will exhibit an increase in $A_{net}$ when exposed to warming. Conversely, if acute warming exceeds a plant’s $T_{opt}$, $A_{net}$ will decrease.
Figure 1.3 Thermal acclimation of net CO$_2$ assimilation rate ($A_{\text{net}}$) of a cold-grown plant and a warm-grown plant.

The warm-acclimated (dashed, red line) plant made constructive adjustments to the higher growth temperature by an increased thermal optimum of $A_{\text{net}}$ (vertical line descending from apex) and a rightward shift in the entire $A_{\text{net}}$ curve, indicating higher net CO$_2$ assimilation rates at higher temperatures that the cold-acclimated plant (solid, blue line). Detractive adjustment is seen in the warm-acclimated plant having a decreased $A_{\text{opt}}$ (maximal rate of $A_{\text{net}}$ at the $T_{\text{opt}}$; filled, red circle) compared to the cold-acclimated plant $A_{\text{opt}}$ (filled, blue circle). Adapted from Way and Yamori (2014).
The $T_{\text{opt}}$ of $A_{\text{net}}$ is determined by the balance between respiration and photorespiration relative to photosynthesis; as temperatures increase past the $T_{\text{opt}}$, respiratory and photorespiratory losses eclipse carbon gain through photosynthesis. At extremely high temperatures, the thermal optimum of photosynthesis is determined by the heat stability of the photosynthetic apparatus and integrity of plant cell membranes (Hüve et al. 2011). Warming increases the fluidity of the plasma membrane and increases proton leakiness across the thylakoid membrane, reducing the efficiency of NADPH and ATP production in the light-dependent reactions of photosynthesis (Hazel 1995, Sage and Kubien 2007). Additionally, at temperatures above the thermal optimum of electron transport, cyclic electron flow can be induced as a method of photoprotection, diminishing NADPH production (Sharkey and Schrader, 2006). Moderate warming (up to 45 °C) does not have direct detrimental effects on the light harvesting complexes (Tang et al. 2006), however, increasing leaf temperature can have detrimental effects on the ability of Rubisco to carboxylate RuBP. The enzyme kinetics of Rubisco are highly temperature-sensitive; while carboxylation rates increase exponentially with increasing temperature and Rubisco is stable at temperatures of up to 50 °C (Crafts-Brandner and Salvucci 2000), Rubisco’s chaperone enzyme, Rubisco activase, is heat-labile (Salvucci and Crafts-Brandner 2004, Sage et al. 2007, Holaday 2009). Rubisco activase maintains the activity of the active sites of Rubisco, but at high temperatures Rubisco activase cannot clear the catalytic sites of Rubisco fast enough to maintain Rubisco’s carboxylation activity (Crafts-Brandner and Salvucci 2000). Rubisco is highly conserved among plant species, but the thermal properties of Rubisco activase are species-specific (Holaday
beyond the optimal temperature for Rubisco activase function, denaturation of Rubisco activase can reduce the carboxylation capacity of the plant.

In addition to the temperature sensitivity of Rubisco, Rubisco substrate specificity also varies with temperature. Rubisco can fix either CO$_2$ or O$_2$ (photorespiration) but the relative affinity of Rubisco for O$_2$ over CO$_2$ increases with increasing temperature (Jordan and Ogren 1984). As well, the relative solubility of O$_2$ in air increases more rapidly than does the solubility of CO$_2$ (Ku and Edwards 1977, Jordan and Ogren 1984). The combined increase in photorespiration and decreased CO$_2$ solubility at high temperatures serve to exacerbate thermal stress in plants at supra-optimal temperatures.

Vapour pressure deficit (VPD) is the difference between the amount of moisture held in the air relative to the amount of moisture that could theoretically be held by saturated air. Vapour pressure deficit will increase with temperature if there is not an increase in relative humidity of the ambient air, resulting in an increase in leaf transpiration. Water loss through increased transpiration can cause a plant to become water-stressed; to combat water loss, plants will reduce their stomatal conductance, which, in turn, reduces CO$_2$ uptake and thereby photosynthetic carbon gain (Way et al. 2015).
1.5.1.2 Respiration

As demonstrated above, fluctuations in temperature and CO$_2$ influence plant carbon gain; however, approximately half of a plant’s carbon balance is dictated by carbon loss through respiration. Some plants can lose up to 80% of their daily carbon gain through $R_{dark}$; on average across plant species, half of the carbon gained through photosynthesis is respired (Amthor 1991, Atkin and Tjoelker 2003). In accounting for plant carbon economics in a future world, it is then imperative to understand how $R_{dark}$ responds to projected temperatures and CO$_2$ levels.

Typically, a linear increase in temperature results in an exponential increase in $R_{dark}$ (Atkin and Tjoelker 2003). Unlike the parabolic response of photosynthesis to temperature, $R_{dark}$ rates continue to increase at an exponential rate until the respiratory thermal optimum is reached. Increases in $R_{dark}$ with high temperatures are caused by increased kinetic rates of mitochondrial enzymes (Raison et al. 1971). Beyond the thermal optimum, $R_{dark}$ plummets as metabolic machinery succumbs to catastrophic heat stress: membranes are permanently damaged and enzymes denature (Hüve et al. 2011, Gauthier et al. 2014, O’Sullivan et al. 2016). It has been proposed that disintegration of mitochondrial membranes uncouple the processes of respiratory CO$_2$ release from electron transport at the high temperatures above the respiratory thermal optimum (i.e. > 50 °C) (Hüve et al. 2011). $R_{dark}$ thermal optima are above 50 °C, much higher than the typical 25 °C thermal optimum of $A_{net}$. At temperatures exceeding the $T_{opt}$ of $A_{net}$ (though not high enough to cause respiratory failure), $A_{net}$ declines while carbon loss continues to escalate.
1.5.2 Long-term plant response to temperature

1.5.2.1 Photosynthesis

If a plant remains in a warmed environment for several days to weeks, thermal acclimation can occur resulting in higher overall rates of $A_{\text{net}}$ at the new, higher temperature (Yamori et al. 2014). Photosynthetic thermal acclimation can be evidenced by a higher thermal optimum of photosynthesis, a higher rate of photosynthesis at that thermal optimum, and/or a higher temperature at which carbon uptake and respiratory losses are in equilibrium ($T_{\text{max}}$ – where $A_{\text{net}}$ equals zero) (Way and Yamori 2014). If the acclimation of these photosynthetic parameters increases carbon gain in a warmed plant under its new growth temperature, this is termed “constructive adjustment”; acclimation of these parameters that does not increase carbon gain is called “detractive adjustment” (*sensu* Way and Yamori 2014) (Figure 1.3).

One of the key mechanisms of photosynthetic thermal acclimation in some species is the production of a heat-stable isoform of Rubisco activase (Law et al. 2001, Scafaro et al. 2016). This allows for a continually high activation state of Rubisco and allows for a warm-acclimated plant to maintain higher rates of carboxylation than a cold-acclimated plant at high temperatures (Sage et al. 2007). Warm-acclimated plants may also express heat shock proteins and chaperone proteins to stabilize protein synthesis at supra-optimal temperatures and repair proteins damaged by high heat (Yamori et al. 2014). Heat shock and chaperone proteins can also confer stability to plant cell membranes (maintaining homeoviscosity) to reduce proton leakiness and therefore electron transport inefficiency (Yamori et al. 2014).
1.5.2.2 Respiration

Plants can acclimate $R_{\text{dark}}$ to warm temperatures over a period of several weeks to months such that plants acclimated to warm temperatures have lower rates of $R_{\text{dark}}$ than cold-acclimated counterparts when measured at a common temperature (typically assessed at 25 °C) (Atkin and Tjoelker 2003). Classically, respiratory acclimation can be characterized as either Type I or Type II (Figure 1.4). Type I acclimation appears as a reduction in the slope of the exponentially increasing curve of $R_{\text{dark}}$ with temperature in the warm-grown plant compared to a control plant (Atkin and Tjoelker 2003). The reduced slope of the $R_{\text{dark}}$ temperature curve indicates a decreased respiratory thermal sensitivity, often measured as a $Q_{10}$. $Q_{10}$ is the change in any physiological rate over a 10 °C increase in temperature; a $Q_{10}$ of 2 means that the rate doubles over the given increase in temperature and a $Q_{10}$ of 1 means that the rate has not changed in response to temperature (homeostasis).

Both Type I and Type II acclimation of respiration reduce the $Q_{10}$ of $R_{\text{dark}}$ in the warm-grown plant compared to the control plant. Type I acclimation yields similar rates of $R_{\text{dark}}$ at cold temperatures but lowers $R_{\text{dark}}$ rates at mid to high (10-35 °C) leaf temperatures (Atkin and Tjoelker 2003), and is thought to be due to substrate and adenylate limitations of $R_{\text{dark}}$ at higher leaf temperatures (Atkin and Tjoelker 2003, Lee 2005). Type I acclimation is typical in cold-acclimated plants which are then subject to a degree of
**Figure 1.4** Two types of respiratory acclimation of a cold-grown and warm-grown plant.

Type I acclimation shows similar respiratory output of the two plants at low measurement temperatures, followed by a decreased slope in warm-acclimated respiration (dashed, red line) compared to a cold-acclimated plant (solid, blue line). There is no difference in the y-intercept of the respiration curves between a Type I-acclimated plant and a cold-acclimated plant. Type II acclimation shows a reduced rate of respiration across the entire range of measurement temperature for the warm-acclimated plant compared to the cold-acclimated plant. Type II acclimation is characterized by a decreased slope *and* y-intercept of respiration in a warm-grown plant. Adapted from Atkin and Tjoelker (2003).
warming over several weeks (Atkin and Tjoelker 2003). In contrast, Type II acclimation of $R_{\text{dark}}$ not only decreases the $Q_{10}$ of $R_{\text{dark}}$, but also reduces the y-intercept of the $R_{\text{dark}}$ temperature response curve as well, such that rates of $R_{\text{dark}}$ are reduced at all leaf temperatures (Atkin and Tjoelker 2003). Type II acclimation results in a greater degree of respiratory suppression than Type I acclimation and occurs via physical changes in the respiratory machinery rather than solely by substrate and adenylate limitations (Atkin and Tjoelker 2003, Campbell et al. 2007); Type II-acclimated plants may show decreases in mitochondrial capacity and density, which may reduce the overall rate of respiratory carbon loss (Klikoff 1966, Miroslavov and Kravkina 1991, Atkin and Tjoelker 2003). Type II acclimation usually occurs in tissues which have developed in the new, warmer environment and confers a greater degree of respiratory homeostasis than does Type I acclimation (Atkin and Tjoelker 2003, Campbell et al. 2007).

