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Joan H M Knoll
Brad L Urquhart
Gabriella Mariani

See next page for additional authors

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Genomic signatures for paclitaxel and gemcitabine resistance in breast cancer derived by machine learning

Stephanie N. Dorman\textsuperscript{a}, Katherina Baranova\textsuperscript{a}, Joan H.M. Knoll\textsuperscript{b,c,d}, Brad L. Urquhart\textsuperscript{e}, Gabriella Mariani\textsuperscript{f}, Maria Luisa Carcangiug, Peter K. Rogan\textsuperscript{a,d,h,i,*}

\textsuperscript{a}Department of Biochemistry, Schulich School of Medicine and Dentistry, University of Western Ontario, London, ON, Canada
\textsuperscript{b}Department of Pathology and Laboratory Medicine, Schulich School of Medicine and Dentistry, University of Western Ontario, London, ON, Canada
\textsuperscript{c}Molecular Diagnostics Division, Laboratory Medicine Program, London Health Sciences Centre, ON, Canada
\textsuperscript{d}Cytognomix Inc., London, ON, Canada
\textsuperscript{e}Department of Physiology and Pharmacology, Schulich School of Medicine and Dentistry, University of Western Ontario, London, ON, Canada
\textsuperscript{f}Department of Physiology and Pharmacology, Schulich School of Medicine and Dentistry, University of Western Ontario, London, ON, Canada
\textsuperscript{g}Department of Medical Oncology, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy
\textsuperscript{h}Department of Diagnostic and Laboratory Pathology, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy
\textsuperscript{i}Department of Computer Science, University of Western Ontario, London, ON, Canada
\textsuperscript{i}Department of Oncology, Schulich School of Medicine and Dentistry, University of Western Ontario, London, ON, Canada

\textbf{ABSTRACT}

Increasingly, the effectiveness of adjuvant chemotherapy agents for breast cancer has been related to changes in the genomic profile of tumors. We investigated correspondence between growth inhibitory concentrations of paclitaxel and gemcitabine (GI\textsubscript{50}) and gene copy number, mutation, and expression first in breast cancer cell lines and then in patients. Genes encoding direct targets of these drugs, metabolizing enzymes, transporters, and those previously associated with chemoresistance to paclitaxel (n = 31 genes) or gemcitabine (n = 18) were analyzed. A multi-factorial, principal component analysis (MFA) indicated expression was the strongest indicator of sensitivity for paclitaxel, and copy number and expression were informative for gemcitabine. The factors were combined using support vector machines (SVM). Expression of 15 genes (\textit{ABCC10}, \textit{BCL2}, \textit{BCL2L1}, \textit{BIRC5}, \textit{BMF}, \textit{FGF2}, \textit{FN1}, \textit{MAP4}, \textit{MAFT}, \textit{NFKB2}, \textit{SLCO1B3}, \textit{TLR6}, \textit{TMEM243}, \textit{TWIST1}, and \textit{CSAG2}) predicted cell line sensitivity to paclitaxel with 82% accuracy. Copy number profiles of 3 genes (\textit{ABCC10}, \textit{NT5C}, \textit{TYMS}) together with expression of 7 genes (\textit{ABCB1}, \textit{ABCC10}, \textit{CMPK1}, \textit{DCTD}, \textit{NME1}, \textit{RRM1}, \textit{RRM2B}) predicted gemcitabine response with 85% accuracy. Expression and copy number studies of two independent sets of patients with known responses were then analyzed with these models. These included tumor blocks from 21 patients that were treated with both paclitaxel and gemcitabine, and 319 patients on paclitaxel and anthracycline therapy. A new paclitaxel SVM was derived from an 11-gene subset since data for 4 of the original genes was...
1. Introduction

Chemotherapeutic agents, such as paclitaxel and gemcitabine, are recommended to patients with developed metastases, basal-like breast cancer, and high-risk indications (premenopausal, ER/PR-negative, HER2-status, large tumors, or node-positive) (Cardoso et al., 2012; Oostendorp et al., 2011). There is currently no gold standard chemotherapy regimen (Cardoso et al., 2012; Oostendorp et al., 2011). Treatment selection is suggested to be individualized and should take into account clinical disease characteristics, treatment history, patient-related factors, and patient preference. However, resistance is one of the major barriers to successful therapy. In a recent study, breast cancer patient response rates to paclitaxel and gemcitabine after 6 cycles of chemotherapy were found to be only 50.0% and 78.6% respectively (Lee et al., 2014). This has motivated a number of groups to develop gene signatures aimed at predicting therapeutic response to these drugs in breast cancer patients (Gąsowska-Bodnar et al., 2014; Hatzis et al., 2011; He et al., 2014).

