

Electronic Thesis and Dissertation Repository

6-20-2017 12:00 AM

Physiological Resolution of Periodic Breath Holding During Heavy Intensity Fartlek Exercise

David J. Lim, *The University of Western Ontario*

Supervisor: Dr. Glen Belfry, *The University of Western Ontario*

A thesis submitted in partial fulfillment of the requirements for the Master of Science degree in Kinesiology

© David J. Lim 2017

Follow this and additional works at: <https://ir.lib.uwo.ca/etd>



Part of the [Exercise Physiology Commons](#)

Recommended Citation

Lim, David J., "Physiological Resolution of Periodic Breath Holding During Heavy Intensity Fartlek Exercise" (2017). *Electronic Thesis and Dissertation Repository*. 4600.
<https://ir.lib.uwo.ca/etd/4600>

This Dissertation/Thesis is brought to you for free and open access by Scholarship@Western. It has been accepted for inclusion in Electronic Thesis and Dissertation Repository by an authorized administrator of Scholarship@Western. For more information, please contact wlsadmin@uwo.ca.

Abstract

The purpose was to compare the physiological responses to periodic 5 s breath holds (CBH), increased power output (FLK), and both (FBH) every 30 s followed by 25 s of continuous heavy intensity cycling exercise with free-breathing (CON). Minute ventilation (\dot{V}_E) was greater ($p < 0.05$) in CBH ($97.58 \pm 16 \text{ L}\cdot\text{min}^{-1}$) and FBH ($100.9 \pm 19 \text{ L}\cdot\text{min}^{-1}$), pulmonary oxygen uptake ($\dot{V}O_{2p}$) was similar in CBH ($2.73 \pm 0.14 \text{ L}\cdot\text{min}^{-1}$) and FBH ($2.73 \pm 0.14 \text{ L}\cdot\text{min}^{-1}$) and greater in FLK ($2.85 \pm 0.12 \text{ L}\cdot\text{min}^{-1}$), compared to CON ($2.71 \pm 0.12 \text{ L}\cdot\text{min}^{-1}$). FBH also resulted in slower $\dot{V}O_{2p}$ kinetics ($62.2 \pm 19 \text{ s}$) and greater blood lactate concentrations ($11.5 \pm 2.7 \text{ mM}$), compared to CON ($48.8 \pm 12 \text{ s}$; $9.0 \pm 2.3 \text{ mM}$). Together, we demonstrated that breath hold-induced hypoxemia and hypercapnia were resolved when not combined with additional work.

Keywords: breath-by-breath pulmonary O_2 uptake, gas exchange, breath hold, hypoxia, muscle deoxygenation, intermittent exercise, heavy intensity

Co-Authorship Statement

This study was designed by G. R. Belfry and D. J. Lim with input from the advisory committee (J. M. Kowalchuk and G. D. Marsh). Majority of the data was collected and analyzed by D. J. Lim with the assistance of J. Kim. D. J. Lim wrote the original manuscript for the study.

Acknowledgments

First and foremost, I would like to extend my gratitude towards my supervisor, Glen R. Belfry for the opportunity to experience and expound my research in integrative exercise physiology at the Master's level. Words cannot contain my gratitude for your encouragement and patience in my development as a researcher and scientific writer. As a role-model and mentor beyond research, the impact you have made in my life will never be forgotten.

I would also like to thank Dr. Greg D. Marsh, for the invaluable insights on the manuscript, and acknowledge my colleagues, Michael Hodgson, Lorenzo Love, André Pelletier, James Vanhie, and Jae Joon Kim for their camaraderie in the lab, the motivation to give my best, and the perspective to enjoy the ride.

Lastly, I would like to express my gratitude to my parents and Grace for their inspiration and unconditional support as I navigate through my convoluted academic journey, and to Esther – you are the love of my life, my best friend, and the light that shines through the murky grime.

I dedicate my thesis to my grandmother, who encouraged my academic endeavors far greater than I could imagine.

Table of Contents

Abstract	ii
Co-Authorship Statement	iii
Acknowledgments	iv
Table of Contents	v
List of Tables	vii
List of Figures	viii
List of Abbreviations	xviii
Chapter 1	1
1 REVIEW OF LITERATURE	1
1.1 Introduction	1
1.2 Energy Systems and Lactate Threshold	2
1.3 Oxygen Uptake Kinetics	6
1.4 Intermittent Exercise	7
1.5 Hypoxia and Breath Holding During Exercise	9
1.6 Breath-by-Breath Analysis by Mass Spectroscopy	11
1.7 Near – Infrared Spectroscopy and Muscle Deoxygenation	12
1.8 Study Rationale	14
1.9 References	16
Chapter 2	24
2 PHYSIOLOGICAL RESOLUTION OF PERIODIC BREATH HOLDING DURING HEAVY INTENSITY FARTLEK EXERCISE	24
2.1 Introduction	24
2.2 Methods	26
2.3 Results	32
2.4 Discussion	49

2.5 Future Directions and Limitations	58
2.6 References	59
Appendix: Ethics Approval Notice.....	68
Curriculum Vitae	69

List of Tables

Table 1. Participant characteristics and performance variables from the ramp incremental test including age, height, body mass, $\dot{V}O_{2\text{peak}}$, estimated LT, peak PO, and PO at $\Delta 50$	36
Table 2. Summary of physiological parameters collected during CON, CBH, FLK, and FBH	37
Table 3. Summary of fluctuations in $\dot{V}O_{2p}$, $\dot{V}CO_{2p}$, \dot{V}_E , $P_{\text{ET}}O_2$, $P_{\text{ET}}CO_2$, $\Delta[\text{HHb}]$, and $\Delta[\text{HHb}]/\dot{V}O_{2p}$ over the 25 s and 5 s intervals within the 30 s cycles of CON, CBH, FLK, and FBH.....	38

List of Figures

Figure 1. Mean pulmonary oxygen uptake ($\dot{V}O_{2p}$) profile during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2peak}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The mean $\dot{V}O_{2p}$ from 120 s to the end of exercise was greater in FLK than all other conditions ($p < 0.05$).....39

Figure 2a. Mean end-tidal partial pressure of oxygen ($P_{ET}O_2$) during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2peak}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The $P_{ET}O_2$ from 120 s to the end of exercise was greater in FLK than CON, greater in FBH than CON, and lower in FBH than FLK ($p < 0.05$).....40

Figure 2b. Fluctuations in mean end-tidal partial pressure of oxygen ($P_{ET}O_2$) during CON and CBH. This reflects the acute resolution of transient hypoxia associated with each breath hold episode. The $P_{ET}O_2$ during the last 5 s of were lower than the peak values during the 25 s periods over the 30 s cycles ($p < 0.05$)..... 41

Figure 3a. Mean end-tidal partial pressure of carbon dioxide ($P_{ET}CO_2$) during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2peak}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The $P_{ET}CO_2$ from 120 s to the end of exercise was lower in FLK than CON, and greater in FBH than FLK ($p < 0.05$).....42

Figure 3b. Fluctuations in mean end-tidal partial pressure of carbon dioxide ($P_{ET}CO_2$) during CON and CBH. This reflects the acute resolution of transient hypercapnia associated with each breath hold episode. The $P_{ET}CO_2$ during the last 5 s were greater than the nadir values during the 25 s periods over the 30 s cycles ($p < 0.05$).....43

Figure 4. Mean minute ventilation (\dot{V}_E) during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2peak}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The \dot{V}_E from 120 s to the end of exercise was greater in FLK and CBH than CON ($p < 0.05$)..... 44

Figure 5a. Mean total hemoglobin concentration changes ($\Delta[Hb_{tot}]$) from baseline values during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2peak}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The mean $\Delta[Hb_{tot}]$ from 120 s to the end of exercise during CBH and FLK were lower than CON and FBH ($p < 0.05$)..... 45

Figure 5b. Mean deoxygenated hemoglobin concentration changes ($\Delta[HHb]$) from baseline values during CON, CBH, FLK, and FBH. The mean $\Delta[HHb]$ from 120 s to the end of exercise were different in all conditions ($p < 0.05$).....46

Figure 5c. Mean tissue hemoglobin saturation ($S_{at}O_2$) during CON, CBH, FLK, and FBH. The mean $S_{at}O_2$ from 120 s to the end of exercise during CBH and FBH were greater than CON ($p < 0.05$).....47

Figure 6. Mean adjustment of normalized [HHb]-to- $\dot{V}O_{2p}$ ratio ($\Delta[HHb]/\dot{V}O_{2p}$) during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2peak}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free-breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free-breathing sprints - peak power output attained during ramp incremental test), and FBH (repeated 25 s free-breathing $\Delta 50$ and 5 s sprints with breath holds). $\Delta[HHb]/\dot{V}O_{2p}$ during the last 5 s were greater than the nadir values during the 25 s periods over the 30 s cycles ($p < 0.05$)..... 48

List of Abbreviations

ADP – Adenosine diphosphate

ATP – Adenosine triphosphate

BH – Breath hold

BPM – Beats per minute

CO₂ – Carbon dioxide

CON – Continuous exercise

CBH – Continuous breath hold

FBH – Fartlek breath hold

FLK – Fartlek

H⁺ – Hydrogen ion

[Hb_{tot}] – Total hemoglobin concentration

[HHb] – Deoxygenated hemoglobin concentration

[HHb]/ $\dot{V}O_{2p}$ – Adjustment of normalized [HHb]-to- $\dot{V}O_{2p}$ ratio

HR – Heart rate

INT – Intermittent exercise

La⁻ – Lactate

LT – Estimated lactate threshold

m – Meter

Mb - Myoglobin

min – Minute

MOD – Moderate intensity

ms – Millisecond

N_2 – Nitrogen

NIRS – Near-infrared spectroscopy

O_2 – Oxygen

[O_2Hb] – Oxygenated hemoglobin concentration

PCr – Phosphocreatine

PCO_2 – Partial pressure of carbon dioxide

$P_{ET}CO_2$ – End-tidal partial pressure of carbon dioxide

$P_{ET}O_2$ – End-tidal partial pressure of oxygen

pKa – Acid dissociation constant

PO – Power output

PO_2 – Partial pressure of oxygen

QO_2 – Blood flow

RAMP – Incremental ramp test

RPM – Revolutions per minute

s – Second

SD – Standard deviation

SE – Standard error

$S_{at}O_2$ – Tissue hemoglobin saturation

τ – Time constant

$\dot{V}CO_{2p}$ – Pulmonary carbon dioxide production

\dot{V}_E – Minute ventilation

$\dot{V}O_{2m}$ – Muscle oxygen uptake

$\dot{V}O_{2p}$ – Pulmonary oxygen uptake

$\dot{V}O_{2\text{peak}}$ – Peak pulmonary oxygen uptake

W – Watts

WR – Work rate

μ_A – Absorption coefficient

μ_S – Scattering coefficient

Chapter 1

1 REVIEW OF LITERATURE

1.1 Introduction

Intermittent exercise patterns are inherent to a variety of sports such as cycling (59), rowing (72), and swimming (16). During training and competition, these intermittent exercise patterns may be comprised of brief periods of higher power output. In the sport of swimming, these periods of higher power output may be performed during the underwater kicking phase after a turn in a 50 m pool, to maintain or increase speed (45, 80). As such, a breath hold is required. Olympic swimming rules (18) dictate that the swimmer may stay under water after a turn for a distance no greater than 15 m.

A swimmer may elect to perform underwater kicking for the total 15 m distance at various intensities. They will cover that 15 m distance in approximately 5 s. Upon reaching the surface of the water, the swimmers continue to kick for approximately 25 s (73) as a complementary propulsive component to the upper limbs for the remainder of the 50 m distance.

Thus, during swimming training and/or competitions requiring multiple lengths of a 50 m pool, numerous cardiovascular, respiratory and metabolic challenges associated with the regular and intermittent changes in power output, and breath hold-induced reductions in O₂ delivery to the tissues (hypoxia) must be resolved. The present study examined the physiological responses associated with the regular insertions of 5 s breath holds and/or periods of higher power output

(sprints) after a 25 s period performed at a work rate that is above the lactate threshold during leg only exercise, on a cycle ergometer.

This chapter reviews the background literature of breath holding and intermittent exercise including energy systems, pulmonary gas exchange, oxygen uptake kinetics, lactate threshold and buffering, and muscle deoxygenation to underpin the rationale for this study. Thereafter, an overview of the equipment used for the cardiorespiratory and metabolic data collection, and their relevance to this research study is presented.

1.2 Energy Systems and Lactate Threshold

Energy Systems

The current study utilized six minute (min) bouts of high intensity constant load exercise. Therefore, the energy systems responsible for providing adenosine triphosphate (ATP) for this intensity and duration will be addressed first. At the onset of exercise, there is an instantaneous increase in ATP requirement at the exercising muscles that matches the prescribed work rate. However, the energy contribution from aerobic metabolism is delayed for a brief period until cardiac output increases and due to the “sluggishness” of the metabolic pathways (91). The phosphorylation necessary during the first ~60 s after the onset of exercise is primarily provided by the adenosine triphosphate-phosphocreatine (ATP-PCr) and the glycolytic systems. Its magnitude and duration determines the ‘O₂ deficit’ (47). Figure 1 below illustrates the interplay between both the PCr-derived, glycolytic and oxidative phosphorylation contributions to 5 min of constant-load exercise (20). Thus, a similar energy system interplay is expected during the 6 min constant-load exercise in the present study.

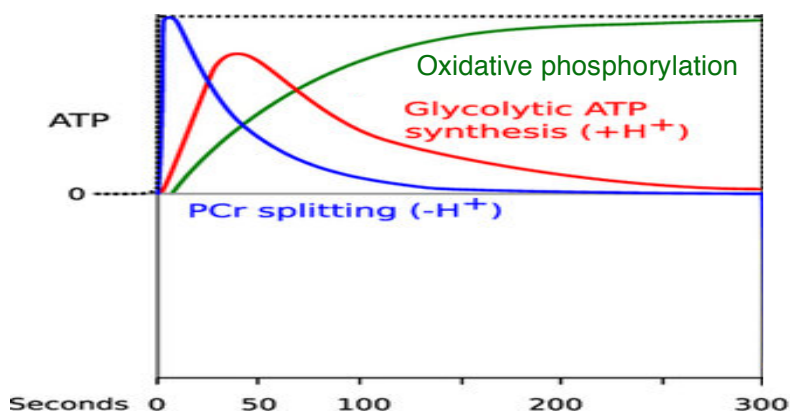


Figure 1. Energy system contribution during a five-minute constant-load exercise bout. The first (blue) line represents the ATP-PCr system, the second (red) line represents anaerobic glycolysis, and the third (green) line represents oxidative phosphorylation (modified from Fielder et al., 2016).

The final stage of the oxidative phosphorylation pathway is the electron transport chain. It is here where the movement of H^+ generates a proton gradient within the inner mitochondrial membrane that facilitates adenosine diphosphate (ADP) phosphorylation in the presence of ATP synthase. The final reaction of the electron transport chain involves the coupling of a pair of H^+ to $\frac{1}{2}$ an O_2 . Oxygen utilization or uptake at the mouth is dictated by this final step (47). Thus, increasing the ATP demand and related O_2 utilization results in decreased partial pressure of O_2 (PO_2) at the muscle. This stimulates offloading of the O_2 from the myoglobin (Mb) in the muscle and subsequently O_2 from the hemoglobin (Hb) in the microvasculature. This in turn results in an increase in deoxygenated Hb and a decrease in the PO_2 (26). This drop in PO_2 also results in increased vasodilation that enhances blood flow, and thus O_2 delivery (55). As this blood reaches the lungs, an elevated alveolar-capillary PO_2 gradient results (83). Under steady-state $\dot{V}O_{2p}$

conditions, deoxygenated hemoglobin concentrations ($[\text{HHb}]$) have been utilized to provide insight to changes in O_2 delivery (55).

Lactate Threshold

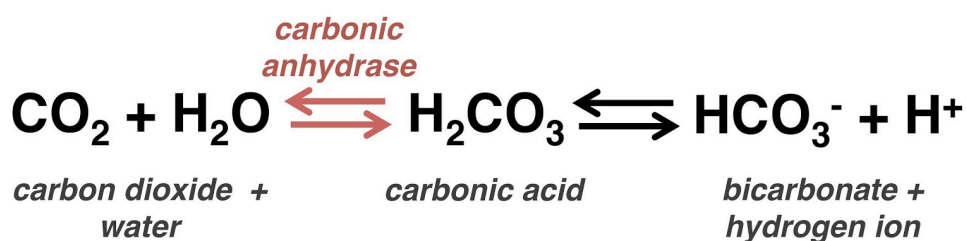
High intensity exercise during training and competition is associated with increased lactate production. Within the context of the high intensity exercise performed in the present study, a greater rate of pyruvate production than its oxidation in Krebs's cycle results in pyruvate accumulation. Initially, the build-up of pyruvate is minimized as lactate dehydrogenase reduces pyruvate into lactic acid (47). Lactic acid, due to its high pK_a , immediately dissociates into lactate (La^-), a strong anion, and equimolar (1.5) amounts of hydrogen ions ($[\text{H}^+]$). High concentrations of La^- and H^+ in the cytosol are co-transported across the sarcolemma via monocarboxylate transporters to the blood (27). Assuming bicarbonate ions are available, H^+ is buffered by the carbonic anhydrase reaction, producing CO_2 (23). This leads to increased partial pressures of CO_2 in the blood (hypercapnia). As peripheral chemoreceptors are most sensitive to changes in PCO_2 , this increase in non-metabolic CO_2 will stimulate the peripheral chemoreceptors (88) to induce hyperventilation for the removal of CO_2 from the blood. If H^+ production continues to increase beyond the maximal capacity of bicarbonate-ventilatory buffering, termed as the lactate threshold, H^+ will accumulate and result in decreased pH.

As exercise intensity increases within work rates below the lactate threshold, the increase in oxidative phosphorylation is matched by an identical increase in respiratory CO_2 production ($\dot{V}\text{CO}_{2p}$) originating from the pyruvate to acetyl-CoA reaction and the Krebs's cycle. Under steady state conditions, $\dot{V}\text{O}_{2p}$ stabilizes after approximately 3 min and $\dot{V}\text{CO}_{2p}$ after 4 min (11).