### 1.5.3 Short-term plant response to CO$_2$

#### 1.5.3.1 Photosynthesis

Exposure to high concentrations of CO$_2$ directly stimulates photosynthesis by increasing the rate of Rubisco carboxylation and suppressing photorespiration (Ogren 1984). The carboxylation rate increases due to increased CO$_2$ substrate availability, which also effectively decreases photorespiration rates, since O$_2$ and CO$_2$ compete for the active site of Rubisco. At low CO$_2$ concentrations (i.e., <150 ppm intercellular CO$_2$), photosynthesis increases sharply as CO$_2$ concentrations increase, since the rate of Rubisco carboxylation is the limiting step in net CO$_2$ assimilation (Sage and Kubien 2007). At high CO$_2$ concentrations (i.e. > 500 ppm), the supply of ATP and NADPH from photosynthetic
electron transport becomes limiting to photosynthesis, and further increases in CO₂ do not stimulate net CO₂ assimilation rates as much.

1.5.3.2 Respiration

Plant Rₐₖₜ does not generally respond to changes in CO₂ concentration that occur over minutes to days. While it is methodologically complicated to measure Rₐₖₜ over a range of ambient CO₂ concentrations using a standard portable photosynthesis system, studies using a variety of techniques (including isotopic techniques) have established that there is no instantaneous stimulation or suppression of Rₐₖₜ by CO₂ (Amthor et al. 2001, Gonzalez-Meler et al. 2004).

1.5.4 Long-term plant response to CO₂

1.5.4.1 Photosynthesis

High CO₂-grown plants usually exhibit decreased Aₙₜₑᵗ when compared to ambient CO₂-grown plants measured at a common CO₂ concentration – a consequence of photosynthetic down-regulation (Sage 1994). The primary mechanism behind long-term photosynthetic CO₂ acclimation is a buildup of excess carbohydrates in leaves under high CO₂ conditions (Moore et al. 1999, Lemoine et al. 2013). As carbon is fixed by Rubisco in the leaves, the resulting sugars are transported to source tissues via the phloem; CO₂-enriched plants produce sugars at a rate that eventually exceeds sink tissue demands (Lemoine et al. 2013). Increased sugars in leaf cells can suppress the transcription of several Rubisco subunit proteins, thereby down-regulating photosynthesis (Moore et al.
The enzyme hexokinase phosphorylates the six-carbon sugars produced from the Calvin-Benson cycle. As sugar concentrations rise within the cells of an elevated CO$_2$-grown plant, so too do hexokinase concentrations; elevated hexokinase levels act as a signal in the reduction of Rubisco subunit transcription (Moore et al. 1999).

The photosynthetic apparatus is costly in terms of plant nitrogen investment – Rubisco accounts for up to 30% of a plant’s nitrogen and 50% of its soluble protein (Sage and Pearcy 1987, Feller et al. 2008). CO$_2$ acclimation reduces the amount of nitrogen devoted to photosynthetic enzymes, though this does not necessarily reduce $A_{\text{net}}$ in an elevated CO$_2$-acclimated plant. The reduced nitrogen investment in photosynthetic enzymes resulting in lowered photosynthetic capacity is offset by a direct CO$_2$ enrichment effect in an elevated CO$_2$-acclimated plant measured at its own growth CO$_2$ concentration (Crous et al. 2008, 2013). Rates of $A_{\text{net}}$ between ambient and elevated-CO$_2$ acclimated plants can be similar when measured in their respective growth CO$_2$ conditions – elevated CO$_2$ acclimated plants can even show higher rates of $A_{\text{net}}$ in such cases (Ellsworth et al. 2004).

Additionally, stomatal conductance ($g_s$) decreases in plants acclimated to high CO$_2$, as less stomatal opening is required to maintain relatively high intracellular CO$_2$ concentrations (Way et al. 2015). Transpiration (E) then decreases as plants reduce their $g_s$ over an acclimation period of several weeks, resulting in an increase in plant water use efficiency (WUE = $A_{\text{net}}$/E) (Ainsworth and Rogers 2007). High WUE can greatly benefit a plant as it mitigates water stress; however, decreased transpiration also reduces
evaporative cooling capabilities under high temperatures (Radin et al. 1994). Since high
temperatures will co-occur with increasing atmospheric CO₂ concentrations, reduced
evaporative cooling may exacerbate temperature stress for plants in a future world.
Additionally, reduced photosynthesis under CO₂-acclimated conditions, coupled with
reduced stomatal conductance, can significantly diminish plant carbon intake (Way et al.
2015).

1.5.4.2 Respiration
Acclimation of $R_{\text{dark}}$ to elevated CO₂ is less straightforward than the acute response of
$R_{\text{dark}}$ and results have varied across studies. Long-term growth in elevated CO₂ conditions
can lead to increased $R_{\text{dark}}$ due to higher concentrations of foliar non-structural
carbohydrates (i.e. the respiratory substrate) (Gonzalez-Meler et al. 2004). Increased $R_{\text{dark}}$
can be further underpinned by increased mitochondrial density in leaves in plants that
develop at high CO₂ concentrations (Griffin et al. 2001, Armstrong et al. 2006a).
However, other studies have shown negligible, or even negative, effects of long-term
CO₂ exposure on $R_{\text{dark}}$ rates; the lower nitrogen content of high CO₂-grown plants can
reduce respiratory capacity and could counteract any stimulation of $R_{\text{dark}}$ via increased
respiratory substrate availability (Gonzalez-Meler et al. 2004).

1.6 Rationale and Objectives
The boreal forest represents a region of paramount importance, economically and
ecologically, not only for Canada, but the entire planet. As the world’s largest terrestrial
biome, the boreal forest forms the foundation for significant biodiversity, as well as crucial emergent properties of its innumerable ecosystems, such as water and nutrient cycling. A significant proportion of the planet’s CO₂ flows through the plants and soils of the boreal forest, underscoring its pivotal role in a world increasingly imperiled by greenhouse gas accumulation. As the climate in northern latitudes is expected to warm at a higher rate than the rest of the world, it is important to understand how the projected changes in climate will affect the species that dominate the CO₂ fluxes of the boreal forest.

In my thesis, I investigated the physiological effects of elevated temperature and CO₂ on two key boreal species: black spruce and tamarack. Specifically, I sought to elucidate the capacity of these two species to acclimate CO₂ assimilation and respiration to increases in temperature and CO₂ concentration. I hypothesized that, given that these species are found in cold, harsh climates and are conservative compared to broad-leaved trees, they will exhibit limited acclimation of \( A_{\text{net}} \) and \( R_{\text{dark}} \) to increasing temperature and CO₂. Further, I hypothesized that tamarack, a deciduous species, will show a stronger acclimation response than will black spruce, as the evergreen leaf strategy of the former is consistent with limited, conservative response to environmental factors.

Future climate models require a high-resolution understanding of how species’ differential responses to climate drivers will feed back to influence the climate itself. The work of my thesis will serve to increase the understanding of how two dominant boreal
species will respond to elevated temperature and CO$_2$, and how they will, in turn, affect CO$_2$ fluxes that feed back onto future climates.
Chapter 2: Materials and Methods

2.1 Experimental design

Seeds of *Larix laricina* (tamarack, from Finch Township, Ontario [45.133 °N, 75.083 °W]) and *Picea mariana* (black spruce, from Seed Zone 27, Ontario [46.522 °N, 80.953 °W]) were obtained from the National Seed Tree Centre in February, 2016 and stored at -20 °C until planted. The parent trees are from provenances that experience climatic conditions similar to London, Ontario, near the southern edge of the natural distribution of both species.

Seedlings were grown from seed in the Biotron Centre for Experimental Climate Change at the University of Western Ontario in London, Ontario, Canada (43.009 °N, 81.274 °W). One week prior to planting seeds, potting medium was prepared using Promix General-Purpose Mycorrhizae (Premier Tech, Rivière-du-Loup, QC) mixed with Miracle-Gro slow release fertilizer (The Scotts Miracle-Gro Company, Marysville, OH, USA), at a ratio of approximately 240 mL of fertilizer per 107 L of potting medium. The potting mix was then moistened and divided into 11.3 L pots, to create a total of 588 pots for the experiment.

Six temperature and CO₂ treatments were used for the study, with each treatment imposed in a separate climate-controlled greenhouse at the University of Western Ontario’s Biotron Centre for Experimental Climate Change. The six treatments were: 1) ambient temperature (AT) with ambient CO₂ (400 µmol mol⁻¹ CO₂; AC); 2) AT with
elevated CO$_2$ (750 µmol mol$^{-1}$ CO$_2$; EC), 3) AT + 4 °C with AC, 4) AT + 4 °C with EC, 5) AT + 8 °C with AC, and 6) AT + 8 °C with EC. AT was determined by a five year (2006-2011) average of daily high and low temperature in London, Ontario and was ramped either up or down every 15 minutes between the two temperature points. Temperature control was maintained by an Argus Advanced Automated Control System (Argus Control Systems Ltd., Surrey, BC). The same system was also used to maintain CO$_2$ concentrations at 750 ppm in the EC greenhouses; AC air was pumped in from outdoors and then filtered. Relative humidity was maintained by the Argus system at a minimum of 60%. The project took place in glass houses therefore photoperiod was that of daily London, Ontario for the duration of the growing season; however, shades were closed over the plants from 10:00 to 14:00 hours daily. The shades served to protect the seedlings from direct, brilliant sunlight.