As in breast cancer patients, breast cancer cell lines show variable responses to growth inhibition by paclitaxel and gemcitabine (Daemen et al., 2013; Shoemaker, 2006). Cell lines mirror many of the pathological features of breast tumors, such as the intrinsic subtypes of breast cancer (Neve et al., 2006; Prat et al., 2013), and can be useful for testing anticancer therapy responses (Heiser et al., 2012). Daemen et al. (2013) employed random forest machine learning to assess genomic information from 70 breast cancer cell lines (including DNA sequence, gene copy number, gene expression, promoter methylation, protein expression, and the corresponding cell line response to 90 anti-cancer compounds) with the objective of establishing pretreatment signatures that predict response. The gene expression profile of the tumor subtype was found to be the most effective way to model response to therapy. However, many molecular signatures derived using genome-wide approaches are inconsistent between different data sets (Ein-Dor et al., 2006; Nilsson et al., 2009). This is partly due to the fact that deriving predictive gene models using thousands of genes risks overtraining, that is, fitting the noise rather than the actual gene signature in the data (Ein-Dor et al., 2006).

We recently defined a set of genes that are stable in gene expression and copy number in the majority (>90%) of breast cancer tumors (Park et al., 2012). The stable gene set is composed of genes that are unmutated in the majority of tumors. Interestingly, many stable gene products were found to be targets of paclitaxel and gemcitabine. We examine the possibility that genomic differences in expression, copy number or mutation in these genes may be related to GI50. Rather than a genome-wide approach to predict sensitivity to paclitaxel and gemcitabine (eg. employed by Daemen et al. (2013)), we analyze stable and linked unstable genes in pathways that determine their disposition (Figure 1).

Figure 1 — Workflow to derive gene signatures. Gene sets were derived for paclitaxel and gemcitabine based on known drug pathways, metabolism, and genes previously implicated in resistance. A multiple factor analysis was completed for each gene to determine which data types (gene expression, copy number, and mutation data) were correlated with the growth inhibitory values for paclitaxel and gemcitabine. Gene expression values were used to derive the paclitaxel SVM classifier, and both gene expression and copy number were used for the gemcitabine SVM. Cell lines were then clustered on optimized gene sets to visualize stratification of tumor subtype and sensitivity. The SVM classifiers were validated using random gene iterations to determine the significance of the classification accuracy, and patient data sets to ensure robustness of the models derived.
Gene panels were established based on biological and experimental studies of paclitaxel and gemcitabine metabolism. Paclitaxel binds to the β subunit of tubulin (TUBB1), inhibiting microtubule formation during mitosis (Jordan and Wilson, 2004). It also binds BCL2, which induces programmed cell death (Ferlini et al., 2009). Paclitaxel is now also recognized to target microtubule–associated proteins 2 (MAP2), 4 (MAP4) and Tau (MAPT) (McGrogan et al., 2008), as well as the xenobiotic receptor (NR112, or PXR) (Harmsen et al., 2009). SLC01B3 transports paclitaxel into cells (Smith et al., 2005), and it is exported by ABCB1 (P-glycoprotein), multidrug resistance–associated proteins ABCC1 (Heijn et al., 1997) and ABCC10 (Chen et al., 2003), and the bile salt export pump ABCB11 (Lecureur et al., 2004). Other genes previously implicated as contributing to paclitaxel resistance include TMEM243 (Duan et al., 2004), BCAP29 (Rao et al., 2008), RBP1 (Duan et al., 2006), TLR6 (Kaczanowska et al., 2013), NFKB2 (Tantivejkul et al., 2005), FGF2 (Carmo et al., 2011), BIRC5 (Lu et al., 2009), TWIST1 (Hong et al., 2011), FN1 (Xing et al., 2008), OPRK1 (Duan et al., 2005), CSAG2 (Duan et al., 1999), and CNGA3 (Duan et al., 2005). Additionally, genes expressed in breast tissue involved in paclitaxel metabolism were included: CYP2C8 and CYP3A4 (Freissner et al., 2010), as well as stable genes in pathways of known direct targets (Park et al., 2012): BAD, BBC3, BCL2L1, BMF, TUBB4A (Kavallaris, 2010), and TUBB4B (Kavallaris, 2010).

Gemcitabine, a deoxycytidine analog, is transported into the cell by SLC29A1 (Marcé et al., 2006), SLC29A2, SLC28A1 (Mackey et al., 1999), and SLC28A3 (Govindarajan et al., 2009). The pro-drug is then phosphorylated by DCK, CMPK1, and NME1 to gemcitabine diphosphate and triphosphate (Ueno et al., 2007). These active forms are incorporated into DNA, which halts replication and cell growth (Plunkett et al., 1995). Gemcitabine di- and triphosphate target ribonucleotide reductase (RRM1, RRM2, and RRM2B), and inhibit DNA synthesis (Mini et al., 2006). An alternative metabolite, difluorodeoxyuridine monophosphate, which is derived by cytidine deaminase (CDA) or dCMP deaminase (DCTD), inhibits thymidylate synthetase (TYMS), resulting in apoptosis (Ueno et al., 2007).

We examine the hypothesis that genomic differences in genotypes, expression and copy number of these genes explain concentration-dependent growth inhibition by gemcitabine and paclitaxel. We then use machine learning to stratify the relative contributions of different genes to chemoresistance, by identifying corresponding genomic signatures at the transcriptional and genomic level in both cell line and patient data.