Under prolonged submaximal exercise intensities, oxidative phosphorylation is responsible primarily for meeting the ATP requirement. However, as mentioned, ATP supply from aerobic metabolism does not increase instantaneously to meet the energy demand at exercise onset (91). During this upregulation period of oxidative phosphorylation ATP is produced primarily via the breakdown of intramuscular PCr stores and substrate level phosphorylation (Wilmore et al., 2008). Moreover, at intensities below the lactate threshold (moderate intensity exercise), this transient mismatch between the rate of ATP hydrolysis required to fuel mitochondrial metabolism and the actual rate of ATP supply, also known as the ‘O₂ deficit’, is overcome and a steady state $\dot{V}O_{2p}$ is achieved (11).

In the present study, exercise will be performed at supra-lactate threshold intensities for 6 min. Thus, increased [La⁻] is expected upon completion of each exercise bout. During exercise at intensities above the lactate threshold, the buffering and subsequent accumulation of H⁺ via the carbonic anhydrase reaction (Equation 1) results in increased in non-respiratory $\dot{V}CO_{2p}$ resulting in the accumulation, and subsequent buffering of H⁺.

Equation 1. Interconversion of carbon dioxide and water to bicarbonate and protons.



This is linked with substrate level phosphorylation and ATP hydrolysis, which is associated with increases in chemoreceptor output from increased $\dot{V}CO_{2p}$, that results in an

increase in ventilation (61). Moreover, work rates above the lactate threshold are also associated with the evolution of the $\dot{V}O_{2p}$ “slow component” that delays the onset of steady state $\dot{V}O_{2p}$ (91). The $\dot{V}O_{2p}$ slow component will be described in depth later in this review.

1.3 Oxygen Uptake Kinetics

At exercise intensities below the lactate threshold (moderate intensity), the rate of adjustment of $\dot{V}O_{2p}$ exhibits a mono-exponential response to match ATP demand during exercise. Three distinct $\dot{V}O_{2p}$ kinetic phases are observed (Whipp et al., 1982). Phase I is the ‘cardio-dynamic’ phase, comprised of a temporal delay (~ 20 s) between the deoxygenated blood arriving at the lungs, prior to the increase in cardiac output. Phase II or the “fundamental” phase is the larger mono-exponential that projects towards steady-state, reflective of both the increased rate of ATP utilization and O_2 delivery (89). At work rates below the lactate threshold Phase III is the point at which $\dot{V}O_{2p}$ reaches a steady-state at which the ATP requirement is met entirely by oxidative phosphorylation.

At exercise intensities above the lactate threshold (heavy intensity domain), similar to the work rates employed in the present study, Phase III exhibits as a second exponential called the “slow component” and the onset of steady state $\dot{V}O_{2p}$ is delayed, and if the work rate is high enough, may not be attained (91). This phenomena during supra-lactate threshold constant-load exercise has been used to describe the increase in $\dot{V}O_2$ /work rate relationship, or gain (35).

The kinetic response of the phase II $\dot{V}O_{2p}$ is described by the time constant tau (τ). This reflects the time it takes $\dot{V}O_{2p}$ to reach 63% of the difference from baseline $\dot{V}O_{2p}$ to steady-state $\dot{V}O_{2p}$. In healthy young adults, the $\tau\dot{V}O_{2p}$ for exercise below the lactate threshold (moderate

intensity) has shown to range from 20 – 35 s (3, 14), and even longer for exercise above the lactate threshold (heavy intensity) (17).

Earlier research has also observed a slowing of $\dot{V}O_{2p}$ kinetics from moderate to severe intensity (upper range of heavy intensity) work rates (9). It has been suggested that this slowing is linked to increased type II muscle fibre recruitment associated with the greater force requirement (62, 87) and the consequent increase in lactate production and H^+ (43), $\dot{V}O_{2p}$ (31), and PCr breakdown (3). This observed slowing of $\dot{V}O_{2p}$ kinetics was attributed to the intrinsic slowness of skeletal muscle oxidative metabolism associated with the increased ATP demand and type II fibre recruitment during high intensity exercise (87).

1.4 Intermittent Exercise

In the present study, intermittent exercise patterns of differing intensities of six min in duration were utilized. From the beginning of research on intermittent exercise (Astrand et al., 1960; Christensen et al., 1960), different durations of the short work: recovery durations have been investigated: 5 min: 2 min (Fox et al., 1975); 30 s: 30 s (Astrand et al., 1960) and 10 s: 5 s (Christensen et al., 1960). The 10 s: 5 s work was only performed with a sample size of two. As such, data from Christensen et al. (1960) were speculative.

More recent work by Belfry et al. (7) has observed the effects of intermittent 5 s recovery periods of two sub-lactate threshold intensities (moderate and light) followed by 10 s of supra-lactate threshold intensity over an 8 min exercise bout (INT). In comparison to the continuous exercise performed at the same power output as the 10 s work periods of the INT, the INT with 5 s of recovery performed at moderate intensity was associated with a lower mean $\dot{V}O_{2p}$ and the 5 s of recovery performed at light intensity exercise elicited a further reduction in $\dot{V}O_{2p}$. These

observations are also consistent with previous literature linking the changes in phosphocreatine concentrations ([PCr]) with changes in $\dot{V}O_{2p}$, which has been suggested to be a proxy for oxidative phosphorylation (67). Oscillations in $\dot{V}O_{2p}$ were also observed over the work: recovery cycles. These fluctuations were matched temporally with NIRS-derived Δ [HHb] (7) that also suggested fluctuations in oxidative phosphorylation.

Since the insertion of 5 s recovery periods has been shown to increase mean [PCr] (8) and decrease mean $\dot{V}O_{2p}$ (7), repeated insertions of 5 s at higher power output in the present study would be expected to result in the opposite $\dot{V}O_{2p}$ response. These 10 s work: 5 s recovery cycles were also associated with increases in PCr-derived phosphorylation substrate level phosphorylation over the first 4 s of the work period (8). It would be expected that the proposed intermittent 5 s sprints would require a similar increase in ATP-PCr derived and substrate level phosphorylation.

Furthermore, these intermittent exercise protocols were associated with lower Δ [HHb] compared to continuous exercise (7). This was attributed to reduced intramuscular pressures (64) and impedance to flow in the exercising muscles during the rest periods (2, 21, 44). Therefore, if the 5 s periods in the present study are performed at higher power output, it is suggested that the opposite response (greater Δ [HHb] and impaired blood flow distribution or O_2 delivery) will be observed.

As previously mentioned, the adjustment of normalized [HHb]-to- $\dot{V}O_2$ ratio (Δ [HHb]/ $\dot{V}O_2$) has been utilized to provide insight to the balance between O_2 utilization to O_2 delivery (i.e. measure of muscle blood flow distribution) (55). Within the intermittent exercise paradigm, our previous work with 10 s work: 5 s recovery (7), 10 s work: 3 s (49) recovery were

associated with decreases in $\Delta[\text{HHb}]/\dot{V}\text{O}_2$ ratio. Thus, it is suggested that intermittent patterns of exercise, in comparison to continuous exercise, improves muscle blood flow distribution.

Conversely, increasing the mean work rate, as is achieved by inserting 5 s sprint periods in the present study would increase the contribution of type II fibre recruitment as well as intramuscular pressures/impedance to muscle blood flow. Thus, as has been suggested previously, the insertions of 5 s higher power output intervals will further increase $[\text{HHb}]$ and improve muscle blood flow distribution consequent to the local vasodilatory effects of La^- (40, 51, 77).

More recently, the effects of 3 s recovery periods during heavy intensity exercise (49) have shown that more frequent short recovery periods were associated with faster $\dot{V}\text{O}_{2p}$ kinetics. It was postulated that the insertions of recovery periods facilitated an improved microvasculature perfusion, and hence O_2 delivery, resulting in faster $\dot{V}\text{O}_{2p}$ kinetics (49). Conversely, if these brief periods were replaced with higher intensity exercise as opposed to recovery, it is expected that the opposite results would occur.

1.5 Hypoxia and Breath Holding During Exercise

Breathing hypoxic gas mixtures ($\sim 15\% \text{O}_2$) during moderate (56, 76) and heavy (4, 17) intensity exercise has shown to slow $\dot{V}\text{O}_{2p}$ kinetics (17, 22, 30, 76). This slowing of $\dot{V}\text{O}_{2p}$ kinetics during exercise in hypoxia has been linked to increased recruitment of less-oxidative type II muscle fibres (24, 57), which also contributes to the development of an increased gain and consequently an increased duration to reach the end of phase II. Hypoxia during exercise also has been associated with greater O_2 deficit, implying a greater reliance on substrate level

and PCr-derived phosphorylation (43), resulting in increased intracellular PCr depletion and blood La^- concentrations (29, 56) that has been associated with slower $\dot{V}\text{O}_{2p}$ kinetics.

The previous research on breath holding during exercise has shown effects comparable to hypoxia (1, 85). During a breath hold, the diffusion of O_2 from the lung to the blood continues. As such, PO_2 in the lung and pulmonary capillaries decreases, giving rise to arterial hypoxemia (65) which reduces the total O_2 delivery to exercising muscles (86, 88). With reduced O_2 availability at the muscle, contribution from substrate level phosphorylation is expected to increase, in order to replace the ATP originally derived from oxidative phosphorylation (53).

Similarly, breath holds of varying durations, from 45 s every 5 min, to 15 s every min, have resulted in reduced $\dot{V}\text{O}_{2p}$, and increased [HHb], arterialized-capillary lactate concentrations ($[\text{La}^-]$), and proton ($[\text{H}^+]$) accumulation (1, 28, 38, 93, 94). Moreover, breath holds have also been associated with increased muscle deoxygenation, as increased O_2 offloading from the Hb attempts to compensate for the reduced O_2 delivery (93). Consequently, $[\text{La}^-]$ would increase, reflecting an increased contribution from substrate level phosphorylation (66).

Consequent to the cessation of ventilation during the periods of the breath holds, increased CO_2 levels in the blood (hypercapnia) have also been observed (28, 41, 48, 85). As CO_2 is a major stimulus to the peripheral chemoreceptors (84, 88), hypercapnia would be responsible for a hyperventilatory response. However, these hyperventilatory responses associated with longer duration breath holds, of approximately 40 s, were insufficient to attenuate the decreases in $\dot{V}\text{O}_{2p}$ (41, 85). In the current study, it is possible that the 25 s free-breathing periods of each 30 s cycle would be sufficient duration to resolve the hypoxic effects of the 5 s breath hold.

Previous work on breath holds also observed a breath hold-induced bradycardia (1, 38, 42). Mechanistically, it has been suggested that breath holds after an inspiration increases intrathoracic pressure and decreases abdominal pressures (28, 60, 75). This creates a pressure gradient between the infra- and supra-diaphragmatic portions of the inferior vena cava that “pulls” the blood towards the right side of the heart (28, 60, 75). This facilitates venous return and diastole to the right side of the heart. It appears that the breath hold-induced increases in stroke volume and mean arterial pressure, and the consequent baroreflex-mediated bradycardia associated with the ‘human diving response’ observed previously with deep diving, is an attempt to reduce cardiac O₂ utilization (25, 70).

Previous work on breath holds during exercise suggests that the compensatory responses associated with breath holds and/or hypoxia are insufficient to resolve successfully the hypoxic and hypercapnic effects demanded of longer duration breath holds. Within the context of shorter duration 5 s breath holds every 30 s in the present study, a physiological resolution of the breath hold effects may be expected. However, if the 5 s breath holds are combined with the sprints, the 25 s of hyperventilation may be insufficient to support the increased ATP demand via oxidative phosphorylation.

1.6 **Breath-by-Breath Analysis by Mass Spectroscopy**

In the present study, $\dot{V}O_{2p}$ will reflect the contribution of oxidative phosphorylation to heavy intensity exercise. $\dot{V}O_{2p}$ will be measured at the mouth for breath-by-breath analysis. These breath-by-breath $\dot{V}O_{2p}$, carbon dioxide production ($\dot{V}CO_{2p}$), and minute ventilation (\dot{V}_E) determinations are calculated using the inspired and expired flow rates from a low dead-space bidirectional turbine. Inspired and expired gases were sampled continuously at the mouth and

analyzed for concentrations of O₂, CO₂, and N₂ by mass spectrometry, which were also calibrated with fixed gas mixtures. Changes in gas concentrations were aligned with gas volumes by measuring the time delay for a square-wave bolus of gas passing through the turbine, resulting in changes in fractional gas concentrations as measured by the mass spectrometer. The data were collected every 20 ms and transferred to a computer, which aligned concentrations with the volumes to build a profile of each breath.

Breath-by-breath alveolar gas exchange was calculated using algorithms (79), which were developed to estimate breath-by-breath alveolar gas exchange by accounting for changes in both functional residual capacity and alveolar gas concentrations during ventilation. Total lung gas exchange was computed with correction for accuracy.

1.7 Near – Infrared Spectroscopy and Muscle Deoxygenation

The previously observed HHb responses have been detected utilizing near-infrared spectroscopy (7, 13, 14, 49). At the onset of high intensity exercise in the present study, O₂ delivery to the muscles were provided by increased O₂ offloading by hemoglobin, consequent to greater PO₂ gradient, until the time which O₂ delivery by blood flow increases (40). Thus, oxygenated hemoglobin, deoxygenated hemoglobin, and total hemoglobin saturation provides useful information to the extent of muscle deoxygenation (15). The near-infrared spectroscopy (NIRS) was utilized to observe these changes in the exercising muscle at the site of interrogation. NIRS enables a continuous and non-invasive monitoring of the relative concentration changes in oxygenated hemoglobin ([O₂Hb]), deoxygenated hemoglobin ([HHb]), total hemoglobin concentrations ([Hb_{tot}]), and tissue hemoglobin saturation (S_{at}O₂) in the muscle microvasculature during dynamic exercise (14).

The theoretical foundation for the NIRS technology are explained in detail by Ferrari et al. (19). In brief, there are varying lengths and frequencies of waves in the electromagnetic spectrum, and infrared is the domain in between the visible and microwave domains. Thus, the infrared spectroscopy exposes organic molecules like hemoglobin (Hb) and myoglobin (Mb) to the infrared radiation, which range from 790-850 nm. Upon infrared radiation, Hb and Mb “resonate” at these wavelength frequencies and is absorbed. When infrared spectroscopy is utilized for organic tissues, it allows for the measurement of muscle $[O_2Hb]$, $[HHb]$, and $[Hb_{tot}]$ (36).

The measurement is determined by the amount of absorption of the near-infrared (NIR) light projected by the diode refracted back to the optode from the organic tissue and the NIR spectrum can particularly penetrate the organic tissue and enable absorption by Hb and Mb, with varying amounts of absorption depending on the extent of O_2 binding of these molecules. The HHb and O_2Hb absorbs different wavelengths of NIR depending on the presence of O_2 -binding (690-760 nm and 800-850 nm, respectively). Therefore, the difference in wavelength emitted by the NIRS optode from the diode provides measures of $[O_2Hb]$, $[HHb]$, and $[Hb_{tot}]$ in the microvasculature (50).

As O_2 is transported from the pulmonary capillaries towards the exercising muscles via the hemoglobin molecules, the decrease in partial pressure of oxygen (PO_2) and the partial pressure gradient facilitates O_2 diffusion (i.e. O_2 -offloading) into the muscles. Thus, the NIRS-derived $[HHb]$ signal provides insight to the balance between oxygen delivery (QO_2) and uptake ($\dot{V}O_2$) at the site of muscle interrogation (14, 46). Furthermore, the sum of $[O_2Hb]$ and $[HHb]$ is utilized to calculate a measure of $[Hb_{tot}]$. Since the total hemoglobin volume is expressed in

concentrations, $[\text{Hb}_{\text{tot}}]$ provides insight to microvascular blood flow changes at the site of interrogation.

Moreover, the adjustment of normalized $[\text{HHb}]$ -to- $\dot{V}\text{O}_2$ ratio ($[\text{HHb}]/\dot{V}\text{O}_2$) provides insight to the matching of O_2 delivery to O_2 utilization, and blood flow distribution at the site of interrogation (55). Thus, a collection of NIRS parameters will provide valuable insight to the physiological effects of 5 s periods of peak aerobic power and breath holds.

Possible limitations of the NIRS is that NIRS-derived $[\text{HHb}]$ signal may be influenced by small arteries and venules, reflecting both the vasculature and the muscle, as the absorption spectra of the myoglobin is similar to that of the hemoglobin. However, the ratio of hemoglobin to myoglobin in human skeletal muscle has been suggested to be approximately 10:1 and it has been accepted that the NIRS-derived $[\text{HHb}]$ signal can be used as a proxy for the dynamic adjustment of O_2 extraction from the hemoglobin molecules (71). In summary, the NIRS enables for a convenient and non-invasive observation of the microvascular oxygenation and blood flow changes at the site of interrogation.

1.8 Study Rationale

The current breath hold literature is limited to longer breath hold durations of 15 – 45 s every min (1, 38, 93, 94). With breathing patterns similar to elite swimming training and competitions in 50 m pools, if the breath holds are shortened to 5 s every 30 s (i.e. 10 s every min), the cardiovascular, respiratory, and metabolic responses associated with the breath holds may be sufficient to resolve the associated transient hypoxia.

Thus, the main purpose of this study was to compare and contrast $\dot{V}\text{O}_{2\text{p}}$, $\tau\dot{V}\text{O}_{2\text{p}}$, \dot{V}_E , $[\text{HHb}]$, HR, and arterialized-capillary lactate concentrations ($[\text{La}^-]$) during a 6 min constant-load

heavy intensity cycle ergometer exercise bout (CON) to three different intermittent protocols consisting of repeated 30 s cycles. The elucidation of the acute effects of the brief breath hold intervals and/or sprints in the present study will provide insight to the associated cardiovascular and respiratory responses.