On 1$^{st}$ May, 2016, 10 seeds of either tamarack or black spruce were planted into each of 49 pots per species per treatment, with a thin layer of perlite on top to prevent seeds from desiccating and being displaced when watered. Seeds were germinated in their respective growth treatments and pots were watered daily to field capacity. After two months of growth (July 2016), seedlings were thinned to one per pot, selecting for seedlings growing farthest from the edge of the pot. Throughout the growing period, plants were rotated periodically within their treatment to mitigate any potential effects of variation in watering and irradiance.
2.2 Gas exchange measurements

Four months after planting (September 1st, 2016), gas exchange measurements were made on five random trees per species per treatment (n=5, N=60 seedlings for the experiment). Gas exchange measurements were conducted using a Li-Cor 6400 XT (Li-Cor BioSciences, Lincoln, NE, USA) which uses an infrared gas analyzer to measure changes in CO2 and water concentration in order to calculate numerous plant physiological parameters including rates of photosynthesis, respiration, and transpiration. The Li-Cor forms an open flow system whereby air passes through a chamber containing the leaf to be analyzed and concentrations of water vapour and CO2 are compared between ambient (reference) air before it goes through the cuvette and the air that is passed through the chamber. There is one infrared gas analyzer measuring reference air and another analyzer to measure the air after it passes over the plant sample; the difference in water vapour and CO2 concentrations are indicative of transpiration, and photosynthetic and respiratory activity of the plant. Measurements were made in a walk-in growth chamber (Environmental Growth Chambers, Chagrin Falls, OH, USA) to allow the seedlings and gas exchange system to reach a full range of measurement temperatures. Net CO2 assimilation rates (A\text{net}) and dark respiration rates (R\text{dark}) were measured at 10 °C, 15 °C, 20 °C, 25 °C, 30 °C, 35 °C, and 40 °C. At each temperature, light-saturated A\text{net} (measured at 1400 \mu mol photons m^{-2} s^{-1}) was first assessed at a cuvette CO2 concentration of 400 ppm, then at 750 ppm after A\text{net} stabilized. The cuvette CO2 was then returned to 400 ppm, the cuvette irradiance was set to 0 \mu mol photons m^{-2} s^{-1}, and R\text{dark} was measured after a 20 minute dark-acclimation period. Only one measurement CO2 (400 ppm) was used in the R\text{dark} measurements, as there is no direct
effect of measurement CO\textsubscript{2} on R\textsubscript{dark} (Amthor et al. 2001). Vapour pressure deficit was maintained between approximately 0.5 and 4.5 kPa across the full 30 °C range of measurement temperatures for all gas exchange measurements. This measurement procedure was repeated at each of the seven measurement temperatures, resulting in two A\textsubscript{net} temperature response curves (one at 400 ppm CO\textsubscript{2}, the other at 750 ppm) and one R\textsubscript{dark} temperature response curve for each tree.

After gas exchange was measured, needles from within the cuvette were stripped from the branch and photographed to analyze projected needle area (using Image J, US National Institutes of Health, Bestheda, MD, USA). The full gas exchange measurements of the 60 trees lasted from September 1\textsuperscript{st} to October 31\textsuperscript{st} 2016, and the treatments were repeatedly sampled in a rotating pattern to minimize any time effect on the variables.

2.3 Photosynthetic calculations
The photosynthetic thermal optimum (T\textsubscript{opt}, the temperature at which the A\textsubscript{net} is maximal) and the maximum A\textsubscript{net} (A\textsubscript{opt}, which occurs at T\textsubscript{opt}) were derived from each individual A\textsubscript{net} temperature response curve at each measurement CO\textsubscript{2} concentration by fitting a second-order polynomial to the data. This same equation was used to estimate T\textsubscript{max}, the CO\textsubscript{2} compensation point (where A\textsubscript{net} equals zero) above T\textsubscript{opt}, which represents the maximum temperature at which photosynthetic carbon gain offsets respiratory carbon loss.
The calculations for gas exchange parameters were made according to the LI-COR 6400 manual. Values for $A_{net}$ were estimated using equation (1)

$$A_{net} = \frac{F(C_r - C_s)}{100S} - C_s E$$  \hspace{1cm} (1)

where F denotes the air flow rate (µmol s$^{-1}$), $C_r$ and $C_s$ denote the reference and sample CO$_2$ concentrations, respectively, S denotes the leaf area (cm$^2$), and E denotes transpiration rate (mol m$^{-2}$ s$^{-1}$). Transpiration (E) was calculated using equation (2)

$$E = \frac{F(W_s - W_r)}{100S(1000 - W_s)}$$  \hspace{1cm} (2)

where $W_s$ and $W_r$ denote the sample and reference water mole fractions (mmol H$_2$O (mol air)$^{-1}$), respectively. Lastly, stomatal conductance was calculated from equation (3)

$$g_{tw} = \frac{E(1000 - \frac{W_l + W_s}{2})}{W_l - W_s}$$  \hspace{1cm} (3)

where $g_{tw}$ denotes total conductance (stomatal conductance plus boundary layer conductance) and $W_l$ denotes the water vapour concentration within the leaf (mmol H$_2$O (mol air)$^{-1}$).

### 2.4 Growth and nutrient analysis

Throughout the growing period of the experiment (May 1$^{st}$ to October 31$^{st}$, 2016), trees were repeatedly measured for shoot height and stem diameter to formulate growth curves. Shoot height was measured using a ruler, starting at the soil line. Stem diameter was measured using a Vernier caliper at the point on the stem where it met the soil line.
Needles used in gas exchange measurements were dried at 60 °C until constant mass and ground into powder using a Wiley mill (Thomas Scientific, Swedesboro, NJ, USA). Approximately 5 mg of each powdered sample was analyzed for percent carbon and nitrogen content using an elemental analyzer (Carlo Erba, Peypin, France). In elemental analysis, samples are combusted instantaneously and the resulting gases emitted are quantified by thermal conduction in a chromatographic column.

2.6 Statistical analysis

JMP software (SAS Institute, Cary, NC, USA) was used for all statistical analyses. Temperature response curves of $A_{\text{net}}$ at 400 ppm, $A_{\text{net}}$ at 750 ppm and $R_{\text{dark}}$ were assessed with repeated-measures ANOVAs within each species, considering measurement temperature, growth CO$_2$ and growth temperature. All other response variables were assessed within each species using two-way ANOVAs for growth CO$_2$ and growth temperature; a Tukey’s Honest Significant Difference post-hoc test was used when significant treatment effects were found.
Chapter 3: Results

3.1 Photosynthetic responses to growth temperature and CO₂

Moderate warming (T4) treatments were an average of 3.63 ± 0.090 °C warmer than ambient temperature (T0) treatments (Fig. 3.1A). Extreme warming (T8) treatments were an average of 3.81 ± 0.067 °C warmer than T4 treatments. Ambient CO₂ biomes maintained average CO₂ concentrations of 405.47 ± 1.96, 401.53 ± 2.75, and 414.32 ± 3.29 ppm CO₂ in the ACT8, ACT4, and ACT0 treatments, respectively (Figure 3.1B). Elevated CO₂ biomes maintained average CO₂ concentrations of 748.19 ± 0.31, 760.18 ± 1.58, and 743.02 ± 0.93 ppm CO₂ in the ECT8, ECT4, and ECT0 treatments, respectively.

3.1.1 Black spruce

3.1.1.1 Net CO₂ assimilation rate measured at a common CO₂ concentration of 400 ppm (A₄₀₀)

There was no significant effect of growth temperature on A₄₀₀ in spruce (p = 0.12); however, there was a trend for a growth CO₂ effect, as EC seedlings tended to have lower A₄₀₀ than AC seedlings (p = 0.086; Figure 3.2A and B, Table 1). There was a significant effect of measurement temperature on A₄₀₀ (p < 0.0001; Table 1); A₄₀₀ was initially stimulated as leaf temperature increased above 10 °C, then declined beyond the Tₜₜₒₜₜ., A significant interaction between growth temperature and growth CO₂ was found (p = 0.014), as EC seedlings had lower rates of A₄₀₀ than AC seedlings in all temperature treatments. There was also a significant interaction of measurement temperature and
Figure 3.1 Temperature and CO₂ levels within the six biomes over the duration of the tree-growing period prior to all physiological experiments.

Day 0 indicates the day of seed planting (1<sup>st</sup> May, 2016) and all temperature (A) and CO₂ (B) readings shown were taken daily at noon, over the growing period. Blue symbols represent ambient temperature growth conditions (T0), purple symbols represent +4 °C-warmed growth conditions (T4), and red symbols represent +8 °C-warmed growth conditions. (8T) Circles represent ambient CO₂ growth conditions of 400 ppm, squares represent elevated CO₂ growth conditions of 750 ppm CO₂.
Figure 3.2 Net CO₂ assimilation rates of black spruce and tamarack measured at 400 ppm CO₂ (A₄₀₀).

Measurements were taken between leaf temperatures of 10 °C and 40 °C and at a saturating light level of 1400 μmol photons m⁻² s⁻¹. Data shown as means ± SE, n = 5. Blue symbols and solid lines represent ambient temperature seedlings (T0), purple symbols and long-dashed lines represent +4 °C-warmed seedlings (T4), and red symbols and short-dashed lines represent +8 °C-warmed seedlings (8T). Circles represent ambient CO₂ growth conditions of 400 ppm (AC), squares represent elevated CO₂ growth conditions of 750 ppm CO₂ (EC).
growth CO\(_2\) (p = 0.0024) in spruce as the temperature response of A\(_{400}\) in EC trees was shifted to the right of the response curve of AC trees. A significant interaction was found between measurement temperature and growth temperature (p = 0.0037), as T0 seedlings tended to have lower A\(_{400}\) at cool leaf temperatures and higher A\(_{400}\) at high leaf temperatures than T4 and T8 seedlings. Lastly, there was a significant interaction between measurement temperature, growth CO\(_2\), and growth temperature (p = 0.0003; Table 1).

3.1.1.2 Net CO\(_2\) assimilation rate measured at a common CO\(_2\) concentration of 750 ppm (A\(_{750}\))

There was no significant effect of growth temperature (p = 0.21) on spruce A\(_{750}\) when measured at 750 ppm CO\(_2\), however, there was a trend for a growth CO\(_2\) effect (p = 0.093) as EC seedlings tended to have lower A\(_{750}\) than AC spruce (Figure 3.3A and B, Table 1). There was a significant effect of measurement temperature on A\(_{750}\) (p < 0.0001; Table 1); A\(_{750}\) was initially stimulated as leaf temperature increased above 10 °C, then declined beyond the T\(_{opt}\). A significant interaction between growth temperature and growth CO\(_2\) was found (p = 0.029) as ECT4 seedlings showed the lowest A\(_{750}\) of all three EC temperature treatments whereas ACT4 seedlings had middling A\(_{750}\) among the three temperature treatments. There was also a significant interaction between measurement temperature and growth CO\(_2\) (p = 0.0063) in spruce. A significant interaction was found between measurement temperature and growth temperature (p = 0.032) whereby AT seedlings tended to assimilate CO\(_2\) at higher rates than warmed seedlings across the
Table 1 Summary ANOVA statistics showing p-values for black spruce and tamarack gas exchange measurements of net CO₂ assimilation at 400 ppm measurement CO₂ ($A_{400}$), and 750 ppm measurement CO₂ ($A_{750}$).

Bolded p values are statistically significant ($p < 0.05$), italicized p values demonstrate $0.10 \geq p \geq 0.05$. $T =$ growth temperature treatment, $CO₂ =$ growth CO₂ concentration, and $T_m =$ measurement leaf temperature.