2. Materials and methods

2.1. Data acquisition

Growth inhibition (GI50), copy number, gene expression, and exome sequencing data were obtained from the supplementary data of Daemen et al. (2013). GI50s (–log10 M, where M is the drug concentration required to inhibit cell line growth by 50%) for paclitaxel were available for 49 cell lines and GI50s for gemcitabine were available for 47 cell lines. Supplementary Table 1 indicates the cell lines used and Supplementary Table 2 indicates the gene, gene product names and their respective drug disposition functions. Supplementary Methods I & II describe copy number and variant calling, results of which are shown in Supplementary Tables 3 and 4. Log2 normalized gene expression data were derived from Affymetrix Gene Chip Human Exon 1.0 ST arrays. Replication studies performed to re-measure and confirm GI50s, verify copy number and mutation data for a subset of the cell lines are outlined in Supplementary Methods III. Figure 1 is an overview of the complete workflow used.

2.2. Cell lines

Cell lines were composed of 10 basal, 9 claudin-low, 25 luminal, and 5 normal-like subtypes. Cell lines were designated resistant, if their GI50 was <8.0 for paclitaxel and <7.0 for gemcitabine, respectively. The threshold values for distinguishing sensitive from resistant cell lines were based on median GI50s for each particular drug (7.99 and 7.13, for paclitaxel and gemcitabine), Daemen et al. (2013) classified cell lines by comparing mean GI50s. We used median GI50, which is not impacted to the same extent by outlier cell lines.

2.3. Multiple factor analysis (MFA)

MFA was used to relate each cell line GI50 according to sets of genomic variables (Abdi and Valentini, 2007). The 44 (gemcitabine) or 45 (paclitaxel) breast cancer cell lines (Supplementary Table 1) were treated as separate individuals. MFA was carried out with the R library “FactoMineR” (Llobet and Esnaola, 2010).

2.4. Support vector classification

A binary support vector machine (SVM) was trained with the Statistics Toolbox in MATLAB (Natick, MA) using fcsvm (linear kernel function) and then tested with a leave-one-out cross-validation (using ‘crossover’ and ‘leaveout’ options). The SVM was trained on the cell lines and explanatory gene variables deemed relevant from the MFA: expression data for the paclitaxel SVM, and copy number and expression data for the gemcitabine SVM. The input data consisted of measurements from all genes used in the MFA. Sequential backward feature selection was performed for feature optimization (Dash and Liu, 1997) to minimize the percentage of misclassified cell lines (classification error) returned from the leave-one-out cross validation (Supplementary Methods IV). Genes that did not reduce or change the classification error were removed from the SVM (one at a time). This procedure was iterated until further gene removal lead to a higher classification error (stopping criterion). The SVM used to analyze patient expression and outcomes of paclitaxel treatment excluded 2 of the 49 cell lines (184A1 and 184BS) from the feature selection process and the subsequent validation. This was done because these lines were not derived from breast cancer tissue, but rather normal tissue that was transformed by exposure to benzo(a) pyrene. By contrast with the SVM, a partial-least squares regression was not effective in relating genomic findings to paclitaxel response (Supplementary Methods V).
The hinge loss was also determined for the subset of genes included in the final SVMs. Hinge loss applies a linear penalty for misclassified data according to their distance from the hyperplane. The loss function is represented by Equation (1) where \( y_j = \{-1,1\} \) and \( f(X_j) \) is the score, i.e. hyperplane distance, for cell line \( j \):

\[
L = \max \left( 0, 1 - y_j f(X_j) \right)
\]

(1)

### 2.5. Applying the cell line SVM to patient data

Formalin fixed, paraffin embedded (FFPE) tumor samples were obtained from the Fondazione IRCCS Istituto Nazionale dei Tumori (Milan, Italy), from leftover material available after diagnostic procedures in consented patients (Musella et al., 2015). Samples obtained were from patients that were first treated with paclitaxel (or in a small number of cases docetaxel) and carboplatin, and then subsequently gemcitabine, upon development of resistance. Clinical information was available as to whether the patients responded to each of the drugs (paclitaxel and gemcitabine). Tumor and control normal tissues were analyzed for expression and copy number of SVM genes, respectively, by real-time reverse-transcriptase polymerase chain reaction (qPCR) and real time PCR (qPCR, methods described in Supplementary Methods, Section VI). The cell line-based SVM models were used to predict patient sample drug responses in a blinded manner. Two SVM models were trained for paclitaxel and gemcitabine: one using the normalized gene expression values, and the other using expression values binned into 10 categories, using the Matlab function: quantile(X,10). Binning was performed because amplifiable RNA template concentration in FFPE blocks is not known precisely, because it is subject to long term degradation and reactivity (Antonov et al., 2005; Fleige and Pfaffl, 2006). Expression measurements were obtained for 11 genes from the paclitaxel SVM, and 6 genes for the gemcitabine SVM. The SVM was trained on the cell line data with these reduced gene sets. Predicted and actual responses were compared, and odds risk ratios (contingency analysis) were calculated (GraphPad Prism, San Diego, California).

Patient data were also obtained from GEO Accession GSE25066, in which expression levels of tumors that were treated with taxane and anthracycline chemotherapy were reported (Hatzis et al., 2011). Expression levels for the paclitaxel SVM genes (except BMF and CSAG2, which were not measured) were extracted for those patients treated with paclitaxel \((n = 319)\). In cases with multiple probe sets per gene, expression levels were averaged. The SVM predictions were then related to response to therapy and residual cancer burden class for each patient.