1.9 References

1. **Andersson JP, Linér MH, Fredsted A, Schagatay EK.** Cardiovascular and respiratory responses to apneas with and without face immersion in exercising humans. *J Appl Physiol* 96: 1005–1010, 2004.
2. **Barcroft BYH, Dornhorst AC.** The blood flow through the human calf during rhythmic exercise. *J Physiol* 109: 402–411, 1949.
3. **Barstow J, Cooper DANM, Thomas J, Buchthal S.** Muscle energetics and pulmonary during moderate exercise oxygen uptake kinetics. *J Appl Physiol* 77: 1742–1749, 1994.
4. **Barstow TJ, Mole PA.** Linear and nonlinear characteristics of oxygen uptake kinetics during heavy exercise. *J Appl Physiol* 71: 2099–2106, 1991.
5. **Belfry GR, Paterson DH, Murias JM, Thomas SG.** The Effects of Short Recovery duration on VO₂ and muscle deoxygenation during intermittent exercise. *Eur J Appl Physiol* 112: 1907–1915, 2012.
6. **Belfry GR, Raymer GH, Marsh GD, Paterson DH, Thompson RT, Thomas SG.** Muscle metabolic status and acid-base balance during 10-s work : 5-s recovery intermittent and continuous exercise. *J Appl Physiol* 113: 410–417, 2012.
7. **Da Boit M, Bailey SJ, Callow S, Dimenna FJ, Jones AM.** Effects of interval and continuous training on O₂ uptake kinetics during severe-intensity exercise initiated from an elevated metabolic baseline. *J Appl Physiol* 116: 1068–77, 2014.
8. **Burnley M, Jones AM.** Oxygen uptake kinetics as a determinant of sports performance. *Eur J Sport Sci* 7: 63–79, 2007.
9. **Chin LMK, Heigenhauser GJF, Paterson DH, Kowalchuk JM.** Effect of hyperventilation and prior heavy exercise on O₂ uptake and muscle deoxygenation

- kinetics during transitions to moderate exercise. *Eur J Appl Physiol* 108: 913–925, 2010.
10. **Delorey DS, Kowalchuk JM, Paterson DH.** Relationship between pulmonary O₂ uptake kinetics and muscle deoxygenation during moderate-intensity exercise. *J Appl Physiol* 7: 113–120, 2003.
 11. **DeLorey DS, Shaw CN, Shoemaker JK, Kowalchuk JM, Paterson DH.** The effect of hypoxia on pulmonary O₂ uptake, leg blood flow and muscle deoxygenation during single-leg knee-extension exercise. *ExpPhysiol* 89: 293–302, 2004.
 12. **Dicker SG, Lofthus GK, Thornton NW, Brooks G.** Respiratory and heart rate responses to tethered controlled frequency breathing swimming. *Med Sci Sports Exerc* 12: 20–23, 1980.
 13. **Engelen M, Porszasz J, Riley M, Wasserman K, Maehara K, Barstow TJ.** Effects of hypoxic hypoxia on O₂ uptake and heart rate kinetics during heavy exercise. *J Appl Physiol* 81: 2500–2508, 1996.
 14. **Fédération Internationale De Natation.** Fina General Rules. *FINA Congr* : 1–11, 2014.
 15. **Ferrari M, Mottola L, Quaresima V.** Principles, techniques, and limitations of near infrared spectroscopy. *Can J Appl Physiol* 29: 463–87, 2004.
 16. **Fiedler GB, Schmid AI, Goluch S, Schewzow K, Laistler E, Niess F, Unger E, Wolzt M, Mirzahosseini A, Kemp GJ, Moser E, Meyerspeer M.** Skeletal muscle ATP synthesis and cellular H⁺ handling measured by localized ³¹P-MRS during exercise and recovery. *Sci Rep* 6: 32037, 2016.
 17. **Folkow B, Gaskell P, Waaler BA.** Blood Flow through Limb Muscles during Heavy Rhythmic Exercise. *Acta Physiol Scand* 80: 61–72, 1970.
 18. **Fukuoka Y, Poole DC, Barstow TJ, Kondo N, Nishiwaki M, Okushima D, Koga S.**

- Reduction of VO₂ slow component by priming exercise: novel mechanistic insights from time-resolved near-infrared spectroscopy. *Physiol Rep* 3: e12432, 2015.
19. **Geers C, Gros G.** Carbon dioxide transport and carbonic anhydrase in blood and muscle. *Physiol Rev* 80: 681–715, 2000.
 20. **Gollnick PD, Sjödin B, Karlsson J, Jansson E, Saltin B.** Human soleus muscle: A comparison of fiber composition and enzyme activities with other leg muscles. *Pflügers Arch Eur J Physiol* 348: 247–255, 1974.
 21. **Gooden BA.** Mechanism of the Human Diving Response. 29: 6–16, 1994.
 22. **Grassi B, Quaresima V, Marconi C, Ferrari M, Cerretelli P.** Blood lactate accumulation and muscle deoxygenation during incremental exercise. *J Appl Physiol* 87: 348–355, 1999.
 23. **Hill A V., Long CNH, Lupton H.** Muscular Exercise, Lactic Acid, and the Supply and Utilisation of Oxygen. *Ergebnisse der Physiol* 24: 43–51, 1925.
 24. **Hoffmann U, Smerecnik M, Leyk D, Essfeld O.** Cardiovascular responses to apnea during dynamic exercise. *Int J Sports Med* 26: 426–431, 2005.
 25. **Hogan MC, Cox RH, Welch HG.** Lactate accumulation during incremental exercise with varied inspired oxygen fractions. *J Appl Physiol* 55: 1134–1140, 1983.
 26. **Hughson RL, Kowalchuk JM.** Kinetics of oxygen uptake for submaximal exercise in hyperoxia, normoxia, and hypoxia. *Can J Appl Physiol* 20: 198–210, 1995.
 27. **Jones AM, Campbell ÆIT, Pringle JSM.** Influence of muscle fibre type and pedal rate on the V rate slope during ramp exercise. *Eur J Appl Physiol* 91: 238–245, 2004.
 28. **Keir DA, Robertson TC, Benson AP, Rossiter HB, Kowalchuk JM.** The influence of metabolic and circulatory heterogeneity on the expression of pulmonary VO₂ kinetics in

- humans. *Exp Physiol* 101: 176–192, 2016.
29. **Koga S, Poole DC, Fukuoka Y, Ferreira LF, Kondo N, Ohmae E, Barstow TJ.** Methodological validation of the dynamic heterogeneity of muscle deoxygenation within the quadriceps during cycle exercise. *Am J Physiol Regul Integr Comp Physiol* 301: R534–R541, 2011.
30. **Kume D, Akahoshi S, Song J, Yamagata T, Wakimoto T, Nagao M, Matsueda S, Nagao N.** Intermittent breath holding during moderate bicycle exercise provokes consistent changes in muscle oxygenation and greater blood lactate response. *J Sports Med Phys Fitness* 53: 327–335, 2013.
31. **Laughlin MH, Armstrong RB.** Muscle blood flow during locomotory exercise. *Exerc Sport Sci Rev* 13: 95–136, 2003.
32. **Lindholm P, Linnarsson D.** Pulmonary gas exchange during apnoea in exercising men. *Eur J Appl Physiol* 86: 487–491, 2002.
33. **Lindholm P, Sundblad P, Linnarsson D.** Oxygen-conserving effects of apnea in exercising men. *J Appl Physiol* 87: 2122–2127, 1999.
34. **Linnarsson D, Karlsson J, Fagraeus L, Saltin B.** Muscle metabolites with exercise and oxygen deficit and hyperoxia. *J Appl Physiol* 36: 399–402, 1974.
35. **Lutjemeier BBJ, Miura AA, Scheuermann BWBW, Koga SS, Townsend DKDK, Barstow TJJ.** Muscle contraction-blood flow interactions during upright knee extension exercise in humans. *J Appl Physiol* 98: 1575–1583, 2005.
36. **Lyttle AD, Blanksby BA, Elliott BC, Lloyd DG.** Net forces during tethered simulation of underwater streamlined gliding and kicking techniques of the freestyle turn. *J Sports Sci* 18: 801–807, 2000.

37. **Mancini DM, Bolinger L, Li H, Kendrick K, Chance B, Wilson JR.** Validation of near-infrared spectroscopy in humans. *J Appl Physiol* 77: 2740–7, 1994.
<http://jap.physiology.org/content/jap/77/6/2740.full.pdf>.
38. **Manning JM.** Physiology of Sport and Exercise. *J Athl Train* 34: 298–299, 1999.
39. **Matheson GO, McKenzie DC.** Breath holding during intense exercise: arterial blood gases, pH, and lactate. *J Appl Physiol* 64: 1947–52, 1988.
40. **McCrudden M, Keir DA, Belfry G.** The effects of short work versus longer work periods within intermittent exercise on VO₂p kinetics, muscle deoxygenation and energy system contribution. *J Appl Physiol* 122: 1435–1444, 2017.
41. **McCully KK, Hamaoka T.** Near-infrared spectroscopy: what can it tell us about oxygen saturation in skeletal muscle? *Exerc Sport Sci Rev* 28: 123–127, 2000.
42. **Murias JM, Dey A, Campos OA, Estaki M, Hall KE, Melling CWJ, Noble EG.** High-Intensity Endurance Training Results in Faster Vessel-Specific Rate of Vasorelaxation in Type 1 Diabetic Rats. *PLoS One* 8: e59678, 2013.
43. **Murias JM, Kowalchuk JM, Paterson DH.** Speeding of VO₂ kinetics with endurance training in old and young men is associated with improved matching of local O₂ delivery to muscle O₂ utilization. *J Appl Physiol* 108: 913–922, 2010.
44. **Murias JM, Spencer MD, Paterson DH.** The Critical Role of O₂ Provision in the Dynamic Adjustment of Oxidative Phosphorylation. *Exerc Sport Sci Rev* 42: 4–11, 2014.
45. **Murphy PC, Cuervo LA, Hughson RL.** A study of cardiorespiratory dynamics with step and ramp exercise tests in normoxia and hypoxia. *Cardiovasc Res* 23: 825–832, 1989.
46. **Nuutinen EM, Nishiki K, Erecinska M, Wilson DF.** Role of mitochondrial oxidative phosphorylation in regulation of coronary blood flow. *Am J Physiol - Hear Circ Physiol*

- 243: H159–H169, 1982.
47. **Paterson DJ, Wood GA, Morton AR, Henstridge JD.** The entrainment of ventilation frequency to exercise rhythm. *Eur J Appl Physiol Occup Physiol* 55: 530–537, 1986.
 48. **Paulev P -E.** Cardiac Rhythm during Breath-Holding and Water Immersion in Man. *Acta Physiol Scand* 73: 139–150, 1968.
 49. **Peronnet F, Aguilaniu B.** Lactic acid buffering, nonmetabolic CO₂ and exercise hyperventilation: A critical reappraisal. *Respir Physiol Neurobiol* 150: 4–18, 2006.
 50. **Poole DC, Barstow TJ, Gaesser, Willis, Whipp BJ.** VO₂ slow component: physiological and functional significance. *Med Sci Sports Exerc* 26: 1354–1358, 1994.
 51. **Rådegran G, Saltin B.** Muscle blood flow at onset of dynamic exercise in humans. *Am J Physiol - Hear Circ Physiol* 274: H314–H322, 1998.
<http://ajpheart.physiology.org/content/274/1/H314.abstract>.
 52. **Richardson RS, Duteil S, Wary C, Wray DW, Hoff J, Carlier PG.** Human skeletal muscle intracellular oxygenation: the impact of ambient oxygen availability. *J Physiol* 571: 415–424, 2006.
 53. **Rossiter HB, Ward SA, Doyle VL, Howe FA, Griffiths JR, Whipp BJ.** Inferences from pulmonary O₂ uptake with respect to intramuscular [phosphocreatine] kinetics during moderate exercise in humans. *J Physiol* 518: 921–932, 1999.
 54. **Rossiter HB, Ward SA, Kowalchuk JM, Howe F a, Griffiths JR, Whipp BJ.** Dynamic asymmetry of phosphocreatine concentration and O₂ uptake between the on- and off-transients of moderate- and high-intensity exercise in humans. *J Physiol* 541: 991–1002, 2002.
 55. **Schagatay E, Andersson JPA, Nielsen B.** Hematological response and diving response

- during apnea and apnea with face immersion. *Eur J Appl Physiol* 101: 125–132, 2007.
56. **Seiyama A, Hazeki O, Tamura M.** Noninvasive quantitative analysis of blood oxygenation in rat skeletal muscle. *J Biochem* 103: 419–24, 1988.
<http://www.ncbi.nlm.nih.gov/pubmed/3391996>.
57. **Siegmund GP, Edwards MR, Moore KS, Tiessen DA, Sanderson DJ, McKenzie DC.** Ventilation and locomotion coupling in varsity male rowers. *J Appl Physiol* 87: 233–242, 1999. <http://jap.physiology.org/content/87/1/233.short>.
58. **Skorski S, Faude O, Caviezel S, Meyer T.** Reproducibility of Competition Pacing Profiles in Elite Swimmers. *Int J Sports Physiol Perform* 9: 217–225, 2013.
59. **Song SH, Lee WK, Chung YA, Hong SK.** Mechanism of apneic bradycardia in man. *J Appl Physiol* 27: 323–327, 1969.
60. **Springer C, Barstow T, Wasserman K, Cooper D.** Oxygen uptake and heart rate responses during hypoxic exercise in children and adults. *Med. Sci. Sports Exerc.* 23: 71–79, 1991. <http://europepmc.org/abstract/MED/1997815>.
61. **Stocker F, Oldershausen C Von, Paternoster FK, Schulz T, Oberhoffer R.** End-exercise DHHb/DVO₂ and post-exercise local oxygen availability in relation to exercise intensity. *Clin. Physiol* (2015). doi: 10.1111/cpf.12314.
62. **Swanson GD, Sherrill DL.** On the breath-to-breath estimation of gas exchange. *J Appl Physiol* 56: 259–261, 1984.
63. **Veiga S, Cala A, Mallo J, Navarro E, Arellano R, Terres-Nicoli JM, Redondo JM, Veiga S, Roig A.** Underwater and surface strategies of 200 m world level swimmers. *J Sports Sci* 34: 1–6, 2015.
64. **Wasserman K, Van Kessel AL, Burton GG.** Interaction of physiological mechanisms

- during exercise. *J Appl Physiol* 22: 71–85, 1967.
65. **Wasserman K, Whipp BJ, Castagna J.** Cardiodynamic hyperpnea: hyperpnea secondary to cardiac output increase. *J Appl Physiol* 36: 457–464, 1974.
 66. **Wein J, Andersson JP, Erdéus J.** Cardiac and ventilatory responses to apneic exercise. *Eur J Appl Physiol* 100: 637–644, 2007.
 67. **Whipp BJ.** The hyperpnea of dynamic muscular exercise. *Exerc Sport Sci Rev* 5: 295–311, 1977.
 68. **Whipp BJ.** The Slow Component of O₂ uptake kinetics during heavy exercise. *Med Sci Sports Exerc* 26: 1319–1326, 1994.
 69. **Whipp BJ, Davis JA.** Peripheral chemoreceptors and exercise hyperpnea. *Med Sci Sports Exerc* 11: 204–212, 1979.
 70. **Whipp BJ, Ward SA, Lamarra N, Davis JA, Wasserman K.** Parameters of ventilatory and gas exchange dynamics during exercise. *J Appl Physiol* 52: 1506–13, 1982.
<http://www.ncbi.nlm.nih.gov/pubmed/6809716>.
 71. **Whipp BJ, Wasserman K.** Oxygen uptake kinetics work for various intensities of constant-load. *J Appl Physiol* : 351–356, 1972.
 72. **Woorons X, Bourdillon N, Vandewalle H, Lamberto C, Mollard P, Richalet JP, Pichon A.** Exercise with hypoventilation induces lower muscle oxygenation and higher blood lactate concentration: Role of hypoxia and hypercapnia. *Eur J Appl Physiol* 110: 367–377, 2010.
 73. **Woorons X, Mollard P, Pichon A, Duvallet A, Richalet JP, Lamberto C.** Prolonged expiration down to residual volume leads to severe arterial hypoxemia in athletes during submaximal exercise. *Respir Physiol Neurobiol* 158: 75–82, 2007.

Chapter 2

2 **PHYSIOLOGICAL RESOLUTION OF PERIODIC BREATH HOLDING DURING HEAVY INTENSITY FARTLEK EXERCISE**

2.1 **Introduction**

Many sports require brief intermittent periods of maximal power output such as cycling (59), rowing (72) and swimming (16), in both training and competition. There are also periods where an athlete must produce maximal power while breathing irregularly or not breathing at all. For example, during backstroke swimming, breath holds are required as the swimmers push off the wall and kick underwater after a turn. In a 50 m pool, elite backstroke swimmers may perform these underwater kicking phases at various intensities that may be sustained for approximately 5 s (80). After surfacing, swimmers are able to breathe freely while completing the remaining distance in approximately 25 s (73). Little is known about the physiological consequences to this type of intermittent breath holding during high intensity exercise. The elucidation and interpretation of the acute singular and combined physiological effects of intermittent 5 s breath holds, while performing various intensities of work, followed by 25 s of free-breathing may provide insights to exercise performance.

Previous research has shown that both breath holding of longer durations (15 to 45 s every min to 5 min, respectively), and breathing low partial pressure of O₂ gases during exercise have been associated with decreases and increases in alveolar and arterial partial pressures of O₂ (PO₂; hypoxemia) and CO₂ (PCO₂; hypercapnia), respectively (38, 82). These conditions resulted

in decreased pulmonary oxygen uptake ($\dot{V}O_{2p}$) (1), slowed $\dot{V}O_{2p}$ kinetics (17, 30, 76), and increased muscle deoxygenation ([HHb]) (38, 92), as well as arterialized-capillary lactate concentrations ([La⁻]), compared to continuous exercise (28, 41, 48, 84, 85, 88).

During steady state exercise, the reduced alveolar and arterial PO₂, and increased PCO₂ are reflected in the decreased end-tidal PO₂ (P_{ET}O₂) and increased PCO₂ (P_{ET}CO₂) in the expired air. It is possible that decreased PO₂, increased PCO₂ and [La⁻] provoked by the 5 s breath hold would induce a compensatory increase in ventilation during the 25 s free-breathing intervals (86), which would be sufficient to resolve the breath hold-induced hypoxemic condition.