<table>
<thead>
<tr>
<th></th>
<th>Black spruce</th>
<th>Tamarack</th>
<th>DF</th>
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<td></td>
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<tr>
<td>CO₂</td>
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<td>$T_m \times CO₂ \times T$</td>
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<td>0.64</td>
<td>12</td>
</tr>
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</table>

|                |              |          |    |
| $A_{750}$      |              |          |    |
| CO₂            | 0.093        | 0.9959   | 1  |
| T              | 0.21         | 0.20     | 2  |
| $T_m$          | $<0.0001$    | $<0.0001$| 6  |
| $T \times CO₂$| 0.029        | 0.082    | 2  |
| $T_m \times CO₂$ | 0.0063     | 0.88     | 6  |
| $T_m \times T$ | 0.032        | 0.011    | 12 |
| $T_m \times CO₂ \times T$ | 0.052   | 0.21     | 12 |
Figure 3.3 Net CO₂ assimilation rates of black spruce and tamarack measured at 750 ppm CO₂ ($A_{750}$).

Measurements were taken between leaf temperatures of 10 °C and 40 °C and a saturating light level of 1400 μmol photons m$^{-2}$ s$^{-1}$. Data shown as means ± SE, n = 5. Blue symbols and solid lines represent ambient temperature seedlings (T0), purple symbols and long-dashed lines represent +4 °C-warmed seedlings (T4), and red symbols and short-dashed lines represent +8 °C-warmed seedlings (8T). Circles represent ambient CO₂ growth conditions of 400 ppm (AC), squares represent elevated CO₂ growth conditions of 750 ppm CO₂ (EC).
measurement temperature range. The interaction between measurement temperature, 
growth CO₂, and growth temperature was not significant but p = 0.052 (Table 1).

3.1.2 Tamarack

3.1.2.1 Net CO₂ assimilation rate measured at a common CO₂ concentration of 400 ppm (A₄₀₀)

There was no significant effect of growth temperature (p = 0.70) on tamarack A₄₀₀, nor 
was there a growth CO₂ effect (p = 0.59; Figure 3.2C and D, Table 1). There was a 
significant effect of measurement temperature on A₄₀₀ (p < 0.0001; Table 1); as in spruce, 
A₄₀₀ was initially stimulated as leaf temperature increased above 10 °C, then declined 
beyond the Tₜₜₜ. There was no significant interaction found between growth temperature 
and growth CO₂ (p = 0.14) nor between measurement temperature and growth CO₂ (p = 
0.95) in tamarack. There was also no significant interaction between measurement 
temperature and growth temperature (p = 0.15). Lastly, there was no interaction between 
measurement temperature, growth CO₂, and growth temperature (p = 0.64; Table 1).

3.1.2.2 Net CO₂ assimilation rate measured at a common CO₂ concentration of 750 ppm (A₇₅₀)

There was no significant effect of growth temperature (p = 0.20) or growth CO₂ (p=0.99) 
on tamarack A₇₅₀ when measured at 750 ppm CO₂ (Figure 3.3C and D, Table 1). There 
was a significant effect of measurement temperature on A₇₅₀ (p < 0.0001); A₇₅₀ was 
initially stimulated as leaf temperature increased, then declined beyond the Tₜₜₜ. There
was no interaction between growth temperature and growth CO₂ (p = 0.082) but ECT4 seedlings showed the lowest A₇₅₀ of all three EC temperature treatments while ACT4 seedlings showed the highest A₇₅₀ of all three AC temperature treatments. There was not, however, a significant interaction of measurement temperature and growth CO₂ (p = 0.88) in tamarack. There was a significant interaction between measurement temperature and growth temperature (p = 0.011), as the temperature response of A₇₅₀ in T₈ seedlings was shifted towards higher leaf temperatures in comparison to T₀ seedlings. Lastly, there was no interaction found between measurement temperature, growth CO₂, and growth temperature (p = 0.21; Table 1).

3.2 Stomatal conductance response to growth temperature and CO₂

3.2.1 Black spruce

There were no significant treatment effects of growth temperature (p = 0.31) or growth CO₂ conditions (p = 0.35) on spruce stomatal conductance (gₛ) when measured at 400 ppm CO₂ (gₛ₄₀₀), nor was there an interaction between the two (p = 0.97; Figure 3.4A and B, Table 2). Measurement temperature did not have a significant effect on gₛ₄₀₀ (p = 0.73) nor was there an interaction between measurement temperature and growth CO₂ (p = 0.13; Table 2). There was a significant interaction between measurement temperature and growth temperature (p = 0.010) as T₀ seedlings consistently maintained the highest gₛ₄₀₀ across the measurement temperatures while warm-grown seedlings maintained lower gₛ₄₀₀. Lastly, there was a significant interaction between measurement temperature, growth temperature, and growth CO₂: EC trees had reduced gₛ₄₀₀ in all temperature
treatments relative to AC trees, while ECT0 trees still maintained the highest $g_{s400}$ compared to warm-grown EC trees (Figure 3.4A and B).

There were no significant treatment effects or interactions when spruce $g_s$ was measured at 750 ppm CO$_2$ ($g_{s750}$; $p > 0.10$ for all; Figure 3.5A and B, Table 2) except for a significant interaction between measurement temperature, growth CO$_2$, and growth temperature ($p = 0.04$; Table 2); EC trees showed reduced $g_{s750}$ compared to AC trees and ECT0 trees had the highest $g_{s750}$ of all three EC temperature treatments (Figure 3.5A and B).

### 3.2.2 Tamarack

There were no significant treatment effects or interactions on tamarack $g_{s400}$ ($p > 0.10$ for all; Figure 3.4C and D, Table 2). There were also no significant treatment effects or interactions on tamarack $g_{s750}$ save for a trend of increasing $g_{s750}$ with increasing measurement temperature ($p = 0.09$; Figure 3.5C and D, Table 2).
**Figure 3.4** Stomatal conductance ($g_{s400}$) of black spruce and tamarack measured at 400 ppm CO$_2$.

Measurements were at a saturating light level of 1400 μmol photons m$^{-2}$ s$^{-1}$. Data shown as means ± SE, n = 5. Blue symbols and solid lines represent ambient temperature seedlings (T0), purple symbols and long-dashed lines represent +4 °C-warmed seedlings (T4), and red symbols and short-dashed lines represent +8 °C-warmed seedlings. (8T) Circles represent ambient CO$_2$ growth conditions of 400 ppm (AC), squares represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$ (EC).
Figure 3.5 Stomatal conductance ($g_{s750}$) of black spruce and tamarack measured at 750 ppm CO$_2$.

Measurements were taken at a saturating light level of 1400 $\mu$mol photons m$^{-2}$ s$^{-1}$. Data shown as means ± SE, n = 5. Blue symbols and solid lines represent ambient temperature seedlings (T0), purple symbols and long-dashed lines represent +4 °C-warmed seedlings (T4), and red symbols and short-dashed lines represent +8 °C-warmed seedlings (8T). Circles represent ambient CO$_2$ growth conditions of 400 ppm (AC), squares represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$ (EC).
Table 2 Summary ANOVA statistics showing p-values for black spruce and tamarack stomatal conductance measured at a CO₂ concentration of 400 (g₄₀₀) or 750 ppm (g₇₅₀), and the ratio of intercellular to atmospheric CO₂ at a measurement CO₂ of 400 ppm (Cᵢ/Cₐ₄₀₀) and 750 ppm (Cᵢ/Cₐ₇₅₀). Bolded p values are statistically significant (p < 0.05), italicized p values represent 0.10 ≥ p ≥ 0.05. T = growth temperature treatment, CO₂ = growth CO₂ concentration, and Tₘ = measurement leaf temperature.

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3.3 C\textsubscript{i}/C\textsubscript{a} response to temperature and CO\textsubscript{2}

3.3.1 Black spruce

There were no treatment effects or interactions on spruce intercellular CO\textsubscript{2} ratios when measured at 400 ppm CO\textsubscript{2} (C\textsubscript{i}/C\textsubscript{a400}; p > 0.21 for all; Figure 3.6A and B, Table 2); however, there was a strong measurement temperature effect (p < 0.0001) as C\textsubscript{i}/C\textsubscript{a400} decreased from 10 °C to 25 °C then increased again up to 40 °C (Figure 3.6A and B).

When spruce intercellular CO\textsubscript{2} ratios were measured at 750 ppm CO\textsubscript{2} (C\textsubscript{i}/C\textsubscript{a750}), the same measurement temperature effect occurred (p < 0.0001) as well as a trend for higher C\textsubscript{i}/C\textsubscript{a750} in T8 trees compared to T4 and T0 trees (p = 0.08; Figure 3.7A and B, Table 2).

3.3.2 Tamarack

There was a trend for a growth temperature effect (p = 0.090) on tamarack C\textsubscript{i}/C\textsubscript{a400} as increasing growth temperature led to increased C\textsubscript{i}/C\textsubscript{a400}, but no significant growth treatment effects were found (p > 0.20 for all; Figure 3.6C and D, Table 2). There was, however, a similar effect of measurement temperature on tamarack C\textsubscript{i}/C\textsubscript{a400} (p < 0.0001) as was seen in spruce. Tamarack C\textsubscript{i}/C\textsubscript{a750} responded in much the same way as did C\textsubscript{i}/C\textsubscript{a400}: there was a growth temperature trend (p = 0.070) on tamarack C\textsubscript{i}/C\textsubscript{a750} as increasing growth temperature led to increased C\textsubscript{i}/C\textsubscript{a750}, but no significant growth treatment effects (p > 0.24 for all; Figure 3.17 and D, Table 2). Again, there was a strong measurement temperature effect (p < 0.0001) as C\textsubscript{i}/C\textsubscript{a750} decreased from 10 °C to 25 °C then increased again up to 40 °C.
Figure 3.6 Ratio of intercellular CO$_2$ concentration to ambient CO$_2$ concentration ($C_i/C_{a400}$) of black spruce and tamarack measured at 400 ppm CO$_2$.

Measurements were taken at a saturating light level of 1400 µmol photons m$^{-2}$ s$^{-1}$. Data shown as means ± SE, n = 5. Blue symbols and solid lines represent ambient temperature seedlings (T0), purple symbols and long-dashed lines represent +4 °C-warmed seedlings (T4), and red symbols and short-dashed lines represent +8 °C-warmed seedlings (8T). Circles represent ambient CO$_2$ growth conditions of 400 ppm (AC), squares represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$ (EC).
Figure 3.7 Ratio of intercellular CO$_2$ concentration to ambient CO$_2$ concentration (C$_i$/C$_{a750}$) of black spruce and tamarack measured at 750 ppm CO$_2$.