### 2.6. Clustering cell lines and patients using expression values of the SVM gene subsets

The unsupervised, hierarchical clustering function ‘clustergram’ in Matlab was used to cluster cell lines and patient data (described in 2.5) according to gene expression values included in the optimized SVM. Expression values were normalized by row so the mean expression of each gene across individuals was 0, and the standard deviation was 1. Clustering was performed by individuals and genes, and dendrograms are displayed for each dimension that indicate relatedness based on their lengths and hierarchical branching.

### 3. Results

#### 3.1. Multiple factor analysis

MFAs were performed using GI50s of 49 cell lines, and genomic measurements of 31 and 18 genes related to paclitaxel and gemcitabine activity from an existing data set (Daemen et al., 2013). We re-confirmed measurements for a subset of the cell lines to ensure consistency between cell line sources (see Supplementary Results I). MFAs were assessed by statistics generated by the program, FactoMineR (Lê et al., 2008). Relationships were stratified by the correlation between the variable and GI50, the RV coefficient (a multivariate generalization of the squared Pearson correlation coefficient), the position of variables on the correlation circle, and the representation quality of each variable group in the first two dimensions (cos2 values). These criteria were used to classify each gene as having a “strong relationship”, “relationship”, “possible relationship” or “no relationship” to GI50 (see Supplementary Table 5 for the thresholds for each class). Examples of correlation circles and individual factor maps for MAPT (paclitaxel) and DCTD (gemcitabine) are illustrated in Supplementary Results II.

MFA revealed “strong relationships” between paclitaxel GI50 and copy number and/or gene expression for 11 genes, consisting of both negative relationships (diminished copy number and gene expression [−] for CYP2C8, CYP3A4, NR1I2 (previously known as PXR), TLR6, and TUBB1) and positive relationships (increased copy number and gene expression [+] for BBC3, BCL2L1, BMF, CNGA3, MAPT, and TUBB4B) with increased chemoresistance (Supplementary Table 6 lists all MFA measurements). The gemcitabine set revealed strong associations between resistance and ABCB1 (+), DCTD (−), and SLC28A1 (+) gene expression as well as strong relationships for ABCG2 (−) and CDA (+) copy number(Supplementary Table 7). The MFA results for paclitaxel (gene expression results only) and gemcitabine treatment (copy number and gene expression), in the respective pathway contexts, are summarized in Figure 2.

Point mutation status was based on 74 deleterious coding mutations (Supplementary Table 4) that were predicted to be damaging (Ng and Henikoff, 2003) or to affect mRNA splicing (Shirley et al., 2013; Viner et al., 2014). Point mutations predicted to be damaging demonstrated strong relationships in ABCB1 \((n = 4, \text{in 2 cell lines})\) to paclitaxel resistance and in SLC28A3 \((n = 3, \text{in 2 cell lines})\) to increased sensitivity to gemcitabine. The limited number of cell lines with mutations in these genes cannot be effectively incorporated into machine learning models, and point mutation results were not included in these analyses.

#### 3.2. Support vector machine (SVM) learning

A binary SVM was employed to develop a predictive multigene classification of genomic signatures for resistance to these
Figure 2 — Genes associated with paclitaxel (A) and gemcitabine (B) mechanism of action (direct targets, metabolizing enzymes), genes previously associated with resistance, and stable genes in the biological pathways targets. Genes with an asterisk (*) are stable genes (Park et al., 2012). Genes highlighted in red showed a positive correlation (within dimension 1 and/or dimension 2) between gene expression or copy number, and resistance in the MFA, whereas genes highlighted in blue demonstrated a negative correlation. Genes outlined in dark grey are those included in the final predictive model that was derived using the SVM. Red T-shaped bars indicate the genes that paclitaxel directly binds/inhibits. Genes outlined in light grey (ie. BAX/BAK) were not included in the analysis because they were not stable genes in the BCL2 pathway.
drugs (Furey et al., 2000). Based on MFA results, data types orthogonal to GI50 were excluded from the SVM (see section 2.4 for details). The classification error of the SVM model was minimized by removing genes, i.e. features, which did not improve accuracy by leave-one-out cross-validation. This feature selection process is illustrated in Figure 3-I. The optimized SVM was then trained, respectively, on 15 gene variables for paclitaxel (49 cell lines) and 10 variables for gemcitabine (44 cell lines). Gene expression values from ABCC10, BCL2, BIRC5, BMF, FGFR2, FN1, MAP4, MAPT, NFKB2, SLCO1B3, TLR6, TMEM243, TWIST1, and CSAG2 comprised the final set of features used to train the SVM for classification of paclitaxel sensitivity. For gemcitabine, both gene expression values (from ABCB1, ABCC10, CMPK1, DCTD, NME1, RRM1, RRM2B) and copy number data (from ABCC10, NT5C, TYMS) were used in the final SVM. The distance of each cell line value from the SVM hyperplane that distinguishes the degree of sensitivity or resistance was plotted against the corresponding GI50 (Supplementary Results III). The trained SVMs misclassified 9 of 49 (18%) cell lines for paclitaxel and 7 of 44 (16%) for gemcitabine, which is comparable to or more accurate than other approaches (Ma et al., 2004, demonstrated 19% misclassification for tamoxifen). Partitioning by histological subtype did not improve the classification accuracy; a single variable SVM model based on subtype misclassified 30% of cell lines for paclitaxel and 45% for gemcitabine (Supplementary Table 8). The feature-optimized SVM outperformed the signature derived from the initial set of genes, which misclassified resistance/sensitivity of 36% cell lines for paclitaxel and 64% for gemcitabine treatments. In addition, multi-gene MFA analyses of the final SVM gene sets demonstrate that the individual factor maps of the resistant and sensitive cell lines segregate to a greater degree than MFAs based on the initial gene sets, which were indistinguishable (Supplementary Results IV). These differences were larger for gemcitabine than paclitaxel.