Breath holding after an inspiration has also resulted in increased intrathoracic pressures, inducing temporary increases in venous return and stroke volumes that results in reflex bradycardia, and thus, reduced O₂ delivery (1, 28, 38, 42, 60, 75). If the aforementioned 5 s breath holds are repeated every 30 s, it is possible that a similar cardiovascular response would be observed. Previously, near-infrared spectroscopy-derived muscle deoxygenation (Δ [HHb]) and tissue hemoglobin saturation (S_{at}O₂) have been shown to reflect changes in PO₂ associated with the coupling of O₂ utilization to oxidative phosphorylation, and/or a change in microvascular blood flow under constant $\dot{V}O_{2p}$ conditions (6). If the breath holds result in hypoxemia and hypercapnia, it is expected that muscle deoxygenation would increase in an attempt to maintain O₂ utilization at the muscle.

In previous intermittent exercise studies, it has been found that reducing the mean power output by inserting 5 s active recovery periods at regular intervals within heavy intensity exercise (supra-lactate threshold) decreased mean $\dot{V}O_{2p}$ and muscle deoxygenation (7). Furthermore, with similar decreases in power output, others have also observed faster $\dot{V}O_{2p}$ kinetics and decreased [La⁻] (9, 58). Thus, it is proposed in the present study that with the

inclusion of 5 s periods of higher power output, known as fartlek exercise, a differential slowing of $\dot{V}O_{2p}$ kinetics, and increased mean $\dot{V}O_{2p}$, and $[La^-]$ would result. Moreover, combining these 5 s periods of higher power output with the breath holds may also result in an attenuation of the expected increase in $\dot{V}O_{2p}$ associated with the increased mean power output, through the replacement of oxidative phosphorylation with increased substrate level phosphorylation, that would be reflected in increased $[La^-]$ (66).

Thus, the purpose of this study was to compare and contrast $P_{ET}O_2$ and $P_{ET}CO_2$, ventilation, $\dot{V}O_{2p}$, $\dot{V}O_{2p}$ kinetics, muscle deoxygenation, and $[La^-]$ during continuous heavy intensity cycling exercise with free-breathing (CON), with three differing exercise conditions: 1) Continuous heavy intensity exercise with repeated 5 s breath holds every 30 s (CBH), 2) Repeated 30 s cycles comprised of 25 s of heavy intensity exercise and 5 s of higher power output (fartlek) with free-breathing (FLK) and, 3) combining the 5 s breath holds with 5 s of higher power output during heavy intensity exercise (fartlek breath hold (FBH)).

It was hypothesized that, in comparison to CON: 1) CBH would result in similar $\dot{V}O_{2p}$ kinetics and mean $\dot{V}O_{2p}$ as a function of increased ventilation stemming from the breath holds; 2) FLK would result in similar $\dot{V}O_{2p}$ kinetics, and increased mean $\dot{V}O_{2p}$, ventilation, and $[La^-]$, however, 3) when the two perturbations are combined, the attempted physiological resolution to the increased $\dot{V}O_{2p}$ demand and the breath hold-induced hypoxemia would be overwhelmed, resulting in similar mean $\dot{V}O_{2p}$, but slower $\dot{V}O_{2p}$ kinetics, and increased ventilation and $[La^-]$.

2.2 Methods

Participants. Ten adult males (24 ± 3 years old) volunteered and gave written consent to participate in this study. All procedures were approved by the Western University Research

Ethics Board for Health Sciences Research Involving Human Participants. All participants were healthy, recreationally active (moderate intensity activities 1-3 times per week), and non-smokers. No participants were taking any medications that would affect the cardiorespiratory or hemodynamic responses to exercise.

Testing protocol. Participants were asked to maintain their usual levels of physical activity throughout their participation in the present study and to refrain from drinking caffeine 6 hours prior to their tests. All tests were performed on an electronically braked cycle ergometer on five separate days, with a minimum of 48 hours of recovery after each test. During each test, the participants were required to wear a nose-clip to prevent the participant from breathing through their nose, and a rubber mouthpiece, similar to that of breathing through a snorkel or diving mask.

Testing Day 1: Incremental ramp test to fatigue on a cycling ergometer with a work rate increment of 25 Watts (W) per minute was performed with verbal encouragement to facilitate peak efforts. These baseline tests took approximately 15 min to complete and were used to determine the $\dot{V}O_{2peak}$ and the estimated lactate threshold (LT) to prescribe the work rate for the heavy intensity (HVY) and peak aerobic power outputs in subsequent tests. The LT was defined as the $\dot{V}O_{2p}$ at which $\dot{V}CO_{2p}$ began to increase out of proportion to $\dot{V}O_{2p}$ with a systematic rise in minute ventilation-to- $\dot{V}O_{2p}$ ratio and end-tidal PO_2 whereas minute ventilation-to- $\dot{V}CO_{2p}$ ratio and end-tidal PCO_2 were stable. The LT was determined through visual inspection by two researchers familiar with this procedure. Data analysis of the LT began after accounting for the delay between $\dot{V}O_{2p}$ and work rate (the cardiodynamic phase) during the incremental ramp test to fatigue.

Testing Day 2: Participants performed a ‘square-wave’ cycling exercise test that began with a 3 min warm-up with light intensity cycling (20 W) followed by a step transition to HVY for 6 min with free-breathing (CON). The work rate during the HVY corresponded to the work rate at which the participant’s $\dot{V}O_{2p}$ was at 50% difference between the LT and $\dot{V}O_{2peak}$ ($\Delta 50$) during their incremental ramp test.

Testing Day 3: Participants performed a ‘square-wave’ cycling exercise test that began with a 3 min light intensity cycle (20 W) followed by a step transition to HVY for 6 min while performing a breath hold protocol (CBH). The breath hold (BH) protocol required repeated 30 s cycles of a 25 s of non-regulated breathing followed by a 5 s of BH. This sequence was performed repeatedly over the total four min light intensity and six 6 min HVY exercise bout. To ensure proper execution, participants were given a 5 s verbal count-down leading into each BH. Participants were also instructed to regulate their breathing during this 5 s lead in period to ensure an inspiration initiated the BH.

Testing Day 4: Participants performed a ‘square-wave’ cycling exercise test that began with a 3 min light intensity cycle (20 W) followed by repeated 30 s cycles comprised of 25 s at $\Delta 50$, and 5 s at the peak work rate attained during ramp incremental test (sprints). Free-breathing was performed throughout.

Testing Day 5: Participants performed a ‘square-wave’ cycling exercise test that began with a 3 min light intensity cycle (20 W) followed by repeated 30 s cycles comprised of 25 s at $\Delta 50$ with free-breathing and 5 s sprints combined with BH (FBH) for 6 min. The order of performing the four conditions (CON, CBH, FLK, and FBH) were systematically randomized via the iPhone application.

Measurements. Inspired and expired gases were measured breath-by-breath by utilizing a mass spectrometer (Innovision, AMIS 2000, Lindvedvej, Denmark). Gas collection for inspired and expired flow rates were also measured with a low-dead-space (90 ml) bidirectional turbine (Alpha Technologies, VMM 110) and pneumotach (Hans Rudolph, Model 4813) which were calibrated before each test by using a syringe of known volume. Inspired and expired gases were sampled continuously at the mouth and analyzed for concentrations of O₂, CO₂, and N₂ by mass spectrometry after calibration with precision-analyzed gas mixtures. Changes in gas concentrations were aligned with gas volumes by measuring the time delay for a square-wave bolus of gas passing the turbine to the resulting changes in fractional gas concentrations as measured by the mass spectrometer. Data were collected every 20 ms and transferred to a computer, which aligned concentrations with volume information to build a profile of each breath. The measurement for each breath began with the inspiration and concluded with the expiration, thus enabling to capture the breath during each breath hold.

During the exercise, the vastus lateralis of the quadriceps muscle was monitored continuously by Near-Infrared Spectroscopy (NIRS; Oxiplex TS, model 95205, ISS, Champaign, IL). The NIRS system was arranged as a single channel consisting of eight laser diodes operating at two wavelengths (690 and 828 nm, 4 at each wavelength) that were pulsed in a rapid succession (frequency modulation of laser intensity was 110 MHz) and a photomultiplier tube. The lightweight plastic NIRS probe (connected to laser diodes and a photomultiplier tube by optical fibres) consisted of two parallel rows of light-emitter fibres and one detector fibre bundle; the source-detector separations for this probe were 2.0, 2.5, 3.0, and 3.5 cm for both wavelengths. The probe was placed on the muscle belly midway between the lateral epicondyle and greater trochanter of the femur; it was secured in place with an elastic strap tightened to

prevent movement. The outline of the probe was marked with a permanent marker for future tests. This placement allowed for accurate and continuous measurement of absolute concentration changes (μM) in muscle oxyhemoglobin ($[\text{O}_2\text{Hb}]$), deoxyhemoglobin ($[\text{HHb}]$), total hemoglobin concentration ($[\text{Hb}_{\text{tot}}]$), and tissue hemoglobin saturation (S_{atO_2}). The location of measurement was covered with an optically dense, black vinyl sheet, to minimize the intrusion of extraneous light. The thigh was wrapped with elastic bandages to further minimize intrusion of extraneous light and movement of the probe. NIRS measurements started 60 s before each test and were collected continuously throughout the entire duration of each test.

The NIRS instrument was calibrated at the beginning of each testing session following a warm-up period of 10 minutes. The calibration was done with the probe placed on a calibration block (phantom), with absorption (μ_{A}) and reduced scattering coefficients (μ_{S}) previously measured; thus, correction factors were determined and were automatically implemented by the manufacturer's software for the calculation of the μ_{A} and μ_{S} for each wavelength during the data collection. Calculation of $[\text{HHb}]$ reflected continuous measurements of μ_{S} throughout each testing session (i.e. constant scattering value not assumed). Data were stored online at an output frequency of 25 Hz but were reduced to 1 s bins for all subsequent analyses and zeroed to the baseline of each test.

Heart rate (HR) was measured continuously by a heart rate monitor (Polar Electro T34) using PowerLab (ML132/ML880, ADInstruments, Colorado Springs, CO) and was calculated (using a 5 s rolling average) based on the RR interval. Data were recorded using LabChart version 6.1 (ADInstruments) on a separate computer.

Arterialized-capillary blood lactate concentrations ($[\text{La}^-]$) were measured 3 min before and 3 min after each test. Prior to the use of the lancet, a topical thermogenic (Finalgon,

Boehringer Ingelheim) was applied onto the left index finger then sterilized with a rubbing alcohol swab for each test. Blood was revealed using an ACCU-CHEK Safe-T-Pro Plus sterile, single use lancing device and was immediately analyzed by SensLab GmbH Lactate SCOUT arterialized-capillary lactate analyzer (mmol L^{-1}). Latex gloves were worn by the attending researcher.

Data analysis. Breath-by-breath gas exchange data were filtered by removal of aberrant data points that lay 3 SD above and below the local mean (39). Data for each protocol were then interpolated linearly to 1 s intervals and time-aligned, such that time 0 represented the increase from the 20 W cycling period to the HVY. This second-by-second data were then averaged into 5 s bins for statistical analysis and graphing. [HHb] and [Hb_{tot}] data were zeroed with the baseline [HHb] and [Hb_{tot}] values determined by the average of 60 s before the step-transition to yield the changes in concentrations respective to their baseline values ($\Delta[\text{HHb}]$ and $\Delta[\text{Hb}_{\text{tot}}]$), and raw S_{atO_2} data was reported. The adjustment of normalized-[HHb]-to- $\dot{V}\text{O}_{2\text{p}}$ ratio ($\Delta[\text{HHb}]/\dot{V}\text{O}_{2\text{p}}$) was obtained by zeroing and normalizing both the [HHb] and $\dot{V}\text{O}_{2\text{p}}$ data as percent changes, as previously described (54). The baseline values were considered “0,” and peak values determined from the final 30 s of the exercise were considered “100%.” to the steady state response. Analysis of the mean data of the different physiological measures were limited from 120 s to end exercise to eliminate the initial $\dot{V}\text{O}_{2\text{p}}$ kinetic response.

The on-transient of each $\dot{V}\text{O}_{2\text{p}}$ profile was modeled with the following mono-exponential function:

Equation 1.

$$y(t) = y_{\text{BSL}} + A_p \cdot (1 - e^{-(t-\text{TD})/\tau})$$

In this equation, $y(t)$ is the value of the dependent variable at any time during the transition, y_{BSL} is the pre-transition baseline value, A_p is the steady-state increase in y above the baseline value, τ is the time constant of the response or the time for y to increase to 63% of the new steady-state, and TD is the time delay. The details of the fitting procedure are described elsewhere (34). Briefly, the Levenberg-Marquardt algorithm was applied to find the minimum sum of squared residuals between the mono-exponential function and the experimental data using specialized software (Origin 8.5; OriginLab, Northampton, MA). The phase I-phase II transition was determined by examining the change in τ and CI_{95} of the fitting window from the end of exercise, and demarcated as the point at which there was a significant increase in τ and/or CI_{95} closer to the onset of exercise. The end of the phase II fitting window was determined by examining the change in τ , CI_{95} , χ^2 , and plotted residuals in response to progressive increases at the end of the fitting window. The point at which there was a systematic increase in τ , CI_{95} , and χ^2 was considered as the end of phase II.

Statistics. Analysis of the results ($n = 10$) between each exercise condition (CON, CBH, FLK, FBH) on changes in $\dot{V}O_2$, [HHb], $\dot{V}CO_{2p}$, \dot{V}_E , $P_{ET}O_2$, HR, and [La⁻] were calculated by one-way repeated-measures (RM) ANOVA. Significant differences were further tested by Tukey post hoc analysis. Data are reported as mean \pm SD unless otherwise presented. Statistical significance was declared when $p < 0.05$.

2.3 Results

Participant characteristics. Summary of the anthropometric characteristics and performance variables assessed during the ramp incremental tests are shown in Table 1.

$\dot{V}O_{2p}$ and $\dot{V}O_{2p}$ kinetics. Mean $\dot{V}O_{2p}$ from 120 s to the end of exercise was greater in FLK than all the other conditions (Table 2; Figure 1; $p < 0.05$). $\dot{V}O_{2p}$ at the end of exercise ($\dot{V}O_{2end}$) during FLK was greater than CON (Table 2; $p < 0.05$). $\dot{V}O_{2p}$ fluctuations were observed during CBH and FBH during each 30 s cycles; last 5 s periods were lower than the greatest 25 s period during FBH (Table 2; $p < 0.05$). There were no fluctuations in CON and FLK over the same period (Table 3; $p > 0.05$). $\tau\dot{V}O_{2p}$ was greater in FBH than CON and FLK (Table 2; $p < 0.05$) but similar in CBH and FLK to CON (Table 2; $p > 0.05$).

End-tidal partial pressure of oxygen ($P_{ET}O_2$) and carbon dioxide ($P_{ET}CO_2$). The mean $P_{ET}O_2$ from 120 s to the end of exercise was greater in FLK and FBH than CON (Table 2; Figure 2a; $p < 0.05$), and similar in CBH to CON (Table 2; $p > 0.05$). Mean $P_{ET}O_2$ fluctuations were observed during the last 5 s of CBH and FBH compared to the highest 5 s period in their respective 25 s periods of each 30 s cycle (Table 3; Figure 2b; $p < 0.05$) but not during FLK (Table 3; $p > 0.05$). The mean $P_{ET}CO_2$ from 120 s to the end of exercise was lower in FLK than CON, greater in FBH than FLK (Table 2; Figure 3a $p < 0.05$) but similar in CBH to CON (Table 3; Figure 2b; $p > 0.05$). Mean $P_{ET}CO_2$ fluctuations were observed during the last 5 s period of CBH and FBH compared to the lowest 5 s period in their respective 25 s periods of each 30 s cycle (Table 3; Figure 3b; $p < 0.05$) but not during FLK (Table 3; $p > 0.05$).

Carbon dioxide production ($\dot{V}CO_{2p}$) and minute ventilation (\dot{V}_E). The mean $\dot{V}CO_{2p}$ from 120 s to the end of exercise was greater in FLK than CON and lower in FBH than FLK (Table 2; $p < 0.05$) but similar in CBH to CON (Table 2; $p > 0.05$). Mean $\dot{V}CO_{2p}$ fluctuations were observed during the last 5 s period of CBH and FBH compared to the lowest 5 s period in their

respective 25 s periods of each 30 s cycle (Table 3; $p < 0.05$) but not during FLK (Table 3; $p > 0.05$). The mean \dot{V}_E from 120 s to the end of exercise was greater in CBH, FLK, and FBH than CON, greater in FBH than CBH (Table 2; Figure 4; $p < 0.05$), and similar in FBH and FLK (Table 2; Figure 4; $p > 0.05$). Mean \dot{V}_E fluctuations were observed during the last 5 s period of CBH and FBH compared to the lowest 5 s period in their respective 25 s periods of each 30 s cycle (Table 3; $p < 0.05$).

Total hemoglobin concentration ($\Delta[Hb_{tot}]$), muscle deoxygenation ($\Delta[HHb]$), tissue hemoglobin saturation ($S_{at}O_2$). A summary of NIRS measures are presented in Table 2. The mean $\Delta[Hb_{tot}]$ changes from baseline values from 120 s to the end of exercise during CBH and FLK was lower than CON, and FLK was lower than FBH (Table 2; Figure 5a; $p < 0.05$). The mean $\Delta[HHb]$ changes from baseline values from 120 s to the end of exercise were different between all conditions; CBH lower than CON, FBH lower than CBH, FLK lower than FBH (Table 2; Figure 5b; $p < 0.05$). The $S_{at}O_2$ from 120 s to the end of exercise during CBH, FLK, and FBH were greater than CON (Table 2; Figure 5c; $p < 0.05$). Mean $\Delta[HHb]$ fluctuations were not observed during the last 5 s of CBH and FBH of each 30 s cycle (Table 3; $p < 0.05$).