Measurements were taken at a saturating light level of 1400 μmol photons m$^{-2}$ s$^{-1}$. Data shown as means ± SE, n = 5. Blue symbols and solid lines represent ambient temperature seedlings (T0), purple symbols and long-dashed lines represent +4 °C-warmed seedlings (T4), and red symbols and short-dashed lines represent +8 °C-warmed seedlings (8T). Circles represent ambient CO$_2$ growth conditions of 400 ppm (AC), squares represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$ (EC).
3.4 Photosynthetic thermal optima responses to growth temperature and CO2

3.4.1 Black spruce

The thermal optima of both A400 and A750 (T_{opt400} and T_{opt750}, respectively) increased significantly as growth temperature increased (p < 0.0025; Figures 3.8A and 3.9A; Table 3). ACT8-grown trees had a 13.3 % higher T_{opt400} and a 15.4 % higher T_{opt750} than ACT0 trees, while T_{opt} increased 11.3% for the same 8 °C increase in growth temperature in the EC seedlings in both measurement CO2 concentrations. At neither measurement CO2 did spruce show a significant T_{opt} response to growth CO2 conditions, nor was there a significant interaction between growth CO2 and temperature on T_{opt} (Table 3).

3.4.2 Tamarack

Tamarack T_{opt} (measured at both 400 and 750 ppm CO2) increased with increasing growth temperature (p < 0.021; Figures 3.8B and 3.9B; Table 3). ACT8 trees had an 18.9 % higher T_{opt400} and a 19.2% higher T_{opt750} than ACT0 trees; in the EC treatment, T_{opt400} increased by 15.9% in the ECT8 seedlings compared to the ECT0 trees and T_{opt750} increased by 20.7 % (Figures 3.8B and 3.9B). At neither measurement CO2 did tamarack show a significant T_{opt} response to growth CO2, nor was there a significant interaction between growth CO2 and growth temperature on T_{opt} (Table 3).
**Figure 3.8** Photosynthetic thermal optimum ($T_{\text{opt}400}$), maximum rate of net CO$_2$ assimilation ($A_{\text{opt}400}$), and high temperature CO$_2$ compensation point ($T_{\text{max}400}$) of black spruce and tamarack.

Measurements were taken at a saturating light level of 1400 µmol photons m$^{-2}$ s$^{-1}$ and a measurement CO$_2$ of 400 ppm. Data shown as means ± SE, n = 5. Blue bars represent ambient temperature seedlings, purple bars represent +4 °C-warmed seedlings, and red bars represent +8 °C-warmed seedlings. Solid bars represent ambient CO$_2$ growth conditions of 400 ppm (AC), hashed bars represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$ (EC). Different letters above bars denote a significant difference (p < 0.05).
Figure 3.9 Photosynthetic thermal optimum ($T_{\text{opt750}}$), maximum rate of net CO$_2$ assimilation ($A_{\text{opt750}}$), and high temperature CO$_2$ compensation point ($T_{\text{max750}}$) of black spruce and tamarack.

Measurements were taken at a saturating light level of 1400 μmol photons m$^{-2}$ s$^{-1}$ and a measurement CO$_2$ of 750 ppm. Data shown as means ± SE, n = 5. Blue bars represent ambient temperature seedlings, purple bars represent +4 °C-warmed seedlings, and red bars represent +8 °C-warmed seedlings. Solid bars represent ambient CO$_2$ growth conditions of 400 ppm (AC), hashed bars represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$ (EC). Different letters above bars denote a significant difference (p < 0.05).
Table 3 Summary ANOVA statistics table for black spruce and tamarack respiratory and photosynthetic characterizations of Q₁₀, photosynthetic thermal optimum measured at 400 ppm CO₂ (T_{opt400}), photosynthetic thermal optimum measured at 750 ppm CO₂ (T_{opt750}), maximal rate of net CO₂ assimilation measured at 400 ppm CO₂ (A_{opt400}), maximal rate of net CO₂ assimilation measured at 750 ppm CO₂ (A_{opt750}), maximal temperature of net CO₂ assimilation measured at 400 ppm CO₂ (T_{max400}), and maximal temperature of net CO₂ assimilation measured at 750 ppm CO₂ (T_{max750}).

Bolded p values are statistically significant (p < 0.05), italicized p values represent 0.10 ≥ p ≥ 0.05. T = growth temperature treatment, CO₂ = growth CO₂ concentration, and Tₘ = measurement leaf temperature.

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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CO₂</td>
<td>0.84</td>
<td>0.59</td>
<td>1</td>
</tr>
<tr>
<td>T</td>
<td><strong>0.0003</strong></td>
<td><strong>0.002</strong></td>
<td>2</td>
</tr>
<tr>
<td>T × CO₂</td>
<td>0.75</td>
<td>0.69</td>
<td>2</td>
</tr>
<tr>
<td>A_{opt400}</td>
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<td></td>
</tr>
<tr>
<td>CO₂</td>
<td>0.088</td>
<td>0.54</td>
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<tr>
<td>T</td>
<td>0.14</td>
<td>0.72</td>
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<tr>
<td>T × CO₂</td>
<td>0.81</td>
<td>0.10</td>
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<tr>
<td>A_{opt750}</td>
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<td></td>
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<tr>
<td>CO₂</td>
<td>0.14</td>
<td>0.93</td>
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<tr>
<td>T</td>
<td><strong>0.066</strong></td>
<td>0.29</td>
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<tr>
<td>T × CO₂</td>
<td>0.65</td>
<td><strong>0.076</strong></td>
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<td></td>
</tr>
<tr>
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<td>0.81</td>
<td>1</td>
</tr>
<tr>
<td>T</td>
<td>0.42</td>
<td>0.98</td>
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</tr>
<tr>
<td>T × CO₂</td>
<td><strong>0.091</strong></td>
<td>0.55</td>
<td>2</td>
</tr>
<tr>
<td>T_{max750}</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CO₂</td>
<td>0.44</td>
<td>0.80</td>
<td>1</td>
</tr>
<tr>
<td>T</td>
<td>0.50</td>
<td><strong>0.00094</strong></td>
<td>2</td>
</tr>
<tr>
<td>T × CO₂</td>
<td>0.42</td>
<td>0.46</td>
<td>2</td>
</tr>
</tbody>
</table>
Spruce A<sub>opt400</sub> did not respond to warming (p = 0.14; Figure 3.8C; Table 3). Although, spruce A<sub>opt400</sub> did tend to decrease at increased growth CO<sub>2</sub> (p = 0.088), with the average EC A<sub>opt</sub> (across all temperature treatments) being 26.5 % lower than that of all AC-grown trees. There was also a trend of A<sub>opt750</sub> decreasing with increasing growth temperature (p = 0.066; Figure 3.9C, Table 3). A<sub>opt750</sub> decreased by 27 % from T0 to T8-grown trees in both CO<sub>2</sub> treatments. There were no significant interactions between growth temperature and growth CO<sub>2</sub> on any spruce A<sub>opt</sub> (Table 3).

Tamarack A<sub>opt400</sub> did not show any response to growth CO<sub>2</sub> or growth temperature (Figure 3.8D, Table 3) – there were also no significant effects of growth temperature or CO<sub>2</sub> on tamarack A<sub>opt750</sub> (Figure 3.9D, Table 3). There was no interaction between growth CO<sub>2</sub> and growth temperature in A<sub>opt400</sub> in tamarack. However, there was a statistical trend for an interaction between growth temperature and growth CO<sub>2</sub> for A<sub>opt750</sub> in tamarack (p = 0.076): as ECT4 seedlings tended to have a lower A<sub>opt750</sub> than ACT4 seedlings (Figure 3.9D, Table 3). The A<sub>opt400</sub> of ECT4 tamaracks was 26.6 % lower than that of ECT0 trees and 33.2 % lower than ECT8 trees (Figure 3.9D). The A<sub>opt750</sub> of ECT4 tamaracks was 27.2 % lower than ECT0 trees and 25.1 % lower than trees grown in the ECT8 treatment (Figure 3.9D).

When measured at either 400 or 750 ppm CO<sub>2</sub>, T<sub>max</sub> did not respond to growth temperature or CO<sub>2</sub> in spruce (p > 0.39 for all; Figures 3.8E and 3.9E, Table 3). There was, however, a trend suggesting an interaction of growth temperature and CO<sub>2</sub> for
$T_{\text{max}400}$ ($p = 0.091$), as $T_{\text{max}400}$ in ECT8 seedlings was slightly higher than in ACT8 spruce. There was no significant interaction between growth CO$_2$ and temperature for spruce $T_{\text{max}750}$ (Table 3). $T_{\text{max}400}$ in tamarack did not respond to growth temperature, growth CO$_2$, nor was there an interaction between the two ($p > 0.55$ for all; Figure 3.8F, Table 3). The $T_{\text{max}750}$ increased with increasing growth temperature in tamarack ($p = 0.00094$), but there was no growth CO$_2$ effect ($p = 0.46$) nor an interaction between growth CO$_2$ and temperature ($p = 0.80$; Figure 3.9F, Table 3).

3.5 Respiratory response to growth temperature and CO$_2$

3.5.1 Black spruce

Elevated growth temperature conditions significantly decreased spruce $R_{\text{dark}}$ ($p = 0.01$) but growth CO$_2$ did not have an effect ($p = 0.85$) (Figure 3.10A and B, Table 4).

Measurement temperature significantly affected spruce $R_{\text{dark}}$ ($p < 0.0001$), as the expected stimulation of $R_{\text{dark}}$ was observed as measurement temperature increase. There was no interaction between growth temperature and growth CO$_2$ on spruce $R_{\text{dark}}$ ($p = 0.099$) nor between measurement temperature and growth CO$_2$ ($p = 0.17$, Table 4). There was a significant interaction between measurement temperature and growth temperature ($p = 0.038$; Table 4) as increasing growth temperatures resulted in lower rates of $R_{\text{dark}}$ across the measurement temperature range. There was also a trend of an interaction among measurement temperature, growth temperature, and growth CO$_2$ ($p = 0.059$), such that ECT4 and ECT8 trees showed slightly lower $R_{\text{dark}}$ than ACT4 and ACT0 trees, respectively (Figure 3.10A and B).
Figure 3.10 Dark respiration (R$_{\text{dark}}$) of black spruce and tamarack.

