# RNA Extraction from Mouse Tissue



#### Prepare your tissue

Perform the dissection on your mouse models and store your tissues of interest in separate 1.5mL centrifuge tubes at -80 °C. Crush and cut 5 mg of your tissue using liquid nitrogen and 3 freeze - thaw cycles.



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#### **Extracting RNA from tissue**

Add 600 µL of Trizol to your sample. Crush the tissue using a pestle, vortex and leave for 10 min on ice. Centrifuge (14000g, 30sec) and extract 550 µL of mixture to new centrifuge tube. Add 550 µL of ethanol.

#### **Purifying RNA**

Add 5  $\mu$ L of DNA digestion buffer and 5  $\mu$ L of DNase I. Incubate for 15 min at room temperature. Add 150  $\mu$ L of Trizol and 200  $\mu$ L of ethanol to centrifuge tube. Extract 400  $\mu$ L of the mixture to a new spin column and collection tube and centrifuge (14000g, 30 sec). Repeat prewash and wash step.

#### **RNA Sequencing Robarts Research**

Send off the RNA samples to the Robarts Research Institute for RNA sequencing. Perform analysis on



### Isolating RNA in spin column

Add 550 µL of the tissue mixture to a spin column with a collection tube. Centrifuge (14000g, 30sec) and repeat with another 550 µL of mixture. RNA is in the spin column now.

#### Prewash and Wash

Add 400  $\mu$ L of the prewash buffer and centrifuge (14000g, 30 sec, x2). Add 700  $\mu$ L of wash buffer and centrifuge (14000g, 2 min).Place spin column with RNA in a new centrifuge tube. Add 40  $\mu$ L of DNase/RNase free water to the spin column and centrifuge (14000g, 30 sec). RNA in centrifuge tube.

## **Protocol and Reagents**