Adjustment of normalized $\Delta[HHb]$ -to- $\dot{V}O_{2p}$ ratio ($\Delta[HHb]/\dot{V}O_{2p}$). Mean $\Delta[HHb]/\dot{V}O_{2p}$ from 0 – 360 s was similar between all conditions (Table 2; Figure 6; $p > 0.05$). Mean $\Delta[HHb]/\dot{V}O_{2p}$ of the last 5 s of each 30 s cycle were greater than the lowest 5 s point during the 25 s in CBH and FBH (Table 3; Figure 3; $p < 0.05$). Mean $\Delta[HHb]/\dot{V}O_{2p}$ fluctuations were observed during the last 5 s of each 30 s cycle during CBH and FBH (Table 3; $p < 0.05$).

Heart rate and O₂-Pulse. The mean heart rate was different between all conditions from 120 s to the end of exercise; CBH lower than CON, FBH lower than FLK, FLK greater than CON (Table 2; $p < 0.05$). The mean O₂-Pulse from 120 s to the end of exercise was greater in CBH than CON and lower in FBH than CON, but similar in FLK to CON (Table 2; $p > 0.05$).

Arterialized-capillary lactate concentration ([La⁻]). The mean pre-exercise [La⁻] across all conditions were similar (Table 2; $p > 0.05$) and the mean post-exercise [La⁻] in FBH was greater than all other conditions (Table 2; $p < 0.05$).

Table 1. Participant characteristics and performance variables from the ramp incremental test including age, height, body mass, $\dot{V}O_{2peak}$, estimated LT, peak PO, and PO at $\Delta 50$.

	Age (years)	Height (cm)	Body mass (kg)	$\dot{V}O_{2peak}$ (L·min ⁻¹)	Estimated LT (L·min ⁻¹)	PO at $\Delta 50$ (W)	Peak PO (W)
Mean	24	179	80	3.17	1.77	218	314
SD	3	6	9	0.52	0.22	30	49

$\dot{V}O_{2peak}$: Peak pulmonary oxygen uptake ($\dot{V}O_2$), estimated *LT*: estimated lactate threshold, *PO at $\Delta 50$* : power output at 50 percent of the difference between the LT and $\dot{V}O_{2peak}$, *Peak PO*: peak power output attained during ramp incremental test (sprints)

Table 2. Summary of physiological parameters collected during CON, CBH, FLK, and FBH.

	CON	CBH	FLK	FBH		CON	CBH	FLK	FBH
Mean PO (Watts)	218	218	234*	234*	HR (bpm)	169	166*†	175*	173*§†
SD	30	30	33	323	SD	7	7	7	6
$\dot{V}O_{2p}$ (L·min ⁻¹)	2.71	2.73	2.85*	2.73†	O ₂ -Pulse (mlO ₂ ·min ⁻¹ ·HR beat ⁻¹)	16.3	16.7*	16.3	15.7*§†
SD	0.12	0.14	0.12	0.14	SD	0.3	0.7	0.4	0.8
$\dot{V}O_{2p}$ BSL (L·min ⁻¹)	0.92	0.90	0.93	0.90	Δ [Hb _{tot}] (uM)	4.94	3.85*	2.44*§	5.42*§†
SD	0.04	0.03	0.04	0.05	SD	0.4	0.7	0.7	0.8
$\dot{V}O_{2p}$ end (L·min ⁻¹)	2.88	2.83	2.97*	2.86	Δ [HHb] (uM)	16.3	14.4*	12.0*§	13.9*§†
SD	0.02	0.10	0.04	0.10	SD	0.7	0.6	0.4	0.5
$\tau\dot{V}O_{2p}$ (s)	48.8	53.2	44.1§†	62.2*	S _{at} O ₂ (%)	51.7	55.5*†	54.4*	55.5*†
SD	12	11	8	19	SD	0.7	0.5	0.2	0.5
$\dot{V}CO_{2p}$ (L·min ⁻¹)	3.12	3.16	3.43*‡	3.28*§	Δ [HHb]/ $\dot{V}O_{2p}$	0.99	0.99	1.00	1.00
SD	0.13	0.38	0.16	0.35	SD	0.13	0.15	0.12	0.15
\dot{V}_E (L·min ⁻¹)	89.74	97.58*	105.0*§	100.9*	pre [La ⁻] (mM)	2.0	1.8	1.7	1.7
SD	7	16	10	19	SD	0.5	0.3	0.4	0.4
P _{ET} O ₂ (mmHg)	103.9	105.4	108.4*	106.4*§	post [La ⁻] (mM)	9.0‡	10.0‡	9.9‡	11.5
SD	2	5	2	5	SD	2.3	2.4	2.0	2.7
P _{ET} CO ₂ (mmHg)	42.3	42.7	40.2*	42.2§†					
SD	2	4	3	4					

Data analyzed by one way repeated measures ANOVA. Values are given as means \pm SD. Mean data are from 120 s – 360 s. *CON*: continuous exercise at Δ 50 with free-breathing; *CBH*: continuous exercise at Δ 50 with repeated 30 s cycles comprised of 25 s free-breathing periods and 5 s breath holds; *FLK*: continuous exercise with repeated 30 s cycles comprised of 25 s Δ 50 and 5 s sprints with free-breathing; *FBH*: 6 min of continuous exercise with repeated 30 s cycles comprised of 25 s Δ 50 with free-breathing and 5 s sprints combined with a breath hold; *Mean PO*: mean power output; $\dot{V}O_{2p}$: mean pulmonary oxygen uptake; *BSL*: baseline during 4 min of 20 W cycling; *end*: last 30 s of exercise; τ : time constant representing 63% of time to reach steady state; $\dot{V}CO_{2p}$: mean carbon dioxide production; \dot{V}_E : mean minute ventilation; *P_{ET}O₂*: mean end-tidal partial pressures of oxygen; *P_{ET}CO₂*: mean end-tidal partial pressures of carbon dioxide; *HR*: mean heart rate; *O₂-Pulse*: mean oxygen utilization per heart beat; Δ [Hb_{tot}]: mean change in total hemoglobin concentration from baseline values; Δ [HHb]: mean change in deoxygenated hemoglobin concentration from baseline values; S_{at}O₂: mean tissue hemoglobin saturation; Δ [HHb]/ $\dot{V}O_{2p}$: adjustment of normalized [HHb]-to-normalized $\dot{V}O_{2p}$ ratio from 0 – 360 s; [La⁻]: arterialized-capillary lactate concentration; * different from CON, § different from CBH, † different from FLK, ‡ different from FBH

Table 3. Summary of fluctuations in $\dot{V}O_{2p}$, $\dot{V}CO_{2p}$, \dot{V}_E , $P_{ET}O_2$, $P_{ET}CO_2$, $\Delta[HHb]$, and $\Delta[HHb]/\dot{V}O_{2p}$ over the 25 s and 5 s intervals within the 30 s cycles of CON, CBH, FLK, and FBH.

	CON		CBH		FLK		FBH	
	25 s	5 s	25 s	5 s	25 s	5 s	25 s	5 s
$\dot{V}O_{2p}$ (L·min ⁻¹)	2.58	2.36	2.67*	2.25*	2.72*	2.48*	2.70*	2.20*‡
SD	0.4	0.5	0.4	0.5	0.4	0.5	0.5	0.5
$\dot{V}CO_{2p}$ (L·min ⁻¹)	2.87	2.60	3.13*	2.03*‡	3.20*	2.83*	3.29*	2.07*†‡
SD	0.6	0.7	0.8	0.5	0.7	0.8	0.9	0.7
\dot{V}_E (L·min ⁻¹)	83.2	72.8	96.9*	59.1*‡	97.7*	83.9*	100*	57.2*†‡
SD	20	20	25	19	23	26	27	16
$P_{ET}O_2$ (mmHg)	103	97	107*	88*‡	107*	100*	108*	89*†‡
SD	4	6	4	12	4	9	5	13
$P_{ET}CO_2$ (mmHg)	41	43	39*	49*‡	39*	42 β	38*	49*†‡
SD	2	2	2	3	3	3	3	3
$\Delta[HHb]$ (uM)	14.3	16.1	14.1*	12.5*	10.5*	11.9*	12.1*†	13.63*§†
SD	4	1	4	1	3	1	4	1
$\Delta[HHb]/\dot{V}O_{2p}$	0.9	1.0	0.9	1.1*‡	0.9	1.0	0.8†	1.1*‡
SD	0.2	0.1	0.2	0.1	0.2	0.1	0.2	0.2

Data analyzed by two way repeated measures ANOVA. Values are given as means \pm SD. 25 s: represents the peak ($\dot{V}O_{2p}$, $\dot{V}CO_{2p}$, \dot{V}_E , $P_{ET}O_2$) or nadir ($P_{ET}CO_2$, $\Delta[HHb]/\dot{V}O_{2p}$) 5 s values during the first 25 s period of each 30 s cycle; 5 s: represents the last 5 s of each 30 s cycle; CON: continuous; CBH: continuous breath hold; FLK: FLK; FBH: FLK breath hold; $\dot{V}O_{2p}$: mean pulmonary oxygen uptake from 0 – 360 s; $\dot{V}CO_{2p}$: mean carbon dioxide production from 0 – 360 s; \dot{V}_E : mean minute ventilation from 0 – 360 s; $P_{ET}O_2$: mean end-tidal partial pressures of oxygen; $P_{ET}CO_2$: mean end-tidal partial pressures of carbon dioxide; $\Delta[HHb]$: change in deoxygenated hemoglobin concentration; $\Delta[HHb]/\dot{V}O_{2p}$: mean ratio of adjustment of normalized [HHb]-to-normalized $\dot{V}O_{2p}$; ‡ different from 25 s within the same condition, * different from CON, § different from CBH, † different from FLK

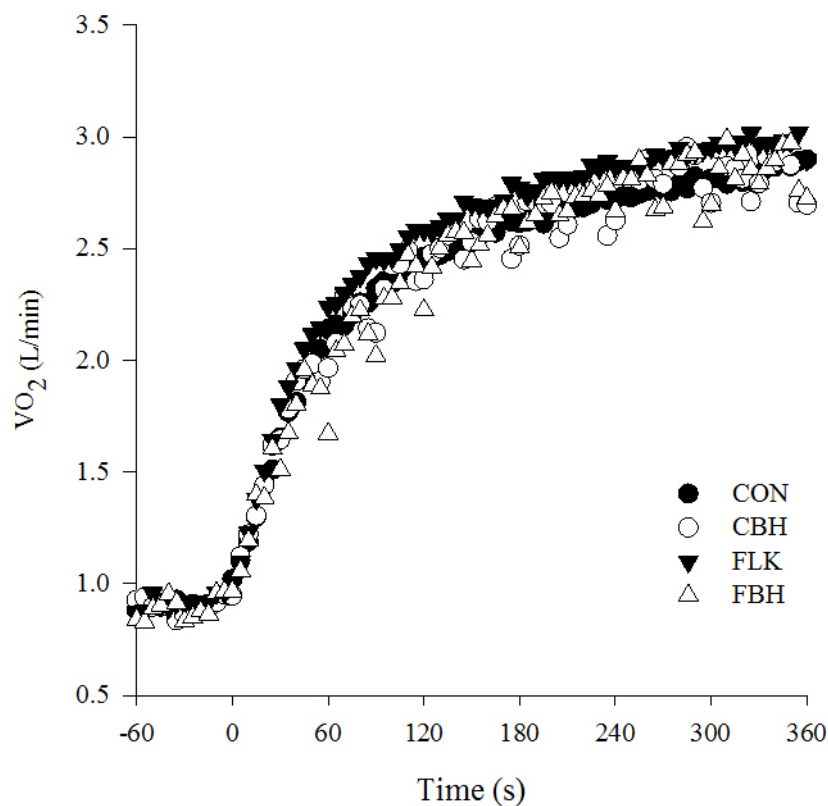


Figure 1. Mean pulmonary oxygen uptake ($\dot{V}O_{2p}$) profile during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2peak}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The mean $\dot{V}O_{2p}$ from 120 s to the end of exercise was greater in FLK than all other conditions ($p < 0.05$).

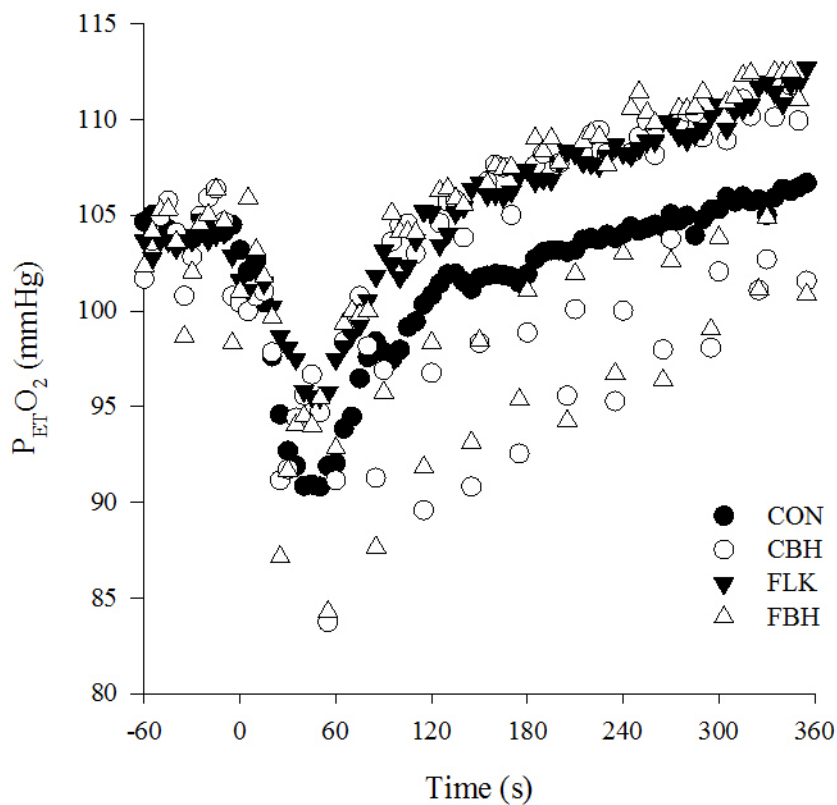


Figure 2a. Mean end-tidal partial pressure of oxygen ($P_{ET}O_2$) during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2peak}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The $P_{ET}O_2$ from 120 s to the end of exercise was greater in FLK than CON, greater in FBH than CON, and lower in FBH than FLK ($p < 0.05$).

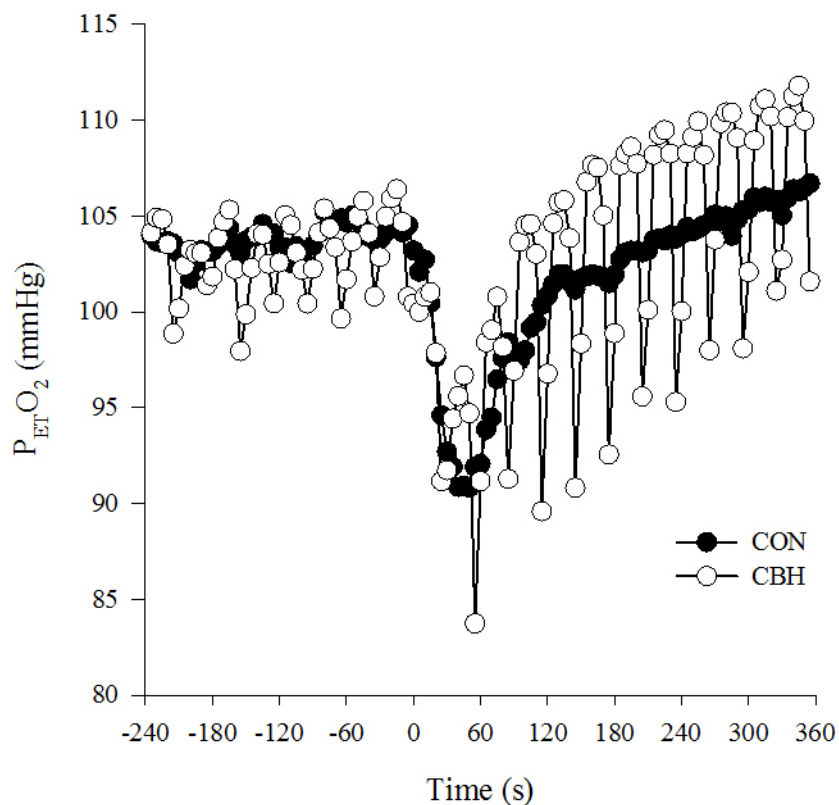


Figure 2b. Fluctuations in mean end-tidal partial pressure of oxygen ($P_{ET}O_2$) during CON and CBH. This reflects the acute resolution of transient hypoxia associated with each breath hold episode. The $P_{ET}O_2$ during the last 5 s (lowest oscillation point shown in the figure) were lower than the peak values during the 25 s periods over the 30 s cycles ($p < 0.05$).

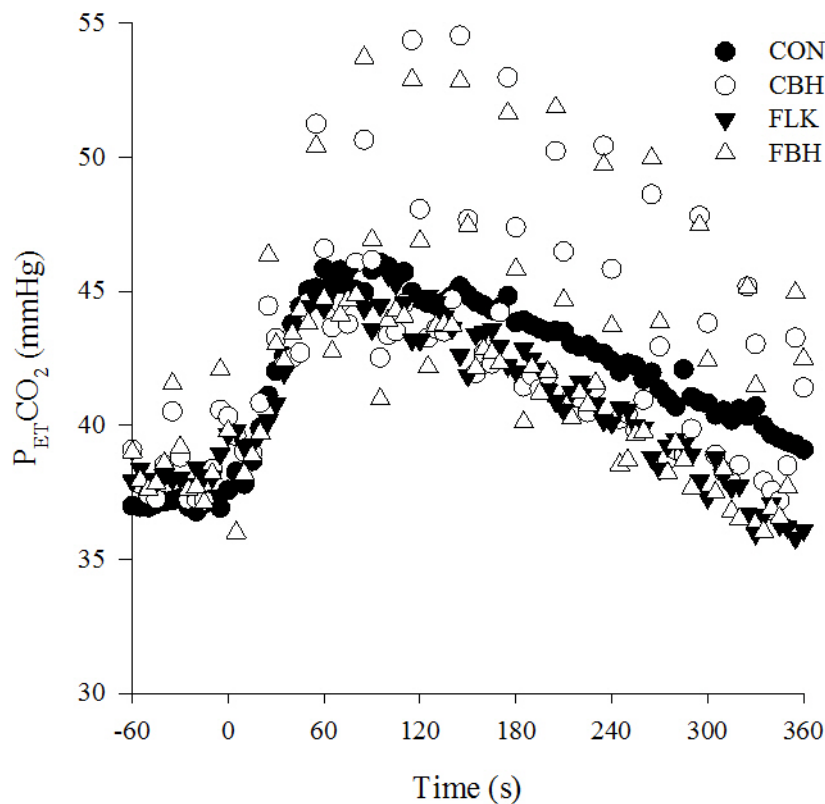


Figure 3a. Mean end-tidal partial pressure of carbon dioxide ($P_{ET}CO_2$) during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2peak}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The $P_{ET}CO_2$ from 120 s to the end of exercise was lower in FLK than CON, and greater in FBH than FLK ($p < 0.05$).