Measurements were taken between 10 °C and 40 °C at a light level of 0 μmol photons m$^{-2}$ s$^{-1}$ and a measurement CO$_2$ of 400 ppm. Data shown as means ± SE, n = 5. Blue symbols and solid lines represent ambient temperature seedlings (T0), purple symbols and long-dashed lines represent +4 °C-warmed seedlings (T4), and red symbols and short-dashed lines represent +8 °C-warmed seedlings (8T). Circles represent ambient CO$_2$ growth conditions of 400 ppm (AC), squares represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$ (EC).
Table 4 Summary ANOVA statistics table for black spruce and tamarack respiratory parameters of dark respiration ($R_{\text{dark}}$), thermal sensitivity of $R_{\text{dark}}$ ($Q_{10}$), dark respiration measured at a fixed temperature of 25 °C ($R_{\text{dark25}}$).

Bolded p values are statistically significant ($p < 0.05$), italicized p values represent $0.10 \geq p \geq 0.05$. T = growth temperature treatment, CO$_2$ = growth CO$_2$ concentration, and T$_m$ = measurement leaf temperature.

<table>
<thead>
<tr>
<th></th>
<th>Black spruce</th>
<th>Tamarack</th>
<th>DF</th>
</tr>
</thead>
<tbody>
<tr>
<td>$R_{\text{dark}}$</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>CO$_2$</td>
<td>0.85</td>
<td>0.68</td>
<td>1</td>
</tr>
<tr>
<td>T</td>
<td>0.01</td>
<td><strong>0.045</strong></td>
<td>2</td>
</tr>
<tr>
<td>T$_m$</td>
<td>&lt;0.0001</td>
<td><strong>0.0001</strong></td>
<td>6</td>
</tr>
<tr>
<td>T $\times$ CO$_2$</td>
<td>0.099</td>
<td>0.35</td>
<td>2</td>
</tr>
<tr>
<td>T$_m$ $\times$ CO$_2$</td>
<td>0.17</td>
<td>0.21</td>
<td>6</td>
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<tr>
<td>T$_m$ $\times$ T</td>
<td><strong>0.038</strong></td>
<td>0.19</td>
<td>12</td>
</tr>
<tr>
<td>T$_m$ $\times$ CO$_2$ $\times$ T</td>
<td><strong>0.059</strong></td>
<td>0.28</td>
<td>12</td>
</tr>
</tbody>
</table>

| $Q_{10}$ (10–40 °C) |              |          |    |
| CO$_2$            | 0.66         | 0.32     | 1  |
| T                | **0.018**    | 0.53     | 2  |
| T $\times$ CO$_2$ | 0.25         | 0.55     | 2  |

| $R_{\text{dark25}}$ |              |          |    |
| CO$_2$            | 0.62         | 0.96     | 1  |
| T                | **0.015**    | **0.012** | 2  |
| T $\times$ CO$_2$ | 0.95         | 0.42     | 2  |
3.5.2 Tamarack

Growth CO₂ conditions did not have a significant effect on tamarack R<sub>dark</sub> (p = 0.68); however, there was a significant effect of growth temperature (p = 0.045), such that needles grown at warmer temperatures had lower rates of R<sub>dark</sub> (Figure 3.10C and D, Table 4). Measurement temperature significantly affected tamarack R<sub>dark</sub> (p = 0.0001) as it did in spruce. There was no significant interaction between growth temperature and growth CO₂ on tamarack R<sub>dark</sub>, nor between measurement temperature and growth CO₂, measurement temperature and growth temperature, nor between measurement temperature, growth temperature, and growth CO₂ (p > 0.19 for all; Table 4).

3.5.3 Respiratory Q₁₀

When the Q₁₀ was calculated across the entire leaf measurement temperature range (10-40 °C), there was a significant effect of growth temperature in spruce (p = 0.018; Figure 3.11A, Table 4), as the Q₁₀ in spruce decreased with increasing growth temperatures. There was no significant growth CO₂ effect nor an interaction between growth temperature and CO₂ in the Q₁₀ from 10-40 °C in spruce. There were no significant effects of growth temperature, growth CO₂, nor growth treatment interactions on tamarack Q₁₀ calculated over the 10-40 °C range (p ≥ 0.32 for all; Figure 3.11B, Table 4).
**Figure 3.11** Respiratory $Q_{10}$ (over the 10-40 °C measurement range) and dark respiration rate at a fixed measurement temperature of 25 °C ($R_{\text{dark}25}$) of black spruce and tamarack. Measurements were taken at a light level of 0 $\mu$mol photons m$^{-2}$ s$^{-1}$ and a measurement CO$_2$ of 400 ppm. Data shown as means ± SE, $n = 5$. Blue bars represent ambient temperature seedlings, purple bars represent +4 °C-warmed seedlings, and red bars represent +8 °C-warmed seedlings. Solid bars represent ambient CO$_2$ growth conditions of 400 ppm (AC), hashed bars represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$ (EC). Different letters above bars denote a significant difference ($p < 0.05$).
3.5.4 $R_{\text{dark}}$ at 25 °C

When $R_{\text{dark}}$ was measured at a common leaf temperature of 25 °C ($R_{\text{dark}25}$), both black spruce ($p = 0.015$) and tamarack ($p = 0.012$) $R_{\text{dark}25}$ decreased with increasing growth temperature (Figure 3.11C and D, Table 4). There were no growth CO$_2$ effects on either species, nor any interactions (Table 4).

3.6 Response of tree growth to elevated temperature and CO$_2$

Elevated growth CO$_2$ increased black spruce height by 21.4 % compared to AC seedlings ($p < 0.0001$; Figure 3.12A, Table 4). There was also a trend for reduced shoot height in spruce with increasing temperature ($p = 0.069$). There was a significant temperature effect on tamarack height ($p = 0.0012$; Table 4); trees were tallest in the T4 treatment in both growth CO$_2$ concentrations (Figure 3.12B). There was a trend of a CO$_2$ enrichment effect increasing tamarack height ($p = 0.054$): ECT0 trees were 15.9 % taller than those grown in ACT0 conditions. There was no significant interaction between CO$_2$ and temperature on shoot height in either species (Table 4).

Growth temperature had a significant effect on spruce stem diameter: extreme warming significantly reduced stem diameter in the T8 treatment at both growth CO$_2$ concentrations compared to the T0 and T4 treatments ($p = 0.00074$; Figure 3.12C, Table 4). Stem diameter was 20.7% and 28.3 % smaller in T8 spruce than in T0 spruce in AC and EC-grown spruce, respectively. Elevated growth CO$_2$ increased black spruce stem diameter ($p = 0.00020$). There was also a trend for an interaction between growth CO$_2$
Figure 3.12 Shoot height and stem diameter of black spruce and tamarack.

Data shown as means ± SE, n = 5. Blue bars represent ambient temperature seedlings, purple bars represent +4 °C-warmed seedlings, and red bars represent +8 °C-warmed seedlings. Solid bars represent ambient CO₂ growth conditions of 400 ppm (AC), hashed bars represent elevated CO₂ growth conditions of 750 ppm CO₂ (EC). Different letters above bars denote a significant difference (p < 0.05).
Table 5 Summary ANOVA statistics table for black spruce and tamarack and anatomical parameters of shoot height, stem diameter, % N, and leaf mass per unit area (LMA).

Bolded p values are statistically significant (p < 0.05), italicized p values represent 0.10 ≥ p ≥ 0.05. T = growth temperature treatment, CO₂ = growth CO₂ concentration, and Tₘ = measurement leaf temperature.

<table>
<thead>
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<th>Shoot height</th>
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<th>Tamarack</th>
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<td>CO₂</td>
<td>&lt;0.0001</td>
<td>0.054</td>
<td>1</td>
</tr>
<tr>
<td>T</td>
<td>0.069</td>
<td>0.0012</td>
<td>2</td>
</tr>
<tr>
<td>T × CO₂</td>
<td>0.65</td>
<td>0.22</td>
<td>2</td>
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<table>
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<th>Tamarack</th>
<th>DF</th>
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</thead>
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<td>0.67</td>
<td>1</td>
</tr>
<tr>
<td>T</td>
<td>0.00074</td>
<td>0.00063</td>
<td>2</td>
</tr>
<tr>
<td>T × CO₂</td>
<td>0.088</td>
<td>0.64</td>
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</table>

<table>
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<th>Tamarack</th>
<th>DF</th>
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<td>0.29</td>
<td>1</td>
</tr>
<tr>
<td>T</td>
<td>0.0008</td>
<td>0.019</td>
<td>2</td>
</tr>
<tr>
<td>T × CO₂</td>
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<table>
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<th>DF</th>
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</thead>
<tbody>
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<td>CO₂</td>
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<td>1</td>
</tr>
<tr>
<td>T</td>
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<td>0.0088</td>
<td>2</td>
</tr>
<tr>
<td>T × CO₂</td>
<td>0.097</td>
<td>0.41</td>
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and T (p = 0.09), such that spruce seedlings had a larger stem diameter in the ECT0 treatment compared to the ACT0 trees.

Tamarack stem diameter showed a significant response to growth temperature (p = 0.00063), but not growth CO$_2$ (p = 0.67), with the lowest stem diameter occurring in 8T seedlings at both growth CO$_2$ conditions (Figure 3.12D, Table 4). There was no interaction between CO$_2$ and temperature on stem diameter in tamarack (Table 4).

3.7 Percent nitrogen and leaf mass per unit area (LMA)

Growth temperature significantly reduced leaf % N in spruce (p = 0.0008; Figure 3.13, Table 4), but growth CO$_2$ had no effect on spruce needle nitrogen content. There was a significant interaction between growth temperature and growth CO$_2$ on leaf % N in spruce (p = 0.002), such that T4 % N was the lowest of the three EC temperature treatments, but T8 was the lowest % N in the AC treatments. Tamarack responded to growth treatments in much the same way as spruce; increasing growth temperatures decreased leaf % N (p = 0.02; Figure 3.13, Table 4) and growth CO$_2$ did not have a significant effect on leaf % N (p = 0.29). There was a statistical trend of an interaction between growth temperature and growth CO$_2$ on leaf % N in tamarack, such that ECT4 % N was the lowest of the three EC temperature treatments, whereas in the AC treatment, % N decreased in order of increasing growth temperature (p = 0.07).
Figure 3.13 Needle percent nitrogen and carbon content of black spruce and tamarack.

Data shown as means ± SE, n = 5. Blue bars represent ambient temperature seedlings, purple bars represent +4 °C-warmed seedlings, and red bars represent +8 °C-warmed seedlings. Solid bars represent ambient CO$_2$ growth conditions of 400 ppm (AC), hashed bars represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$ (EC). Different letters above bars denote a significant difference (p < 0.05).
Black spruce LMA did not respond significantly to growth temperature (p = 0.24) or growth CO₂ (p = 0.46); however, there was a statistical trend for an interaction between growth temperature and CO₂ as LMA was highest in the T0 treatment at AC, but in the T4 treatment at EC (p = 0.097; Figure 3.14A, Table 4). Tamarack LMA decreased significantly with increasing growth temperature (p = 0.0089) but did not respond to growth CO₂ (p = 0.85); there was no significant interaction between growth temperature and growth CO₂ (p = 0.41; Figure 3.14B, Table 4).
Figure 3.14 Leaf mass per unit area (LMA) of black spruce and tamarack.