To assess the individual impacts of a gene on SVM accuracy, each gene remaining in the optimized SVM was removed, and the misclassification rate was redetermined

![Figure 3](image-url)
such as (Kutuk and Letai, 2008). The loss of taxel is known to trigger apoptosis through these pathways (Ferlini et al., 2009). Additional apoptotic regulators, expression and increasing paclitaxel sensitivity.

3.3. Applying the cell line-trained SVM to patient data

Formalin fixed paraffin embedded (FFPE) tissue blocks were obtained from patients that were treated with paclitaxel and gemcitabine, and whose response to both drugs are known. Gene expression measurements for 11 genes from the paclitaxel SVM, and gene expression (6 genes) and copy number (CN; 3 genes) from the gemcitabine SVM were obtained using qRT-PCR and qPCR (Supplementary Tables 9 and 10). Gene expression measurements for 11 genes from the paclitaxel (Bhat and Setaluri, 2007), which is supported by our analysis.

For gemcitabine, removing NT5C copy number, NME1 gene expression, ABCC10 gene expression, and RRM2B gene expression had the largest effects, by respectively increasing misclassification rates to 34%, 32%, 32%, and 30% (Figure 3B-II). NT5C is located on 17q25.1 a region associated with cancer (Fukino et al., 1999). Allelic imbalances in TYMS have previously been hypothesized to be involved in drug resistance in renal cell carcinoma (Colavito et al., 2009) and ABCG1 has been associated with drug resistance (Hopper-Borge et al., 2009). NME1 is a known metastasis suppressor gene which may have great prognostic value (Shoushtari et al., 2011). RRM2B and RRM1 have been suggested to be associated with gemcitabine resistance (Aye et al., 2014) and have been shown to be overexpressed in a gemcitabine-resistant pancreatic cancer cell line (Wang et al., 2015).

Table 1 — Using the SVM to predict patient response from archived FFPE tissue.

<table>
<thead>
<tr>
<th></th>
<th>Paclitaxel</th>
<th>Gemcitabine</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>NORM 10 bins</td>
<td>NORM 10 bins</td>
</tr>
<tr>
<td>No. of accurate predictions</td>
<td>12</td>
<td>9</td>
</tr>
<tr>
<td>Total</td>
<td>17a</td>
<td>17</td>
</tr>
<tr>
<td>Percent accurate</td>
<td>71%</td>
<td>71%</td>
</tr>
<tr>
<td>Odds ratio</td>
<td>5.83</td>
<td>6.00</td>
</tr>
<tr>
<td>P-value</td>
<td>0.1534</td>
<td>0.1534</td>
</tr>
</tbody>
</table>

a 4 patients were treated with docetaxel instead of paclitaxel, and were not included in this summary.
b Fisher’s exact test. Gene expression values were either normalized (NORM) or binned into 10 categories (10 bins), as described in the methods. Please refer to Supplementary Table 11 for all FFPE clinical response and prediction data.
described in Hatzis et al., 2011 for both sensitive and resistant patients. The odds ratio of the 11-gene SVM was 4.484 (Fisher’s exact test, p < 0.0001), compared to the odds ratio of 3.181 of the predictive signature described in that study (Fisher’s exact test, p < 0.0001).

3.4. Clustering cell line and patient data based on SVM gene subsets

Two distinct groups emerge from unsupervised clustering using the SVM gene set for paclitaxel in the cell line data (Figure 4A). The left cluster (highlighted in light grey) corresponds with the luminal subtype, and the right corresponds (Figure 4A). The left cluster (highlighted in light grey) corresponding the SVM gene set for paclitaxel in the cell line data.

Two distinct groups emerge from unsupervised clustering using the SVM gene set for paclitaxel in the cell line data (Figure 4A). The left cluster (highlighted in light grey) corresponds with the luminal subtype, and the right corresponds (Figure 4A). The left cluster (highlighted in light grey) corresponding the SVM gene set for paclitaxel in the cell line data.

Table 2 – SVM predictions on 319 patients treated with paclitaxel from Hatzis et al. (2011).