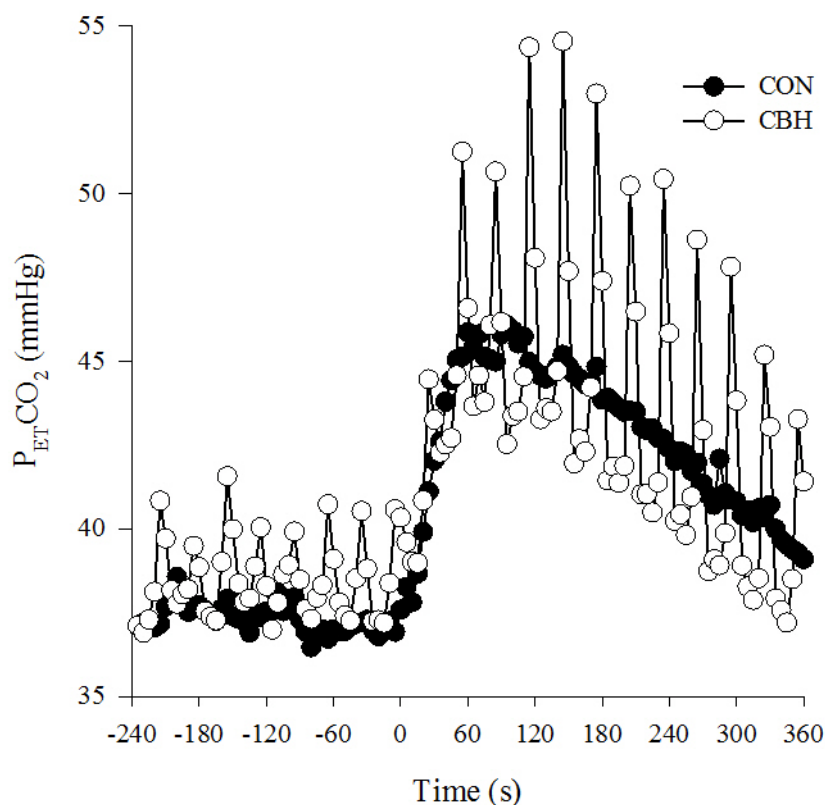


Figure 3b. Fluctuations in mean end-tidal partial pressure of carbon dioxide ($P_{ET}CO_2$) during CON and CBH. This reflects the acute resolution of transient hypoxia associated with each breath hold episode. The $P_{ET}CO_2$ during the last 5 s (highest oscillation point shown in the figure) were greater than the nadir values during the 25 s periods over the 30 s cycles ($p < 0.05$).

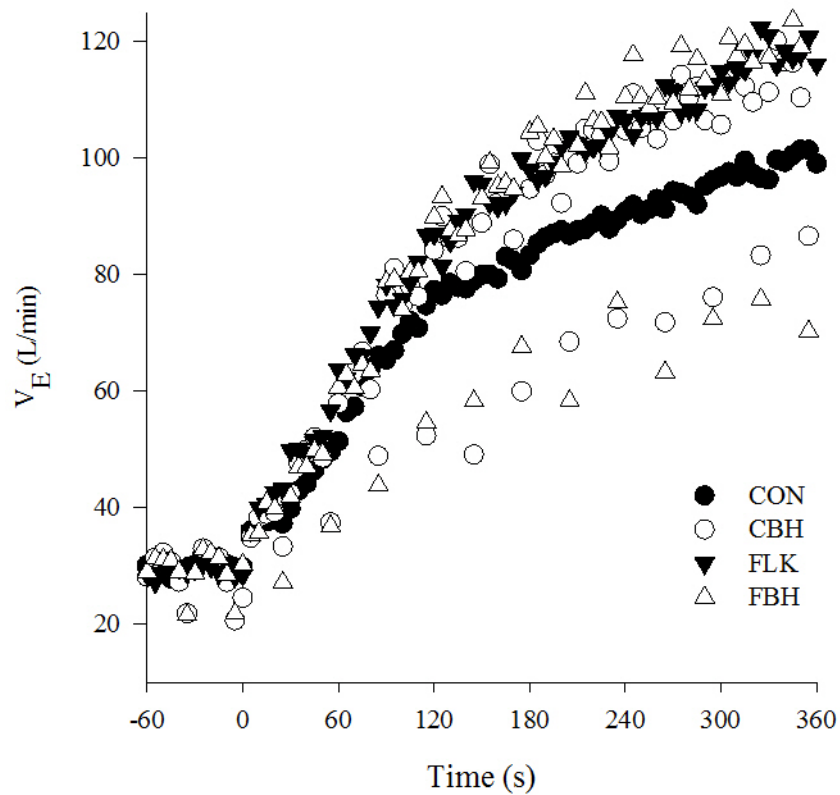


Figure 4. Mean minute ventilation (\dot{V}_E) during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2\text{peak}}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The \dot{V}_E from 120 s to the end of exercise was greater in FLK and CBH than CON ($p < 0.05$).

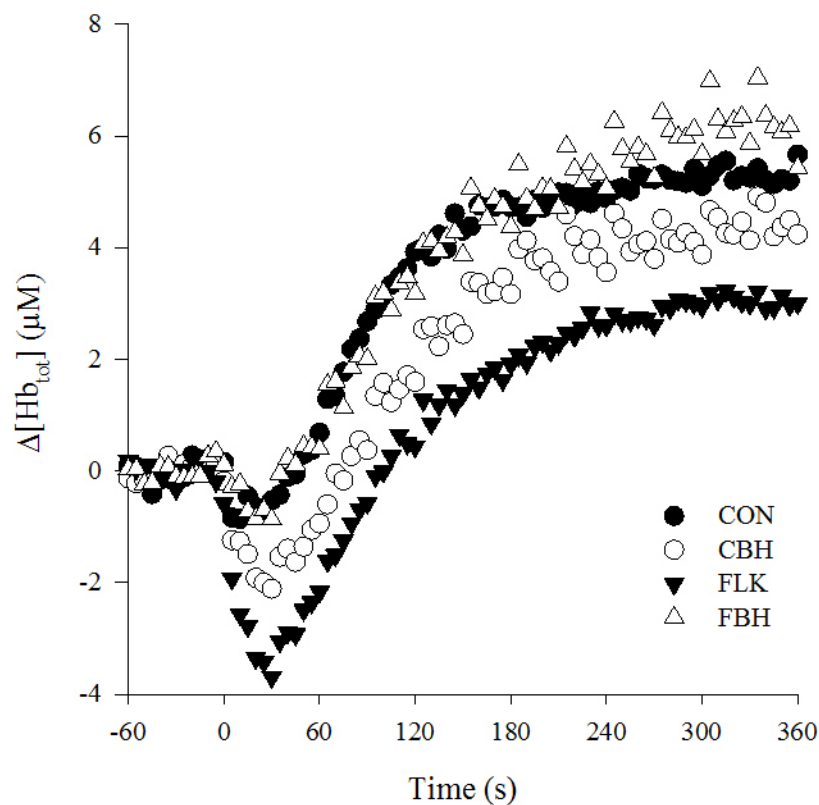


Figure 5a. Mean total hemoglobin concentration changes ($\Delta[\text{Hb}_{\text{tot}}]$) from baseline values during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}\text{O}_{2\text{peak}}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free breathing sprints - peak work rate attained during ramp incremental test), and FBH (repeated 25 s free breathing $\Delta 50$ and 5 s sprints with breath holds). The mean $\Delta[\text{Hb}_{\text{tot}}]$ from 120 s to the end of exercise during CBH and FLK were lower than CON and FBH ($p < 0.05$).

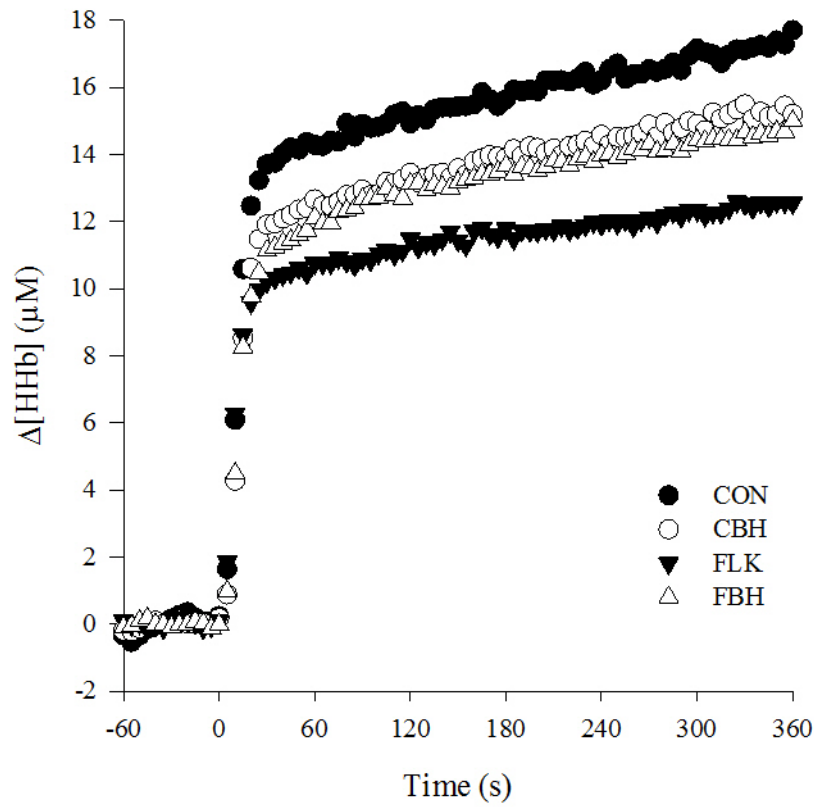


Figure 5b. Mean deoxygenated hemoglobin concentration changes ($\Delta[\text{HHb}]$) from baseline values during CON, CBH, FLK, and FBH. The mean $\Delta[\text{HHb}]$ from 120 s to the end of exercise were different in all conditions ($p < 0.05$).

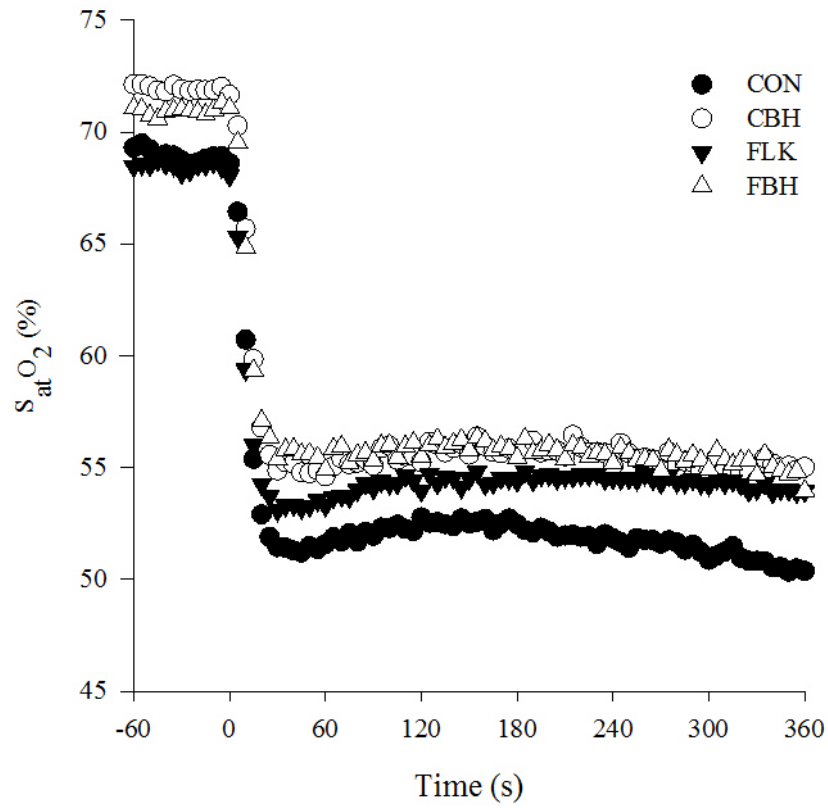


Figure 5c. Mean tissue hemoglobin saturation (S_{atO_2}) during CON, CBH, FLK, and FBH. The mean S_{atO_2} from 120 s to the end of exercise during CBH and FBH were greater than CON ($p < 0.05$).

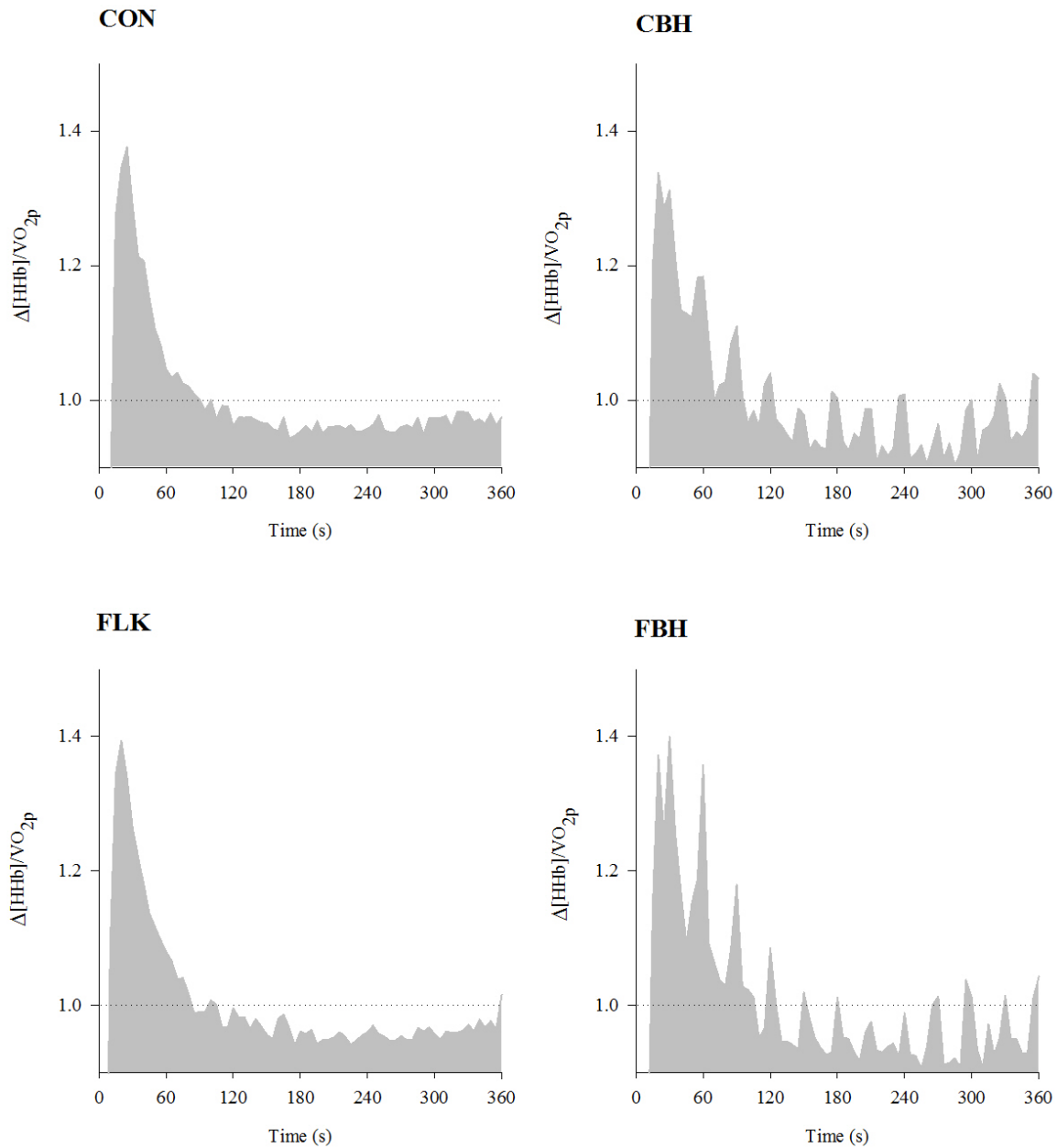


Figure 6. Mean adjustment of normalized [HHb]-to- $\dot{V}O_{2p}$ ratio ($\Delta[\text{HHb}]/\dot{V}O_{2p}$) during CON (continuous exercise at $\Delta 50$: 50 percent of the difference between the LT and $\dot{V}O_{2\text{peak}}$), CBH (continuous exercise at $\Delta 50$ with repeated 25 s free-breathing and 5 s breath holds), FLK (repeated 25 s $\Delta 50$ and 5 s free-breathing sprints - peak power output attained during ramp incremental test), and FBH (repeated 25 s free-breathing $\Delta 50$ and 5 s sprints with breath holds). $\Delta[\text{HHb}]/\dot{V}O_{2p}$ during the last 5 s were greater than the nadir values during the 25 s periods over the 30 s cycles ($p < 0.05$).

2.4 Discussion

The novel purpose of this study was to compare and contrast mean $\dot{V}O_{2p}$, $\dot{V}O_{2p}$ kinetics, muscle deoxygenation ($\Delta[\text{HHb}]$), heart rate, and arterialized-capillary lactate concentrations ($[\text{La}^-]$) during heavy intensity constant-load cycle ergometer exercise (CON), to three different intermittent, heavy intensity exercise protocols. These protocols included 5 s periods of breath holding (CBH), 5 s periods of higher power output (FLK), or a combination of the two (FBH).