Data shown as means ± SE, n = 5. Blue bars represent ambient temperature seedlings, purple bars represent +4 °C-warmed seedlings, and red bars represent +8 °C-warmed seedlings. Empty bars represent ambient CO$_2$ growth conditions of 400 ppm, hashed bars represent elevated CO$_2$ growth conditions of 750 ppm CO$_2$. 
Chapter 4: Discussion

4.1 The future of black spruce and tamarack in the boreal forest

Black spruce acclimated net photosynthesis to warming with primarily detractive adjustments, demonstrating that the physiology of the seedlings used in this experiment was best suited to current, ambient conditions and higher temperatures impair net carbon gain. Spruce was effective at acclimating respiratory losses to increased growing temperatures; however, shoot height and stem diameter decreased in the warmer treatments, suggesting that aboveground carbon sequestration will be weakened in future warming scenarios despite acclimating $R_{\text{dark}}$. While the detrimental effects of moderate warming on spruce stem diameter were fully mitigated by enriched growth CO$_2$, extreme warming still caused a significant reduction in stem diameter regardless of growth CO$_2$ treatment.

Tamarack, on the other hand, did not acclimate $A_{\text{net}}$ to increasing temperatures; instead, net carbon gain remained constant across temperature treatments. However, when measured at 750 ppm CO$_2$, extreme warming decreased rates of $A_{\text{net}}$. Tamarack seedlings acclimated $R_{\text{dark}}$ to a lesser extent than spruce; however, increased aboveground biomass allocation with moderate warming may have required higher rates of growth respiration. Tamarack height and stem diameter increased with 4 degree warming and declined considerably with 8 degree warming, suggesting that seedlings may benefit from projected mid-century climates where spruce, ostensibly, will not.
Declining foliar nitrogen concentration underpinned the acclimation of $A_{\text{net}}$ and $R_{\text{dark}}$ in black spruce and tamarack. Photosynthetic enzymes and pigments are the largest nitrogen pools in leaves (Sage and Pearcy, 1987) and low leaf N usually translates into low photosynthetic capacity, such that the biochemical demand for CO$_2$ is reduced. As the seedlings of this experiment were well fertilized, changes in leaf nitrogen were not likely due to poor supply. But if both species had reduced leaf N and reduced photosynthetic capacity (which is corroborated by unpublished data from these same trees), how could tamarack maintain high rates of $A_{\text{net}}$? The answer likely lies in the ability to supply CO$_2$ to the site of carboxylation. Whereas spruce stomatal conductance ($g_s$) was lowest in warm-grown trees, tamarack appeared to mitigate the effects of low foliar nitrogen in warm-grown trees by increasing $g_s$, which would increase CO$_2$ substrate availability to the Calvin Cycle, thereby boosting $A_{\text{net}}$. The fundamental differences in acclimation of carbon dynamics between the two species became manifest in their accumulation of aboveground biomass, where spruce aboveground biomass responded negatively in all warming scenarios whereas tamarack increased height and diameter with 4 degree warming.

While the reality of field experiments is quite different than the highly-controlled laboratory setting of my experiment, the physiological limitations of spruce $A_{\text{net}}$ in response to warming suggest that future climates will impair the ability of this species to compete in the boreal forest. Tamarack may become a more dominant species in the
future, as the predicted temperature and CO\textsubscript{2} conditions of coming decades exceed the ability of black spruce to acclimate carbon gain.

4.2 Thermal acclimation of A\textsubscript{net} in black spruce and tamarack

4.2.1 Black spruce

Under no circumstances did spruce A\textsubscript{net} improve at high measurement temperatures when grown in warm conditions. Thermal acclimation of A\textsubscript{net} occurred; however, all adjustments were detractive. Moreover, rates of A\textsubscript{net} in warm-grown trees were consistently lower at low measurement temperatures, indicating that T4 and T8 warming regimes reduced net photosynthetic carbon gain in spruce relative to trees grown in current thermal conditions. The limited acclimation potential of black spruce therefore appears to be insufficient for the species to maintain photosynthetic homeostasis at moderate and extreme warming scenarios, let alone improve net photosynthetic efficiency relative to seedlings grown in current, ambient conditions. Previous studies with black spruce have found similar limitations of A\textsubscript{net} in high temperature-grown trees, also attributed to lower leaf nitrogen concentrations (Way and Sage 2008a, 2008b).

Black spruce T\textsubscript{opt} increased with growing temperature in both ambient and elevated CO\textsubscript{2}-grown seedlings at both measurement CO\textsubscript{2} concentrations, indicating a partial degree of thermal acclimation (Way and Sage 2008a, Way and Yamori 2014). However, spruce A\textsubscript{opt} (the maximum rate of carbon assimilation at T\textsubscript{opt}) was lower in warm-grown treatments, meaning that warm-grown trees still had lower carbon gain relative to cool-
grown trees. As well, the $T_{\text{max}}$ of $A_{\text{net}}$ (where photosynthetic carbon gain and respiratory carbon loss are in equilibrium) should shift to a higher temperature in a warm-acclimated plant, such that carbon gain at higher measurement temperatures is improved relative to a cool-acclimated plant (Yamori et al. 2014), but warm-grown black spruce seedlings did not have increased $T_{\text{max}}$. Taken together, the data indicate little plasticity in spruce for $A_{\text{net}}$ to acclimate to increasing growth temperatures.

4.2.2 Tamarack

Tamarack, on the other hand, showed no significant acclimation of $A_{400}$ to temperature, indicating that net carbon gain was not sensitive to increases in growth temperature. This is consistent with previous research on thermal acclimation of $A_{\text{net}}$ in tamarack seedlings (Tjoelker et al. 1998). At $A_{750}$, there was evidence of constructive adjustment of $A_{\text{net}}$, as $T_{\text{max}}$ increased with moderate warming, although T8 trees had lower $A_{\text{net}}$ compared to T0 trees. $A_{\text{opt}}$ also tended to be lower in T8 than T0 tamarack seedlings, which would suggest that extreme warming impaired $A_{\text{net}}$, but a larger sample size may be required to demonstrate a statistical significance for this effect. The degree of thermal acclimation of carbon assimilation was most clear in the $A_{750}$ curves, where the higher $A_{\text{net}}$ rates allow for greater separation of the data. ACT4 seedlings showed a classic thermal acclimation response as rates of $A_{750}$ were slightly higher than ACT0 at higher measurement temperatures. ACT8 seedlings had lower rates of $A_{750}$ compared to ACT0 seedlings at all but the highest measurement temperatures, at which point they were commensurate with the $A_{750}$ rates of ACT0. These results suggest that tamarack responses to moderate (T4) warming may lead to improvements in $A_{\text{net}}$, or at the very least, moderate warming does
not reduce $A_{\text{net}}$, while extreme warming (T8) of tamarack seedlings results in diminished net photosynthesis. As with black spruce, tamarack $T_{\text{opt}}$ increased with increasing growth temperature and the $A_{\text{opt}750}$ remained constant among all temperature treatments, indicating a constructive adjustment in net carbon assimilation.

Interestingly, $A_{\text{net}}$ was lower in ECT4 spruce and tamarack then in either ECT0 or ECT8. This “anomaly” is, however, underpinned by lower leaf nitrogen concentration ECT4 leaves of both species. Investment in photosynthetic apparatus is directly influenced by leaf nitrogen concentration (Warren et al. 2003, Way and Sage 2008a, Scafaro et al. 2016) so the low $A_{\text{net}}$ in the ECT4 treatment is likely attributable to the low needle nitrogen concentration.

### 4.3 CO$_2$ acclimation of $A_{\text{net}}$ in black spruce and tamarack

In black spruce, $A_{\text{net}}$ showed significant acclimation to CO$_2$: $A_{\text{net}}$ was lower in EC-grown trees at all growth temperatures compared to AC-grown trees, when measured at a common CO$_2$ concentration (either $A_{400}$ or $A_{750}$). Photosynthetic down-regulation has been found before in black spruce seedlings grown at high CO$_2$ and was found to be a result of decreased photosynthetic capacity (Tjoelker et al. 1998). When $A_{\text{net}}$ was assessed at growth CO$_2$ conditions ($A_{\text{growth}}$), net carbon assimilation was higher in EC-grown trees, likely as the instantaneous effect of high measurement CO$_2$ allowed for high levels of substrate availability for CO$_2$ fixation in EC trees (Crous et al. 2008). Growth
CO$_2$ did not have any effect on spruce $T_{opt}$; however, there was a trend of decreased $A_{opt400}$ in EC-grown spruce seedlings, supporting the finding of CO$_2$ acclimation of $A_{net}$.

Unlike black spruce, tamarack did not acclimate to elevated growth CO$_2$ conditions, and EC-grown trees did not show a down-regulation of $A_{net}$ compared to AC-grown trees. This is contrary to previous results which have found acclimation of $A_{net}$ in elevated CO$_2$-grown tamarack (Tjoelker et al. 1998). This difference could be attributed to the small pot size used by Tjoelker et al. (1998) which would limit seedling growth and therefore carbohydrate sink demand. The only exception to this was the unusual suppression of $A_{net}$ in ECT4 trees at both $A_{400}$ and $A_{750}$ mentioned earlier. There were no effects of growth CO$_2$ on $T_{opt}$ or $A_{opt}$ (save for a decreased $A_{opt750}$ in the ECT4 treatment as discussed previously). The lack of CO$_2$ acclimation in tamarack may be indicative of an absence of carbohydrate sink limitations in high CO$_2$-grown seedlings conditions (Moore et al. 1999, Ainsworth and Rogers 2007, Lemoine et al. 2013), which may be linked to the higher growth of tamarack seedlings in EC (Coleman et al. 2008), demonstrating a notable difference between black spruce and tamarack response to elevated growth CO$_2$.