<table>
<thead>
<tr>
<th>Cell line 11-gene SVM</th>
<th>Hatzis “Rx” prediction</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>RD</td>
</tr>
<tr>
<td>All RCB classes</td>
<td></td>
</tr>
<tr>
<td>Predicted insensitive</td>
<td>119</td>
</tr>
<tr>
<td>Predicted sensitive</td>
<td>138</td>
</tr>
<tr>
<td>Odds ratio</td>
<td>4.484</td>
</tr>
<tr>
<td>P-value</td>
<td></td>
</tr>
<tr>
<td>RCB class 0/1 only</td>
<td>11</td>
</tr>
<tr>
<td>Predicted insensitive</td>
<td>19</td>
</tr>
<tr>
<td>Predicted sensitive</td>
<td></td>
</tr>
<tr>
<td>Odds ratio</td>
<td>3.011</td>
</tr>
<tr>
<td>P-value</td>
<td></td>
</tr>
</tbody>
</table>

RD = recurrent disease (designated “insensitive” patient response), pCR = pathological complete response (designated as “sensitive” patient response), RCB = residual cell burden (as described in Symmans et al., 2007).

a p-values were determined using a Fisher’s exact test. Please refer to Supplementary Table 12 for all predictions and patient information.

3.5. Significance of SVM classification accuracy

To assess the significance of the derived SVM, we selected 100,000 random sets of 15 genes from a set of expression values (to compare to the paclitaxel SVM) and 10 genes from a set of copy number and expression values (gemcitabine SVM) for 23,030 genes. Only 0.14% of paclitaxel and 0.01% of gemcitabine random gene combinations exceeded the classification accuracy of the derived SVMs. (Figure 6 – A/C). The hinge loss, which increases based on the misclassified object’s distance to the hyperplane, was 0.64 for the paclitaxel SVM and 0.66 for the gemcitabine SVM (optimal is close to zero). Among the random gene combinations, the likelihood of deriving SVMs with equal or lower scores was 1.45% and 0.83% for paclitaxel and gemcitabine, respectively (Figure 6 – B/D). Thus, the accuracy of the SVMs achieved for both drugs were not likely due to random chance (p < 0.05 in all cases, Table 3).

Nearly all of the high performance random gene set combinations appear to be statistical artifacts. Analysis of 10,000 random gene selections found 18 combinations with lower paclitaxel misclassification response rates. All 18 signatures were unique (2 transcripts occurred twice) and transcript combinations were dominated (24%) by alternative splice variants and expressed pseudogenes. None of the random gene combinations were significantly associated with known biological pathways. Six of the random signatures contained ≥10 gene expression values in the patient data. None of these signatures predicted paclitaxel sensitivity, except one set containing WWP1, which has previously been suggested to be a prognostic indicator in breast cancer (Nguyen Huu et al., 2008). This signature (and one based on WWP1 expression alone) predicted more patients to be sensitive to paclitaxel (Hatzis et al., 2011) than our derived SVM. Similar numbers of patients predicted to be sensitive by both SVM models exhibited complete remission (52 vs. 55), however the WWP1-based SVM predicted sensitivity in a greater number of non-responders (n = 178) than our derived SVM (n = 138) and misclassified 41% of the cell lines. For the gemcitabine response, the SVM of a single random gene set had a lower misclassification rate than our derived SVM. The genes in this set were unrelated to gemcitabine metabolism, with 9 of 10 SVM variables exhibiting copy number changes, two of which involved non-coding RNA genes.
Figure 4 – Expression heatmap of the paclitaxel and gemcitabine SVM derived genes for the tested cell lines. Each row represents a gene and each column a cell line. Red indicates higher expression and blue represents lower expression, as shown by the color bar on the left. ‘Resistant’ cell lines are colored grey and ‘sensitive’ cell lines are colored white in the row labeled ‘response’. Cell lines are labeled by subtype and copy number according to the legends. Clustering was done based on the similarity of each cell line’s expression profile in the 1st (column) dimension and each gene’s expression profile in the 2nd (row) dimension. The dendrograms on the top and left indicate the relatedness of each cell line and gene by the length and subdivision of the branches, with deeper branches indicating a stronger relationship and branches in the same ‘tree’ being more closely related to each other than data in other ‘trees’. A) A section of the dendrogram for paclitaxel is shaded grey to indicate a cluster composed entirely of luminal cell lines and a higher proportion of resistant cell lines. The other section is white to indicate a cluster with very few luminal cell lines and a higher proportion of sensitive cell lines. B) A section of the dendrogram for gemcitabine is shaded grey to indicate a cluster composed of a higher proportion of resistant cell lines. The other section is white to indicate a cluster with a higher proportion of sensitive cell lines.
We have developed an online web-based SVM calculator (http://chemotherapy.cytognomix.com) to predict chemosensitivity from normalized gene expression and copy number (gemcitabine) values. The user may either input their own values or populate them from available patient or cell line results that determined the accuracy of the SVM.

3.6. Translation of signature to other cancer types

To mitigate tissue-specific effects, we rederived SVM models specific to lung cancer (lung) and hematopoietic and lymphoid tissue cancer (hematopoietic) cell lines using expression data from the broad institute (www.broadinstitute.org/ccle/home; “CCLE_Expression_2012-09-29.res” and “CCLE_NP24.2009_profiling_2012.02.20.csv”). Lung and hematopoietic tissue types were chosen because they contained the highest number of cell lines with expression and paclitaxel GI50s. The final lung SVM contained 14 genes, and classified cell lines with 72% accuracy (Supplementary Results VIII – A). The final hematopoietic SVM was composed of 8 genes, and classified cell lines with 75% accuracy (Supplementary Results VIII – B). Four genes were present in all three (breast, lung and hematopoietic) cancer cell line SVMs (BMF, FGF2, TMEM243, and TWIST1), and 8 genes were eliminated from all of the SVMs (ABCB11, BBC3, CNGA3, CYP2C8, CYP3A4, NR1I2, TUBB4A, and TUBB4B; Supplementary Results VIII – C). MFAs using the Lung and Hematopoietic SVM gene sets do not show the same degree of segregation between resistant and sensitive cell lines as the breast SVM (Supplementary Results VIII – D/E).