The main findings demonstrated that, compared to CON: 1) CBH resulted in similar mean $\dot{V}O_{2p}$ and $\dot{V}O_{2p}$ kinetics, increased mean \dot{V}_E , decreased $\Delta[\text{HHb}]$, similar $[\text{La}^-]$, 2) FLK resulted in increased mean $\dot{V}O_{2p}$, similar $\dot{V}O_{2p}$ kinetics, increased \dot{V}_E , decreased $\Delta[\text{HHb}]$, and similar $[\text{La}^-]$, and 3) FBH resulted in similar mean $\dot{V}O_{2p}$, slower $\dot{V}O_{2p}$ kinetics, increased \dot{V}_E , decreased $\Delta[\text{HHb}]$, and increased $[\text{La}^-]$.

Continuous Breath Hold (CBH).

Before discussing the outcomes of the addition of breath holds to continuous heavy intensity exercise (CBH), the effects of breath holds in the present study on pulmonary gas pressures, arterial gas pressures, and the subsequent effects on O_2 delivery will be discussed. Initially, the breath holds resulted in decreased $P_{\text{ET}}\text{O}_2$ and increased $P_{\text{ET}}\text{CO}_2$ (Table 2; Figure 2b and 3b). Under sub-lactate threshold exercise intensities, end-tidal partial pressures have been shown to be in equilibrium with that of arterial partial pressures (78). However, during supra-lactate threshold intensities, such as were utilized in the present study, $P_{\text{ET}}\text{O}_2$ and $P_{\text{ET}}\text{CO}_2$ are greater and lower than pulmonary capillary pressures, respectively (86). This response is linked to the increased breathing frequencies associated with the ventilatory buffering that occurs at these intensities, and the associated faster breathing frequencies that reduce alveolar-capillary

diffusion times for both gases, and as such, may not be an accurate reflection of pulmonary capillary gas partial pressures (86, 88). However, in the present study, the $P_{ET}O_2$ and $P_{ET}CO_2$ of CBH and FBH are determined utilizing the partial pressure of these gases from the last inspiration before the breath hold, and the expiration 5 s later. Consequently, the confounding effects of the faster breathing frequencies are not an issue, and it is suggested that the fluctuations in $P_{ET}O_2$ and $P_{ET}CO_2$ consequent to the breath holds (Table 3; Figure 2b and 3b) reflect a distinct hypoxemia and hypercapnia within the pulmonary capillaries (90). It is also suggested that pursuant to hypoxemia and hypercapnia, the hyperventilatory response (Table 2 and 3; Figure 4), presumably via the peripheral and central chemoreceptors (88), was observed. Together, the unchanged mean $\dot{V}O_{2p}$ during CBH, compared to CON (Table 2; Figure 1), suggests that the momentary, breath hold-induced hypoxemia and hypercapnia has been resolved by the hyperventilatory responses to the breath holds (Table 3; Figure 4), reflected by the increased mean \dot{V}_E throughout the duration of CBH (Table 2; Figure 4).

Previous literature has shown that increases or decreases in NIRS-derived muscle deoxygenation ($\Delta[HHb]$) and tissue hemoglobin saturation ($S_{at}O_2$) reflect increases or decreases in the PO_2 . This reflects O_2 utilization from oxidative phosphorylation, and changes in O_2 delivery under constant $\dot{V}O_{2p}$ conditions within the microvasculature in the muscle under NIRS interrogation (6). CBH, compared to CON, resulted in decreased heart rate, increased O_2 -Pulse, and a decrease in total hemoglobin concentration ($\Delta[Hb_{tot}]$) (Table 2; Figure 5a). This suggests that any decreases in heart rate was made up by increases in stroke volume as reflected with increased O_2 -Pulse, and thus O_2 delivery was maintained and local muscle blood flow decreased, similar to decreased leg blood flow observed by others under similar hypoxemic conditions (37). However, the decreased $\Delta[HHb]$ and increased $S_{at}O_2$ in CBH, compared to CON (Table 2;

Figure 5b and 5c) suggests, despite decreased local blood flow, a dramatic enhancement of microvascular blood flow redistribution has occurred in the muscle under NIRS interrogation (15, 52). Effectively, doing more with less. The appropriate temporal response of the microvascular endothelium facilitating this redistribution has been demonstrated in previous animal work (51). These authors observed a $3\% \cdot s^{-1}$ increase in vessel radius, after a time delay of ~ 4 s from the endothelial vasodilatory stimulus (51). It is suggested that the ~ 10 s of hypoxemia and hypercapnia associated with the 5 s breath holds observed at the lung (Table 3; Figure 2b and 3b) would also be present at the muscle for a similar duration, which would have been of sufficient duration for the targeted vasodilatory response to increase microvascular blood flow distribution in the muscle under NIRS interrogation.

Furthermore, increases in $S_{at}O_2$ and O_2 -Pulse under hypoxic and hypercapnic conditions have also been observed at rest (74) and during moderate intensity exercise (33). These investigators suggested that increased breathing frequencies and \dot{V}_E , similar to that observed during CBH, would have elicited a decrease in alveolar gas, and subsequently, blood temperature. This would have shifted the O_2 dissociation curve to the left, increasing the $S_{at}O_2$ at a given PO_2 (47). While it is possible that the increase in $S_{at}O_2$ and O_2 -Pulse observed in CBH reflects a similar change in blood temperature, our increased $\Delta[Hb_{tot}]$, $\Delta[HHb]$, and $S_{at}O_2$ data suggest the aforementioned enhancement in microvascular blood flow distribution in the muscle under NIRS interrogation.

Conventionally, increases and decreases in the adjustment of the $\Delta[HHb]$ -to- $\dot{V}O_{2p}$ ratio ($\Delta[HHb]/\dot{V}O_{2p}$) have been utilized to reflect increases and decreases in muscle arterial-venous O_2 differences and/or O_2 delivery as per the Fick equation (32, 55). Within the 30 s cycles of CBH, $\Delta[HHb]/\dot{V}O_{2p}$ oscillations were observed (Table 3; Figure 6), yet no fluctuations in $\Delta[HHb]$ were

detected (Table 3). This suggests that the $\Delta[\text{HHb}]/\dot{V}\text{O}_{2p}$ oscillations during CBH (Table 3; Figure 6) were due to fluctuations in $\dot{V}\text{O}_{2p}$ (Table 3; Figure 1). It appears that the transient 5 s breath hold-induced reductions of $P_{\text{ET}}\text{O}_2$ and the subsequent hypoxemia reduced the O_2 delivery to the exercising muscle, resulting in a short-lived decrease in oxidative phosphorylation. This elicited both the observed rhythmical decreases of $\dot{V}\text{O}_{2p}$ (Table 3; Figure 1) and the synchronous overshoots in $\Delta[\text{HHb}]/\dot{V}\text{O}_2$ (Table 3; Figure 6). Notably, similar temporal oscillations in $\dot{V}\text{O}_{2p}$ have been observed in previous intermittent exercise protocols during 5 s active recovery periods, which reduced O_2 demand (7). Taken together, this suggests that, with either a 5 s period of reduced O_2 demand or O_2 availability, a concomitant reduction in oxidative phosphorylation results.

CBH, compared to CON, also resulted in bradycardia (Table 2). Similar breath hold-induced bradycardia has been observed in previous breath hold research (1, 38, 42). Mechanistically, it has been suggested that a breath hold performed after an inspiration increases the intrathoracic pressures and decreases the abdominal pressures (28, 60, 75), creating a pressure gradient between the infra- and supra-diaphragmatic portions of the inferior vena cava, that draws blood towards the right side of the heart (28, 60, 75). This facilitates venous return and diastole of the right side of the heart, thus increasing stroke volume concurring with Starling's law (10). It is suggested that this mechanism was responsible for the observed bradycardia during the breath holds in CBH (Table 2).

The previously observed slowing of $\dot{V}\text{O}_{2p}$ kinetics during exercise in hypoxia (15, 17, 30, 76) and longer duration breath holds (28, 38, 70), suggested that breath holds in CBH would result in a similar slowing of $\dot{V}\text{O}_{2p}$ kinetics, compared to CON. However, the expectation was that the breath hold-induced hypoxemia and the decrease in oxidative phosphorylation would be

replaced by increased substrate level phosphorylation (15, 17, 24, 30, 57, 76). As such, despite increased $\dot{V}CO_{2p}$ and \dot{V}_E , the similar $[La^-]$ in CBH, compared to CON (Table 2) suggests that the expected metabolic stress associated with increased substrate level phosphorylation was resolved. It appears that increased \dot{V}_E during CBH (Table 2; Figure 4) reconciled the transient hypoxemia and hypercapnia through increased CO_2 elimination during the 25 s free-breathing periods (Table 3). Moreover, it is suggested that the improvement in microvascular distribution of blood flow in CBH, combined with the increased \dot{V}_E , have also averted any slowing of $\dot{V}O_{2p}$ kinetics and increase in $[La^-]$.

Fartlek (FLK).

Our previous work on intermittent exercise patterns have shown that regular insertions of short recovery periods (5 s and 3 s) during heavy intensity cycling exercise resulted in decreased mean $\dot{V}O_{2p}$ and $\Delta[HHb]$ (7, 49). It was expected in the present study that, if these short recovery periods were replaced with periods of higher power outputs, the increased ATP demand would result in increased mean $\dot{V}O_{2p}$, $\dot{V}CO_{2p}$, \dot{V}_E , $\Delta[HHb]$, and $[La^-]$, compared to CON.

Indeed, the insertions of 5 s periods of higher power outputs (sprints), which increased the mean power output, was resulted in increased mean $\dot{V}O_{2p}$, $\dot{V}CO_{2p}$, and \dot{V}_E , and, unexpectedly, similar $[La^-]$, compared to CON (Table 2; Figure 1 and 4). Utilizing the standard O_2 cost of work ($\sim 10 \text{ ml } O_2 \cdot \text{min}^{-1} \cdot \text{W}^{-1}$) (63), our data have shown that the increased ATP demand from the increased mean power output from CON to FLK has been ultimately met by increased mean $\dot{V}O_{2p}$ (Table 2; Figure 1). However, previous work from our lab has demonstrated that during the first 4 s of work, following brief recovery periods from an elevated baseline during intermittent exercise, has shown that contributions from PCr-derived

phosphorylation and substrate level phosphorylation were substantial (8). Similarly, in the present study, the repeated transitions from 218 W during the 25 s period, to 5 s sprints at 314 W (Table 1), would have increased PCr-derived phosphorylation and substrate level phosphorylation and resulted in increased type II fibre recruitment (24, 57), intracellular and blood $[La^-]$ (4, 43). Unexpectedly the post-exercise $[La^-]$ in FLK was similar to CON (Table 2). Despite the accumulation of La^- and H^+ that would have developed from the 5 s sprints, how was $[La^-]$ in FLK, similar to CON? It is suggested that the similar $[La^-]$ was mechanistically possible through increased \dot{V}_E , reflective of increased ventilatory buffering (Table 2 and 3; Figure 4) of the additional CO_2 that had been produced via the carbonic anhydrase reaction (61, 86).

FLK, compared to CON, also resulted in increased mean heart rate and similar O_2 -Pulse (Table 2), reflecting an increase in O_2 delivery. This increase in mean heart rate and the expected increase in systolic blood pressure associated with the 5 s sprints, may have resulted from a central attempt, via increased baroreceptor-mediated sympathetic activation (69). Furthermore, the decreased $\Delta[Hb_{tot}]$ and $\Delta[HHb]$, and increased $S_{at}O_2$, suggests that the increase in O_2 delivery was greater than the increased O_2 demand associated with FLK (Table 2; Figure 5a, 5b, and 5c). Thus, it appears that the increase in O_2 delivery as suggested from increased mean heart rate and O_2 -Pulse was proportionally greater than what was required, and a maldistribution of microvascular blood flow in the muscle of NIRS interrogation occurred, resulting in the decreased muscle deoxygenation (Table 2; Figure 5b).

Previous intermittent exercise research has also observed a similar maldistribution of microvascular blood flow during muscle contractions performed at greater, compared to lower work rates (68, 81). This was linked to increased intramuscular pressures (64) which impeded blood flow in the exercising muscles (2, 21, 44). As such, it is suggested that the 44% increase in

power output, performed during the 5 s sprints of FLK (Table 2), would have similarly impeded blood flow and elicited the observed decrease in $\Delta[\text{Hb}_{\text{tot}}]$. Thus, the increase in mean heart rate during FLK may have resulted from a central attempt to improve local muscle blood flow and overcome the aforementioned occlusion-mediated increase in vascular resistance (68).

FLK also resulted in similar $\dot{V}\text{O}_{2\text{p}}$ kinetics to CON (Table 2). The previously suggested mechanisms by which $\dot{V}\text{O}_{2\text{p}}$ kinetics may have been slowed (30, 43) are suggested to have been offset by increased mean $\dot{V}\text{O}_{2\text{p}}$ and O_2 delivery, compared to CON (Table 2; Figure 1). It appears that the increased mean \dot{V}_E and pulmonary-arterial PO_2 , as suggested by the greater $\text{P}_{\text{ET}}\text{O}_2$ in FLK (Table 2; Figure 2a), increased the alveolar-arterial PO_2 gradient, and thus, O_2 delivery (61).

Fartlek Breath Hold (FBH).

In FBH, it was expected that the combined responses to the insertions of 5 s breath holds and sprints would overwhelm any hyperventilatory attempts to increase $\dot{V}\text{O}_{2\text{p}}$ and attenuate increases in $[\text{La}^-]$, that was observed in the singular interventions in CBH and FLK (83).

Consequent to the insertions of 5 s sprints, FBH resulted in greater mean power output and ATP demand, as well as increased $\dot{V}\text{CO}_{2\text{p}}$ and mean \dot{V}_E , compared to CON (Table 2; Figure 4). In comparison to FLK, FBH resulted in lower $\dot{V}\text{CO}_{2\text{p}}$ and similar mean \dot{V}_E (Table 2; Figure 4). This suggests that the increase in mean \dot{V}_E during FLK was comparable to that of the breath hold-induced hyperventilatory responses that was observed during the 25 s free-breathing periods of FBH (Table 3; Figure 4), allowing sufficient ventilation to result in similar and lower mean $\dot{V}\text{O}_{2\text{p}}$, compared to CON and FLK, respectively (89). Nevertheless, the increased $[\text{La}^-]$ in FBH, compared to all other conditions (Table 2) suggests that the ventilatory responses in FBH

were overwhelmed by the combination of the breath holds and sprints, and was insufficient to resolve the increased La^- production from the sprints.

Moreover, FBH resulted in greater heart rate than CON, and lower heart rate than FLK, but lower O_2 -Pulse compared to both CON and FLK, suggesting increased O_2 delivery (Table 2). As suggested earlier, the increased sympathetic activation (69) may be responsible for the increased heart rate in FBH compared to CON, and the breath hold-induced bradycardia (60, 75), for the decreased heart rate in FBH relative to FLK.

FBH also resulted in increased $\Delta[\text{Hb}_{\text{tot}}]$, compared to both CON and FLK, decreased and increased $\Delta[\text{HHb}]$, compared to CON and FLK, respectively, and increased $\text{S}_{\text{at}}\text{O}_2$, compared to both CON and FLK (Table 2; Figure 5a, 5b, and 5c). These responses suggest that the addition of the breath holds to the sprints increased local muscle blood flow, presumably consequent to the local vasodilatory effects of La^- (27, 40). It has been observed previously that increased $[\text{La}^-]$ and associated acidosis induces local vasodilation through activation of cyclic guanine monophosphate (cGMP) within smooth muscle cells of the muscle microvasculature (12).

Furthermore, in comparison to CON, the increased heart rate, $\Delta[\text{Hb}_{\text{tot}}]$, and decreased O_2 -Pulse during FBH, (Table 2; Figure 5a), suggest an increase in blood velocity that would have decreased Hb capillary transit and diffusion time within the microvasculature. This would have resulted in a reduction of O_2 availability to the working muscle, and thus, resulted in the observed decrease in muscle deoxygenation (Table 2; Figure 5b). In comparison to FLK, in which the blood velocity is relatively slower due to breath hold-induced bradycardia, the smaller decrease in Hb capillary transit and diffusion time within the microvasculature, has resulted in increased muscle deoxygenation (Table 2; Figure 5b). Similar effects of Hb capillary and diffusion time on muscle deoxygenation have been suggested elsewhere under similar

hypoxemia during exercise in hypoxia (15). Despite the increased power output and ATP demand of FBH, compared to CON, the unchanged mean $\dot{V}O_{2p}$ during FBH (Table 2; Figure 1) suggests a similar reduction in O_2 availability for oxidative phosphorylation.

FBH also resulted in slower $\dot{V}O_{2p}$ kinetics than both CON and FLK (Table 2), consistent with previous observations of slower $\dot{V}O_{2p}$ kinetics during exercise in hypoxia (15, 17, 30, 76), and the associated increased type II muscle fibre recruitment (24, 57) and blood $[La^-]$ (56).

In summary, the metabolic stress resulting from the addition of sprints to the breath holds were unable to be resolved by the ventilatory responses, as was observed in CBH, notably resulting in decreased mean $\dot{V}O_{2p}$, slower $\dot{V}O_{2p}$ kinetics, and increased $[La^-]$ (Table 2; Figure 1).

Practical Applications

In swimming, athletes utilize different strategies of achieving the ~5 s underwater kicking phase. In backstroke swimming, swimmers tend to be faster kicking underwater and therefore may perform these underwater breath hold phases at greater intensity to maximize their velocity over these 5 s. The present study provides preliminary insight into the singular and combined physiological resolutions associated with 30 s cycles, comprised of 5 s breath holds and/or sprints, followed by 25 s of free-breathing performed at an identical power output. Our data suggest that choosing to perform such breath hold phases at greater intensity places a greater demand on substrate level phosphorylation and will precipitate an earlier onset of fatigue.

Conclusion

To our knowledge, this is the first study to examine the physiological responses to the insertions of intermittent 5 s breath holds and/or sprints during heavy intensity cycling exercise.