4.4 Stomatal conductance and intercellular CO$_2$ ratio response to elevated temperature and CO$_2$

Stomatal conductance ($g_s$) was consistently highest in $T0$ spruce in both ambient and elevated growth CO$_2$ conditions and at both measurement CO$_2$ concentrations. Measurement CO$_2$ did not influence spruce $g_s$, suggesting that stomatal conductance was not responsive to acute changes in ambient CO$_2$. This result is consistent with previous
research characterizing conifer species as having a conservative, insensitive stomatal response to changes in ambient CO₂ compared to broad-leaved species (Tjoelker et al. 1999a, Medlyn et al. 2001). At both measurement CO₂ concentrations, gₛ was suppressed in EC-grown spruce, indicating acclimation of gₛ to high-CO₂ growing conditions. Acclimation of gₛ to elevated CO₂ has been found across several tree species (reviewed in Medlyn et al. 2001). Suppressed gₛ in EC-grown spruces is consistent with the suppressed Aₙₑₙ curves of the same trees, potentially implying a causal effect of stomatal limitations on Aₙₑₙ reduction seen in EC trees.

Tamarack gₛ was not significantly influenced by growth temperature or growth CO₂ concentration; however, an apparent stratification of gₛ by growth temperatures is visible, suggesting increased gₛ with increasing growth temperatures. A larger sample size may have been able to detect a growth temperature effect on gₛ if it exists; however, higher gₛ in warm-grown trees would certainly explain the constructive adjustments of Aₙₑₙ seen in tamarack.

The ratio of intracellular CO₂ (Cᵢ) to ambient CO₂ (Cₐ) represents the balance of CO₂ diffusing into the leaf and CO being fixed by Rubisco. If a plant’s CO₂ intake is not commensurate with its rate of CO₂ fixation, the former becomes a constraint on Aₙₑₙ (Farquhar et al. 1980). None of the growth treatments induced significant changes in Cᵢ/Cₐ for spruce, suggesting that reduced gₛ limiting CO₂ supply to photosynthesis was balanced by a reduced biochemical ability to fix CO₂. Tamarack Cᵢ/Cₐ, however, showed
an increasing trend matching the stratification of $g_s$ whereby $C_i/C_a$ increased with increasing growth temperature. This suggests that diminishing biochemical demand for CO$_2$ in photosynthesis was mitigated in tamarack by increasing substrate supply.

4.5 Acclimation of R$_{dark}$ to elevated temperature and CO$_2$

Black spruce showed strong thermal acclimation of R$_{dark}$; rates of R$_{dark}$ decreased with increasing growth temperature. While average air temperature in the Canadian boreal could warm by up to 8 °C by 2100 (IPCC, 2014), leaf temperatures have been shown to exceed air temperature by more than 6 °C (Martin et al. 1999). Therefore, acclimation of R$_{dark}$ at high measurement temperatures is central to maintaining a positive carbon balance. The significant reduction in spruce R$_{dark25}$ with increasing growth temperature obviates a reduced slope of the R/T curve and is consequently strong evidence of Type I respiratory acclimation (Atkin and Tjoelker 2003).

Like spruce, tamarack seedlings acclimated R$_{dark}$ to temperature such that warm-grown trees maintained lower rates of R$_{dark}$ than cool-grown trees. While spruce showed a discernible stratification of decreasing R$_{dark}$ with growth temperature, such a pattern only appeared in AC-grown tamaracks. ECT4 tamarack R$_{dark}$ was as low as the ECT8, again perhaps underpinned by the low foliar nitrogen concentration (as explained previously). While R$_{dark25}$ decreased significantly with increasing growth temperature, Q$_{10}$ did not respond to growth temperature, again indicating Type I acclimation of R$_{dark}$ in tamarack. Other studies have shown that conifers are able to successfully acclimate respiration to
increasing growth temperature, even to a greater extent than broad-leaved species
however growth temperature and CO$_2$ concentrations were maintained at lower levels
than this project (Tjoelker et al. 1999a, Tjoelker et al. 1999b, reviewed in Way and Oren
2010).

Neither species showed a response of $R_{\text{dark}}$ to elevated growth CO$_2$, nor effects on $R_{\text{dark}25}$
or Q$_{10}$. This suggests that foliar mitochondrial structure and density did not respond to
enriched growth CO$_2$, and changes in photosynthetic carbohydrate production did not
translate into altered rates of $R_{\text{dark}}$ (Griffin et al. 2001, Armstrong et al. 2006b). Previous
studies on trees have found a similar lack of growth CO$_2$ effect on respiratory

4.6 Effects of elevated temperature and CO$_2$ on foliar nitrogen
Both species showed a reduction in foliar nitrogen concentration with increasing growth
temperature. The acclimation of $R_{\text{dark}}$ to high temperature likely resulted from a lower
energy requirement to sustain the fewer enzymes caused by lower leaf nitrogen (Tjoelker
et al. 1999b). Declining foliar nitrogen concentration likely contributed to the reduction
in spruce $A_{\text{net}}$ in T4 and T8 warming treatments; in tamarack, the negative effects of low
nitrogen on $A_{\text{net}}$ appear to have been counteracted by the acclimation of $g_s$, facilitating
higher rates of CO$_2$ diffusion into the needles.
There was no overall significant growth CO₂ effect on spruce leaf nitrogen concentration, implying that high CO₂ acclimation of $A_{\text{net}}$ in spruce was not underpinned by reductions in foliar nitrogen and was rather likely a function of decreased $g_s$ in EC-grown trees as well as a regulatory carbon-sink feedback. The uniquely low nitrogen concentration of the ECT4 spruces (compared to other EC temperature treatments) may have been low enough (i.e., below a required nitrogen threshold) to significantly reduce $A_{\text{net}}$, whereas the overall lack of growth CO₂ effect on leaf nitrogen indicates that nitrogen concentration alone did not lead to acclimation of $A_{\text{net}}$ to high CO₂. There was no effect of growth CO₂ concentration on tamarack leaf nitrogen concentration; however, like spruce, there was a significant reduction in leaf nitrogen of ECT4 tamaracks. As was the case with spruce, the low nitrogen concentration of ECT4 trees may have caused reduced tamarack $A_{\text{net}}$ in the ECT4 treatment.

### 4.7 Effects of elevated temperature and CO₂ on tree growth

Black spruce seedlings showed a decreasing trend in height with increasing growth temperature. Previous work with black spruce has similarly found that high temperature-grown seedlings show impaired growth, both above- and belowground (Way and Sage 2008b). There was, however, a strong positive effect of elevated growth CO₂ in spruce height, such that ECT8 trees were as tall as ACT0 trees; this suggests that predicted rises in CO₂ by the end of the century may be sufficient to offset the detrimental effects of elevated temperature on spruce height. Tamarack shoot height increased with moderate warming, while extreme warming caused a slight decline in height. An increase in shoot height with moderate warming, coupled with the constructive adjustments seen in $A_{\text{net}}$ of
ACT4 tamarack, suggests that tamarack seedling growth will benefit with a slight increase in global temperature. However, previous work that found reduced allocation to root mass in high temperature-grown conifers (a parameter not measured in my thesis) suggests that improvements in aboveground growth with warming may be counteracted by impaired ability to access belowground resources (Way and Sage 2008b), which could increase drought susceptibility.

With extreme warming of 8 °C, black spruce stem diameter decreased relative to the T0 and T4 treatments. Overall, spruce stem diameter benefited from enriched growth CO₂ concentrations as EC spruce seedlings had a greater stem diameter than AC trees. The positive effect of elevated growth CO₂ was not sufficient to counteract the negative effect of extreme warming on spruce stem diameter. In both CO₂ treatments, tamarack stem diameter increased with moderate warming, but decreased with extreme warming. Diminishing stem diameter under extreme warming treatments suggests less carbon allocation to wood in the seedlings of both species and is consistent with decreased rates of Aₙₑₙ in warm-grown trees. Lignin formation in wood is a major carbon sink for trees and forms up to 20 % of the carbon sink in needle-leaved species (Thompson and Randerson 1996); lignin is also a recalcitrant carbon compound which decomposes slowly in forest soils, making it an effective form of long-term carbon sequestration (De Deyn et al. 2008). Whereas tamarack showed an increase in stem diameter of T4 seedlings, spruce did not – this suggests that tamarack seedlings may have an improved capability to survive in a future, warmer boreal forest compared to black spruce. Previous
dendroclimatological work has shown that field-grown black spruce radial growth was negatively affected by increasing temperatures (Huang et al. 2010).

Black spruce LMA decreased with increasing growth temperature, with the exception of ECT4 seedlings, which had a higher LMA than ECT0 trees. This result is surprising given that $A_{\text{net}}$ in ECT4 spruce seedlings was lower than ECT0 seedlings; one would therefore expect less allocation of carbon to needle biomass. However, this was not the case. Perhaps allocation to needles mass came at the expense of allocation elsewhere, such as root mass. There was no effect of growth CO$_2$ concentration on spruce LMA. Tamarack LMA also decreased with increasing growth temperature and, like spruce, did not respond to growth CO$_2$. Decreasing LMA suggests thinner needles in both species which would facilitate increased CO$_2$ diffusion into the chloroplasts in warm-grown trees (Hultine and Marshall 2000).

4.8 Feedback on future climates

Black spruce and tamarack are dominant species of the boreal forest and the results of my experiment suggest that net carbon fluxes will be reduced in both species in climates predicted for the end of the 21$^{\text{st}}$ century. Tamarack may experience a slight increase in growth with moderate warming; however, black spruce $A_{\text{net}}$ acclimated detractively in all warming scenarios, and growth was suppressed in spruce, even though $R_{\text{dark}}$ was reduced by high growth temperatures. Together this suggests that these two species will not be as effective carbon sinks in the future as they are now. Reduced photosynthetic CO$_2$ uptake
by two major species in the boreal forest, caused by increased temperatures, could cause a significant accumulation of atmospheric CO$_2$, perpetuating further climate warming into the 22nd century.

4.9 Future directions

The greatest limitation in the design of my experiment was the sample size of five seedlings for all the gas exchange analyses. Ideally, this experiment could be replicated with at least one more Li-Cor 6400 XT in order to double the sample size. Additionally, increasing the time scope of the experiment would be useful; all the seedlings in this experiment were less than six months old. By measuring seedling carbon flux across seasons, over several years, ontogenetic information about acclimation capacity could be gained. Characterizing carbon flux of other boreal species will also be imperative in creating high-resolution climate models, therefore I propose a similar experimental design with other dominant boreal species such as paper birch (*Betula papyrifera*) and trembling aspen (*Populus tremuloides*).

In understanding acclimation capacity, it is imperative to characterize photosynthetic capacity; maximum rates of Rubisco carboxylation and rates of photosynthetic electron transport are crucial parameters to quantify across plant functional types and traits in order to create accurate climate models. While these parameters were not part of my own work, they formed a substantial part of this overall project. Future studies must study these parameters with all plant types. As the species composition changes in a warming
world, understanding soil dynamics will aid in predicting which types of plant species will succeed, and the consequences on higher trophic levels that follow.
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