4. Discussion

This paper describes the development of genomic signatures using support vector machines that can predict breast cancer tumor response to paclitaxel and gemcitabine. We used a biologically-driven approach to identify a meaningful group of genes whose expression levels and copy number may be useful in guiding selection of specific chemotherapy agents during patient treatment. Previous studies have derived associations between the genomic status of one or more genes and tumor response to certain therapies (Duan et al., 2003; Glinsky et al., 2005; Hatzis et al., 2011; Ma et al., 2004; Rajput et al., 2013; van’t Veer et al., 2002). Correlations between single gene expression and tumor resistance (Duan et al., 2003,
do not take into account multiple mechanisms of resistance or assess interactions between multiple genes. ABC transporter overexpression has long been shown to confer resistance, but enzymatic or functional inhibition has not substantially improve patient response to chemotherapy (Samuels et al., 1997).

Multi-gene analytical approaches have previously been successful in deriving prognostic gene signatures for metastatic risk stratification (Oncotype DX®, MammaPrint®, subtypes (PAM50), and efforts have been made to predict chemotherapy resistance (Hess et al., 2006; Hatzis et al., 2011). Given the complexity of genomic changes and the fundamental

Table 3 – SVM performance using randomly selected genes based off 100,000 iterations.

<table>
<thead>
<tr>
<th></th>
<th>Minimum</th>
<th>Maximum</th>
<th>Average</th>
<th>Standard deviation</th>
<th>Drug SVM</th>
<th>z-score</th>
<th>p-value</th>
<th>No. random SVM ≤ drug SVMs*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Percent misclassification of cell lines in leave-one-out analysis</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>15-gene</td>
<td>12.2%</td>
<td>83.7%</td>
<td>42.7%</td>
<td>8.8%</td>
<td>18.4%</td>
<td>−2.78</td>
<td>0.0027</td>
<td>141</td>
</tr>
<tr>
<td>10-gene</td>
<td>12.2%</td>
<td>90.2%</td>
<td>48.0%</td>
<td>10.5%</td>
<td>15.9%</td>
<td>−3.06</td>
<td>0.0011</td>
<td>10</td>
</tr>
<tr>
<td>Hinge loss score</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>15-gene</td>
<td>0.39</td>
<td>2.02</td>
<td>1.05</td>
<td>0.18</td>
<td>0.64</td>
<td>−2.04</td>
<td>0.0207</td>
<td>1453</td>
</tr>
<tr>
<td>10-gene</td>
<td>0.30</td>
<td>2.02</td>
<td>1.05</td>
<td>0.18</td>
<td>0.66</td>
<td>−2.16</td>
<td>0.0153</td>
<td>826</td>
</tr>
</tbody>
</table>

Misclassification rates and hinge loss scores were determined from SVMs derived using 100,000 random combinations of gene expression and copy number values from 23,030 genes. The minimum, maximum, average, and standard deviations of each 100,000 iterations were determined, and compared to the paclitaxel and gemcitabine SVMs ("drug SVM").

a The number of random gene combinations with equal or lower misclassification rates or hinge loss scores compared to the drug SVMs.
b Random selection of 15 gene expression values were compared to the paclitaxel SVM.
c Random selection of 10 gene expression or copy number values were compared to the gemcitabine SVM.
biological differences among the intrinsic subtypes of breast cancer (Cancer Genome Atlas Network, 2012; Dorman et al., 2014), this approach has advantages over analysis of isolated genes. Reasonable gene signatures associated with breast cancer outcome can be obtained by chance alone (Venet et al., 2011), however our results show that such signatures are especially rare. Gene signatures derived without reference to the underlying mechanisms of chemotherapy response do not capture meaningful biological results (Drier and Domany 2011).

Our approach started with a focused biologically-relevant initial gene set, rather than taking a genome-wide approach. The derived signatures were demonstrated to significantly outperform random selected combinations of genes in prediction of sensitivity and resistance. The random gene sets may be statistical artifacts, as they were not enriched for any biological relevant pathways, and included expressed pseudogenes. The compositions of these other gene sets were distinct from the set used to derive the SVM and another 20-gene signature for taxane sensitivity (He et al., 2014).

Our analysis highlights the importance of the expression of genes encoding microtubule-associated proteins and apoptotic regulators in paclitaxel resistance (McGrogan et al., 2008; Tanaka et al., 2009; Wang et al., 2013). MAPT expression was significantly correlated with drug resistance, and both MAPT and MAP4 were components of the optimized paclitaxel SVM gene set. In clustering analysis of both cell lines and patients, MAPT was differentially expressed between tumor clusters stratified by subtype and outcome (Figures 4 and 5). Our results confirm that apoptosis-related proteins, particularly BCL2L1, but also BCL2, BMF, and BIRC5, contribute to paclitaxel sensitivity (Flores et al., 2012). BCL2L1, BCL2 and BMF were found to be stable in breast cancer tumors, reinforcing the notion that alterations in stable genes contribute to drug resistance (Park et al., 2012). Supplementary Table 13 describes genes analyzed in the context of their biological pathways and relevant literature.

The gemcitabine metabolic pathway has been well characterized (Alvarellos et al., 2014), however the critical genes have not been treated as an ensemble in conferring resistance (see Supplementary Table 14 for interpretation of the MFA results for all genes). The MFA analyses indicated gemcitabine genes predominantly contribute to drug resistance through overexpression. For DCTD, however, underexpression is associated with increased resistance in the MFA analysis. DCTD deficiency causes an imbalance in the dNTP pool (Eriksson et al., 1984), which affects control of DNA replication. DCTD is inhibited by dFdCTP (a gemcitabine metabolite) through a mechanism by which gemcitabine exhibits self-potentiation (the reduction of competing natural metabolites) (Xu and Plunkett, 1992). Lower DCTD expression and as a consequence, activity, would reduce gemcitabine self-potentiation by altering the dNTP pool. This state is related to drug resistance, which was noticeably lower in 4 cell lines with increased resistance (HCC1187, HCC1428, HCC202, and MDAMB134VI). Like DCTD, CDA also catalyzes the conversion of gemcitabine monophosphate to difluorodeoxyuridine monophosphate (Figure 2B), and accounts for 90% of this conversion in the cell (Govindarajan et al., 2009). However, drug resistance was associated with CDA overexpression. Likewise, the ribonucleotide reductase subunits RRM1 and RRM2B make significant contributions to the gemcitabine SVM. The RRM1-RRM2B complex is associated with mitochondrial genomic integrity (Bourdon et al., 2007) and RRM2B is necessary for nucleotide synthesis in DNA repair (Kuo et al., 2012). Changes in RRM2B expression could be associated with mitochondrial dysfunction, or may result from loss of p53 expression, which usually induces RRM2B expression (Tanaka et al., 2000).

The 11-gene paclitaxel SVM was able to classify FFPE patient samples we obtained and measured in our lab with similar accuracy to that of the cell lines. In addition, the same SVM model was able to predict complete pathological response on a second patient data set, with greater accuracy than the originally reported gene signature (Hatzis et al., 2011). The SVM performed particularly well for predicting drug-sensitive tumors with low or no minimal residual disease (Table 2). The SVM gene signature proved to be resilient as a diagnostic marker, as the performance was not compromised by the lack of expression data for 4 genes.

Unlike paclitaxel, gemcitabine was not used to treat patients in the study by Hatzis et al. (2011) or other publically available data sets. The SVM analysis on RNA expression and DNA copy number from the FFPE-derived tumor punches appeared to predict response more accurately when expression values were obtained for the majority of genes in the SVM. Obtaining high quality gene expression measurements from FFPE samples was especially difficult from older tissue blocks (Supplementary Results V – B) as previously noted (Choudhary et al., 2014). Consequently, the SVM analysis may be better suited for fresh-frozen tumor tissue or more sensitive gene expression analyses (such as mRNA sequencing). Missing data appeared to impact the gemcitabine SVM to a greater extent than the paclitaxel SVM, which may be due to the smaller number of gene measurements required for this SVM.

Including gene expression subtype in the SVM did not improve the classification accuracy even though subtype is known to contribute to tumor biology (Heiser et al., 2012). However, the two paclitaxel PAM50 genes (MAPT and BCL2) partially stratify the cell lines by subtype during unsupervised clustering (Figure 5). This is not the case in the gemcitabine gene set. In patient data, clustering by expression of the SVM genes also revealed statistically significant deterioration in outcome for low MAPT expressing luminal tumors (Supplementary Information VII).

Machine learning may be a fruitful approach in the selection of other chemotherapy agents. Translating our results to the assessment of human tumor samples (Gąsowska-Bodnar et al., 2014) confirmed our gene signature’s relevance to predicting chemoresistance by SVM. However, both SVMs were not integrated because cell lines were treated with individual drugs, so predicting whether patient response to these drug interactions will be synergistic or antagonistic is difficult with our approach. In addition, while point mutations are well known contributors to chemoresistance of other drugs, this approach – for either SVM training or testing – is not conducive for prediction of chemosensitivity given the sparse number of observations for these types of mutations.

In cases without residual disease, the paclitaxel SVM was particularly effective in predicting which tumors would...
show complete pathological response. Docetaxel is prescribed somewhat interchangeably (Crown et al., 2004; Hatzis et al., 2011; O’Shaughnessy et al., 2013) and both paclitaxel and docetaxel act through similar biological pathways (Oshiro et al., 2009). However, the performance of the paclitaxel SVM on patients treated with docetaxel was reduced. This SVM contains 8 paclitaxel resistance genes. Predictions of docetaxel sensitivity might be improved by rederiving a specific SVM using taxane pathway genes (Oshiro et al., 2009), and those known to be associated with resistance to docitaxel (such as those known to be associated with resistance to docitaxel)

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References


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Appendix A.
Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.molonc.2015.07.006.

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Appendix A.
Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.molonc.2015.07.006.