It has been established that the breath hold-induced hyperventilatory responses and improved microvascular distribution of blood flow during CBH, were sufficient to maintain similar $\dot{V}O_{2p}$ to CON. However, when breath holds were combined with the sprints, the hyperventilatory responses were overwhelmed and precipitated greater metabolic stress.

2.5 Future Directions and Limitations

Future Directions

The present study on the physiological responses to the breath holds during heavy intensity exercise were observed among but limited to recreationally active, healthy young male participants. Investigation of similar protocols in women and/or older populations would enable comparisons between sex and with aging, enabling to further characterize the physiological responses to breath holds and sprints during heavy intensity exercise.

Limitations

The NIRS signals are affected by subcutaneous adipose tissue thickness (5). The penetration depth is roughly half of the optode distance (19). Since our measures were not corrected for the adipose tissue thickness, subject variabilities in adipose tissue thickness may have influenced the NIRS signals.

2.6 References

1. **Andersson JP, Linér MH, Fredsted A, Schagatay EK.** Cardiovascular and respiratory responses to apneas with and without face immersion in exercising humans. *J Appl Physiol* 96: 1005–1010, 2004.
2. **Barcroft BYH, Dornhorst AC.** The blood flow through the human calf during rhythmic exercise. *J Physiol* 109: 402–411, 1949.
3. **Barstow TJ, Mole PA.** Linear and nonlinear characteristics of oxygen uptake kinetics during heavy exercise. *J Appl Physiol* 71: 2099–2106, 1991.
4. **van Beekvelt MC, Borghuis MS, van Engelen BG, Wevers RA, Colier WN.** Adipose tissue thickness affects in vivo quantitative near-IR spectroscopy in human skeletal muscle. *Clin Sci (Lond)* 101: 21–28, 2001.
5. **Behnke BJ, Kindig CA, Musch TI, Koga S, Poole DC.** Dynamics of microvascular oxygen pressure across the rest-exercise transition in rat skeletal muscle. *Respir Physiol* 126: 53–63, 2001.
6. **Belfry GR, Paterson DH, Murias JM, Thomas SG.** The Effects of Short Recovery duration on VO₂ and muscle deoxygenation during intermittent exercise. *Eur J Appl Physiol* 112: 1907–1915, 2012.
7. **Belfry GR, Raymer GH, Marsh GD, Paterson DH, Thompson RT, Thomas SG.** Muscle metabolic status and acid-base balance during 10-s work : 5-s recovery intermittent and continuous exercise. *J Appl Physiol* 113: 410–417, 2012.

8. **Da Boit M, Bailey SJ, Callow S, Dimenna FJ, Jones AM.** Effects of interval and continuous training on O₂ uptake kinetics during severe-intensity exercise initiated from an elevated metabolic baseline. *J Appl Physiol* 116: 1068–77, 2014.
9. **Braunwald E, Frahm CJ.** Studies on Starling's Law of the Heart: IV. Observations on the Hemodynamic Functions of the Left Atrium in Man. *Circulation* 24: 633–642, 1961.
10. **Chen Y, Messina J, Wolin S.** Evidence arteriolar for cGMP mediation dilation to lactate of skeletal muscle. *J Appl Physiol* 81: 349–354, 1996.
11. **DeLorey DS, Shaw CN, Shoemaker JK, Kowalchuk JM, Paterson DH.** The effect of hypoxia on pulmonary O₂ uptake, leg blood flow and muscle deoxygenation during single-leg knee-extension exercise. *ExpPhysiol* 89: 293–302, 2004.
12. **Dicker SG, Lofthus GK, Thornton NW, Brooks G.** Respiratory and heart rate responses to tethered controlled frequency breathing swimming. *Med Sci Sports Exerc* 12: 20–23, 1980.
13. **Engelen M, Porszasz J, Riley M, Wasserman K, Maehara K, Barstow TJ.** Effects of hypoxic hypoxia on O₂ uptake and heart rate kinetics during heavy exercise. *J Appl Physiol* 81: 2500–2508, 1996.
14. **Ferrari M, Mottola L, Quaresima V.** Principles, techniques, and limitations of near infrared spectroscopy. *Can J Appl Physiol* 29: 463–87, 2004.
15. **Folkow B, Gaskell P, Waaler BA.** Blood Flow through Limb Muscles during Heavy Rhythmic Exercise. *Acta Physiol Scand* 80: 61–72, 1970.

16. **Gollnick PD, Sjödín B, Karlsson J, Jansson E, Saltin B.** Human soleus muscle: A comparison of fiber composition and enzyme activities with other leg muscles. *Pflügers Arch Eur J Physiol* 348: 247–255, 1974.
17. **Hill A V., Long CNH, Lupton H.** Muscular Exercise, Lactic Acid, and the Supply and Utilisation of Oxygen. *Ergebnisse der Physiol* 24: 43–51, 1925.
18. **Hoffmann U, Smerecnik M, Leyk D, Essfeld O.** Cardiovascular responses to apnea during dynamic exercise. *Int J Sports Med* 26: 426–431, 2005.
19. **Hughson RL, Kowalchuk JM.** Kinetics of oxygen uptake for submaximal exercise in hyperoxia, normoxia, and hypoxia. *Can J Appl Physiol* 20: 198–210, 1995.
20. **Karpman VL.** The theoretical analysis of Fick's equation. On the centennial of the use of Fick's principle in physiology. *Z Kardiol* 64: 801–808, 1975.
<http://www.ncbi.nlm.nih.gov/pubmed/769376>.
21. **Katayama K, Sato Y, Ishida K, Mori S, Miyamura M.** The effects of intermittent exposure to hypoxia during endurance exercise training on the ventilatory responses to hypoxia and hypercapnia in humans. *Eur J Appl Physiol Occup Physiol* 78: 189–94, 1998.
22. **Keir DA, Murias JM, Paterson DH, Kowalchuk JM.** Breath-by-breath pulmonary O₂ uptake kinetics: effect of data processing on confidence in estimating model parameters. *Exp Physiol* 99: 1511–22, 2014.
23. **Koskolou MD, Calbet JAL, Rådegran G, Roach RC.** Hypoxia and the cardiovascular response to dynamic knee-extensor exercise. *Am J Physiol* 272: H2655–H2663, 1997.

24. **Kume D, Akahoshi S, Song J, Yamagata T, Wakimoto T, Nagao M, Matsueda S, Nagao N.** Intermittent breath holding during moderate bicycle exercise provokes consistent changes in muscle oxygenation and greater blood lactate response. *J Sports Med Phys Fitness* 53: 327–335, 2013.
25. **Lamarra N, Whipp BJ, Ward SA, Wasserman K.** Effect of interbreath fluctuations on characterizing exercise gas exchange kinetics. *J Appl Physiol* 62: 2003–12, 1987.
<http://www.ncbi.nlm.nih.gov/pubmed/3110126>.
26. **Laughlin MH, Armstrong RB.** Muscle blood flow during locomotory exercise. *Exerc Sport Sci Rev* 13: 95–136, 2003.
27. **Lindholm P, Linnarsson D.** Pulmonary gas exchange during apnoea in exercising men. *Eur J Appl Physiol* 86: 487–491, 2002.
28. **Lindholm P, Sundblad P, Linnarsson D.** Oxygen-conserving effects of apnea in exercising men. *J Appl Physiol* 87: 2122–2127, 1999.
29. **Linnarsson D, Karlsson J, Fagraeus L, Saltin B.** Muscle metabolites with exercise and oxygen deficit and hyperoxia. *J Appl Physiol* 36: 399–402, 1974.
30. **Lutjemeier BJJ, Miura AA, Scheuermann BWB, Koga SS, Townsend DKDK, Barstow TJJ.** Muscle contraction-blood flow interactions during upright knee extension exercise in humans. *J Appl Physiol* 98: 1575–1583, 2005.
31. **Manning JM.** Physiology of Sport and Exercise. *J Athl Train* 34: 298–299, 1999.
32. **Matheson GO, McKenzie DC.** Breath holding during intense exercise: arterial blood

- gases, pH, and lactate. *J Appl Physiol* 64: 1947–52, 1988.
33. **McCrudden M, Keir DA, Belfry G.** The effects of short work versus longer work periods within intermittent exercise on VO₂p kinetics, muscle deoxygenation and energy system contribution. *J Appl Physiol* 122: 1435–1444, 2017.
 34. **Murias JM, Dey A, Campos OA, Estaki M, Hall KE, Melling CWJ, Noble EG.** High-Intensity Endurance Training Results in Faster Vessel-Specific Rate of Vasorelaxation in Type 1 Diabetic Rats. *PLoS One* 8: e59678, 2013.
 35. **Murias JM, Keir DA, Spencer MD, Paterson DH.** Sex-related differences in muscle deoxygenation during ramp incremental exercise. *Respir Physiol Neurobiol* 189: 530–536, 2013.
 36. **Murias JM, Kowalchuk JM, Paterson DH.** Speeding of VO₂ kinetics in response to endurance-training in older and young women. *Eur J Appl Physiol* 111: 235–243, 2011.
 37. **Murias JM, Spencer MD, Paterson DH.** The Critical Role of O₂ Provision in the Dynamic Adjustment of Oxidative Phosphorylation. *Exerc Sport Sci Rev* 42: 4–11, 2014.
 38. **Murphy PC, Cuervo LA, Hughson RL.** A study of cardiorespiratory dynamics with step and ramp exercise tests in normoxia and hypoxia. *Cardiovasc Res* 23: 825–832, 1989.
 39. **Nuutinen EM, Nishiki K, Erecinska M, Wilson DF.** Role of mitochondrial oxidative phosphorylation in regulation of coronary blood flow. *Am J Physiol - Hear Circ Physiol* 243: H159–H169, 1982.
 40. **Ozyener F, Rossiter HB, Ward S a, Whipp BJ.** Influence of exercise intensity on the

- on- and off-transient kinetics of pulmonary oxygen uptake in humans. *J Physiol* 533: 891–902, 2001.
41. **Paterson DJ, Wood GA, Morton AR, Henstridge JD.** The entrainment of ventilation frequency to exercise rhythm. *Eur J Appl Physiol Occup Physiol* 55: 530–537, 1986.
 42. **Paulev P -E.** Cardiac Rhythm during Breath-Holding and Water Immersion in Man. *Acta Physiol Scand* 73: 139–150, 1968.
 43. **Peronnet F, Aguilaniu B.** Lactic acid buffering, nonmetabolic CO₂ and exercise hyperventilation: A critical reappraisal. *Respir Physiol Neurobiol* 150: 4–18, 2006.
 44. **Poole DC, Gaesser GA, Hogan MC, Knight DR, Wagner PD.** Pulmonary and leg VO₂ during submaximal exercise: implications for muscular efficiency. *J Appl Physiol* 72: 805–810, 1992.
http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?cmd=Retrieve&db=PubMed&dopt=Citation&list_uids=1559962.
 45. **Rådegran G, Saltin B.** Muscle blood flow at onset of dynamic exercise in humans. *Am J Physiol - Hear Circ Physiol* 274: H314–H322, 1998.
<http://ajpheart.physiology.org/content/274/1/H314.abstract>.
 46. **Rossiter HB, Ward SA, Doyle VL, Howe FA, Griffiths JR, Whipp BJ.** Inferences from pulmonary O₂ uptake with respect to intramuscular [phosphocreatine] kinetics during moderate exercise in humans. *J Physiol* 518: 921–932, 1999.
 47. **Saltin B, Rådegran G, Koskolou MD, Roach RC.** Skeletal muscle blood flow in

- humans and its regulation during exercise. *Acta Physiol Scand* 162: 421–436, 1998.
48. **Saul JP, Rea RF, Eckberg DL, Berger RD, Cohen RJ.** Heart rate and muscle sympathetic nerve variability during reflex changes of autonomic activity. *Am J Physiol* 258: H713-21, 1990.
49. **Schagatay E, Andersson JPA, Nielsen B.** Hematological response and diving response during apnea and apnea with face immersion. *Eur J Appl Physiol* 101: 125–132, 2007.
50. **Siegmund GP, Edwards MR, Moore KS, Tiessen DA, Sanderson DJ, McKenzie DC.** Ventilation and locomotion coupling in varsity male rowers. *J Appl Physiol* 87: 233–242, 1999. <http://jap.physiology.org/content/87/1/233.short>.
51. **Skorski S, Faude O, Caviezel S, Meyer T.** Reproducibility of Competition Pacing Profiles in Elite Swimmers. *Int J Sports Physiol Perform* 9: 217–225, 2013.
52. **Somers VK, Mark a L, Zavala DC, Abboud FM.** Contrasting effects of hypoxia and hypercapnia on ventilation and sympathetic activity in humans. *J Appl Physiol* 67: 2101–2106, 1989.
53. **Song SH, Lee WK, Chung YA, Hong SK.** Mechanism of apneic bradycardia in man. *J Appl Physiol* 27: 323–327, 1969.
54. **Springer C, Barstow T, Wasserman K, Cooper D.** Oxygen uptake and heart rate responses during hypoxic exercise in children and adults. *Med. Sci. Sports Exerc.* 23: 71–79, 1991. <http://europepmc.org/abstract/MED/1997815>.
55. **Suskind M, Bruce RA, McDowell ME, Yu PNG, Lovejoy FWJ.** Normal variations in

- end-tidal air and arterial blood carbon dioxide and oxygen tensions during moderate exercise. *J Appl Physiol* 3: 282–290, 1950.
56. **Veiga S, Cala A, Mallo J, Navarro E, Arellano R, Terres-Nicoli JM, Redondo JM, Veiga S, Roig A.** Underwater and surface strategies of 200 m world level swimmers. *J Sports Sci* 34: 1–6, 2015.
57. **Walløe L, Wesche J.** Time course and magnitude of blood flow changes in the human quadriceps muscles during and following rhythmic exercise. *J Physiol* 405: 257–73, 1988.
58. **Ward DS, Nguyen TT.** Ventilatory response to sustained hypoxia during exercise. *Med Sci Sports Exerc* 23: 719–726, 1991.
59. **Wasserman K, Van Kessel AL, Burton GG.** Interaction of physiological mechanisms during exercise. *J Appl Physiol* 22: 71–85, 1967.
60. **Wasserman K, Whipp BJ, Castagna J.** Cardiodynamic hyperpnea: hyperpnea secondary to cardiac output increase. *J Appl Physiol* 36: 457–464, 1974.
61. **Wein J, Andersson JP, Erdéus J.** Cardiac and ventilatory responses to apneic exercise. *Eur J Appl Physiol* 100: 637–644, 2007.
62. **Whipp BJ.** The hyperpnea of dynamic muscular exercise. *Exerc Sport Sci Rev* 5: 295–311, 1977.
63. **Whipp BJ, Davis JA.** Peripheral chemoreceptors and exercise hyperpnea. *Med Sci Sports Exerc* 11: 204–212, 1979.
64. **Whipp BJ, Ward SA, Lamarra N, Davis JA, Wasserman K.** Parameters of ventilatory

and gas exchange dynamics during exercise. *J Appl Physiol* 52: 1506–13, 1982.

<http://www.ncbi.nlm.nih.gov/pubmed/6809716>.

65. **Whipp BJ, Wasserman K.** Alveolar-arterial during graded gas tension differences exercise. *J Appl Physiol* 27: 361–365, 1969.
66. **Woorons X, Bourdillon N, Lamberto C, Vandewalle H, Richalet J-P, Mollard P, Pichon A.** Cardiovascular Responses During Hypoventilator at Exercise. *Int J Sports Med* 32: 438–445, 2011.

Appendix

Ethics Approval Notice



**Western
Research**

Research Ethics

**Western University Health Science Research Ethics Board
HSREB Full Board Initial Approval Notice**

Principal Investigator: Glen Belfry
Department & Institution: Health Sciences/Kinesiology, Western University

Review Type: Full Board
HSREB File Number: 107170
Study Title: The Effects of Periodic Breath-Holding During Intermittent Exercise on Energy System Contribution, Lactate Threshold, and Regional Blood Flow Distribution.
Sponsor: Natural Sciences and Engineering Research Council

HSREB Initial Approval Date: January 08, 2016
HSREB Expiry Date: January 08, 2017

Documents Approved and/or Received for Information:

Document Name	Comments	Version Date
Advertisement	Received November 24, 2015	
Other	PAR-Q form - Received December 18, 2015	
Letter of Information & Consent		2015/11/24
Western University Protocol		2015/12/18

The Western University Health Science Research Ethics Board (HSREB) has reviewed and approved the above named study, as of the HSREB Initial Approval Date noted above.

HSREB approval for this study remains valid until the HSREB Expiry Date noted above, conditional to timely submission and acceptance of HSREB Continuing Ethics Review.

The Western University HSREB operates in compliance with the Tri-Council Policy Statement Ethical Conduct for Research Involving Humans (TCPS2), the International Conference on Harmonization of Technical Requirements for Registration of Pharmaceuticals for Human Use Guideline for Good Clinical Practice Practices (ICH E6 R1), the Ontario Personal Health Information Protection Act (PHIPA, 2004), Part 4 of the Natural Health Product Regulations, Health Canada Medical Device Regulations and Part C, Division 5, of the Food and Drug Regulations of Health Canada.

Members of the HSREB who are named as Investigators in research studies do not participate in discussions related to, nor vote on such studies when they are presented to the REB.

The HSREB is registered with the U.S. Department of Health & Human Services under the IRB

This is an official document. Please retain the original in your files

Curriculum Vitae

Name: David John Lim

Post-secondary Education and Degrees:

The University of Western Ontario
London, Ontario, Canada
2017-2021 DDS

The University of Western Ontario
London, Ontario, Canada
2016-2017 M.Sc.

The University of Western Ontario
London, Ontario, Canada
2014-2016 B.A.

The University of Western Ontario
London, Ontario, Canada
2009-2014 B.Sc.

Honours and Awards:

Western Graduate Research Scholarship
2016-2017

UWO In-Course Scholarships Year II
2015

Western's 125th Anniversary Alumni Awards
2015

Laurene Paterson Estate Scholarship
2012

Related Work Experience:

Teaching Assistant
The University of Western Ontario
2016-2017

Publications: