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Giant beaver palaeoecology inferred from stable isotopes

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This is a multi-individual ($n = 11$), stable carbon and nitrogen isotope study of bone collagen ($\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$) from the giant beaver (genus *Castoroides*). The now-extinct giant beaver was once one of the most widespread Pleistocene megafauna in North America. We confirm that *Castoroides* consumed a diet of predominantly submerged aquatic macrophytes. These dietary preferences rendered the giant beaver highly dependent on wetland habitat for survival. *Castoroides'* $\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$ do not support the hypothesis that the giant beaver consumed trees or woody plants, which suggests that it did not share the same behaviours as *Castor* (i.e., tree-cutting and harvesting). The onset of warmer, more arid conditions likely contributed to the extinction of *Castoroides*. Six new radiocarbon dates help establish the chronology of the northward dispersal of the giant beaver in Beringia, indicating a correlation with ice sheet retreat.

The Pleistocene giant beaver. The one hundred kilogram giant beaver (genus *Castoroides*) inhabited North America throughout the mid- to late Pleistocene^{1–3}. The giant beaver went extinct along with dozens of other genera during the late Pleistocene megafauna extinction^{4,5}. The underlying mechanism(s) behind this global extinction remains contested, though it is often attributed to a combination of climate change and anthropogenic impacts^{6–12}. Current information regarding giant beaver ecology is insufficient to evaluate various hypotheses posited to be responsible for *Castoroides'* extinction. The last appearance date of $10,150 \pm 50$ years BP for a *Castoroides ohioensis* specimen from Wayne County, New York demonstrates that it was also one of the late surviving members of the Pleistocene megafauna community in North America⁶. Here we use stable isotopes to explore the ecology of *Castoroides*, which allows us to better understand its diet, its impact on the surrounding Pleistocene landscape, and the mechanisms that led to its demise. Previous models of *Castoroides'* diet and habitat preference are based on skeletal morphology, fossil depositional environment, and the behaviour of the only remaining extant member of the Castoridae family, *Castor*.

Extant analogue species. The short limbs and bulky body of *Castoroides* made it poorly adapted to a life spent predominantly on land and it must have relied on access to the water for shelter from terrestrial predators¹³. Previous studies based on stable isotope analysis of single specimens suggest that the giant beaver consumed aquatic vegetation, and thrived in mid-latitude regions of North America during warm, strongly seasonal conditions between 125,000 and 75,000 BP^{14,15}. Other studies hypothesize that the giant beaver was a relatively cold-tolerant species, preferentially consumed emergent macrophytes, and lived in ponds and shallow lakes bordered by marshlands^{16–19}. The presence of giant beaver fossils in high latitude regions such as Alaska and Yukon Territory confirm it could persist through harsh arctic climatic and low light conditions in the winter. It is unknown if *Castoroides* shared any behavioural characteristics with the genus *Castor*, and the possibility that the giant beaver engineered its habitat by constructing dams or lodges remains a topic of debate^{1,17,20–24}.

Two competing models of *Castoroides* diet and behaviour have emerged based on extant North American semi-aquatic rodents with well-documented ecologies and distinctly different impacts on the surrounding ecosystem^{14,19,25–27}. The first model predicts that *Castoroides* filled an econiche similar to that of the muskrat (*Ondatra zibethicus*), as a semi-aquatic rodent that primarily consumed submerged and emergent freshwater macrophytes in calm wetlands with an expansive littoral zone^{14,19–21}. Such feeding would have kept shallow waterways clear of excess macrophyte growth and promoted aquatic biotic diversity. The second model suggests that *Castoroides* filled an econiche similar to that of the extant North American beaver (*Castor canadensis*), as a semi-aquatic rodent that consumed both terrestrial and aquatic vegetation, and practiced tree-cutting behaviour for the

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Figure 1. Terrain map of North America indicating sample collection sites. 1 – Old Crow, Yukon Territory, Canada. 2 – Whitehorse, Yukon Territory, Canada. 3 – Pinery Provincial Park and London Area, Ontario, Canada. 4 – Ohio, United States of America. Map data from Google, INEGI, 2019.

purpose of lodge and dam building²⁵. The bulk of its diet would have been foliage from deciduous trees and its engineering habits would have promoted local biotic diversity and significantly impacted the hydrological patterns of the Pleistocene landscape.

Problematic to the latter hypothesis is that there are neither confirmed discoveries of dam or lodge structures built by giant beavers, nor substantiated evidence of Pleistocene-era cut wood that matches the occlusal surface size and angle of *Castoroides* incisors^{28,29}. The angle of protrusion of the incisors from the skull and their lack of a thin, chisel-like edge rendered giant beaver incisors ineffective tools for cutting down trees²¹. This has not stopped speculation that masses of branches discovered in proximity to *Castoroides* fossils in Pleistocene sediments were built by the giant beaver²⁵. In addition, tree harvesting behaviours have been present in certain branches of the Castoridae family since the Miocene^{30,31}.

Stable carbon and nitrogen isotopes and dietary mixing models. The stable carbon and nitrogen isotope compositions ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$) of an animal's body tissues closely reflect those of its diet, with adequately known isotopic discrimination factors occurring between each trophic level^{32–34}. Thus, the isotopic composition of modern and ancient bone collagen serves as a useful proxy for diet^{35–38}. To correctly assess trophic position and forage preference, however, potential dietary sources must be established³⁹.

Here, *Castoroides*, *O. zibethicus* and *C. canadensis* bone collagen carbon and nitrogen isotope compositions ($\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$) are assessed within the context of modern terrestrial and freshwater plant carbon and nitrogen isotope signatures. Modern plant samples were collected from shallow lakes, marshes, and creeks located in the Great Lakes region of southwestern Ontario (Pinery Provincial Park and the London area) and Yukon Territory (Whitehorse and Old Crow Basin), Canada, whereas *Castoroides* specimens originate from sites in Yukon Territory, Canada, and Ohio, USA (Fig. 1). The isotopic data were incorporated into a statistically-based dietary mixing model (SIAR V4: Stable Isotope Analysis in R. An Ecologists' Guide) to determine both forage choice and relative input proportions for each rodent species⁴⁰. This allows us to determine the degree of dietary overlap amongst rodent species, *Castoroides*' level of dependence on aquatic habitat space, and the viability of using *C. canadensis* or *O. zibethicus* as extant analogues in palaeoecological models incorporating the giant beaver. A series of new radiocarbon dates obtained from *Castoroides* bone collagen from Yukon Territory and Ohio are also reported to evaluate the chronology of regional extirpation.

Stable carbon and nitrogen isotopes of freshwater plants. Carbon and nitrogen dynamics are complex in freshwater systems. As a result, freshwater plant, or macrophyte $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ vary widely^{39,41–43}. Plant physiology and local environmental conditions account for broad-scale isotope variability; site specific characteristics (*i.e.*, wetland habitat type, water turbulence, latitude, local geology, water pH, water temperature) also impact the isotopic composition of bioavailable carbon and nitrogen sources. Habitat division within a wetland also has a strong impact on macrophyte $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ ³⁹. Plant functional groups used here (terrestrial trees and shrubs, emergent macrophytes, floating macrophytes, and submerged macrophytes) were categorized not by taxonomic relationship, but by the plant's exposure to the atmosphere, the water column, and/or substrate. These

| Project sample ID | Institute accession ID | Specimen locality | ¹⁴ C age (BP) | ± | AMS Lab code |
|-------------------|------------------------|---------------------------------|--------------------------|------|--------------|
| C3-TP2014 | CMN 18306 | Old Crow Basin, Yukon Territory | >44,600 | | UCLAMS151528 |
| C4-TP2014 | CMN 18707 | Old Crow Basin, Yukon Territory | >43,700 | | UCLAMS151529 |
| C6-TP2014 | CMN 14711 | Old Crow Basin, Yukon Territory | 44,600 | 2600 | UCLAMS151530 |
| C8-TP2014 | CMN 33640 | Old Crow Basin, Yukon Territory | >42,200 | | UCLAMS151531 |
| C11-TP2014 | OHS N9087 | Williams County, Ohio | 11,961 | 80 | AA105557 |
| C22-TP2014 | OHS N8739 | Harmony Township, Ohio | 11,168 | 53 | AA105559 |

Table 1. Radiocarbon ages for *Castoroides* specimens (bone and tooth dentin collagen) analyzed in this study.

three media determine which sources of bioavailable carbon and nitrogen the plant can access, and ultimately control plant $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$.

Complexities arising from the use of modern plants in palaeodietary studies. Anthropogenic influence on global or regional carbon and nitrogen isotope compositions at the base of the food web is one concern when utilizing modern plant $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ as a basis for palaeodietary interpretations. Photosynthetic pathway is the dominant control on plant $\delta^{13}\text{C}$ and atmospheric CO_2 is the primary carbon source for photosynthetic organisms. A Suess effect correction can be applied to modern $\delta^{13}\text{C}$ data to account for the recent addition (primarily through the burning of fossil fuels) of ^{12}C -enriched CO_2 to the atmosphere⁴⁴.

Assessment of the nitrogen isotope baseline is more complicated as it is specific to region and ecosystem. Anthropogenic activities (particularly land development and agricultural practices) can contribute nitrogen sources that enter the local environment and impact the nitrogen isotope composition of primary producers. This effect is particularly pronounced in modern wetland habitats⁴⁵.

There is also evidence that regional nitrogen isotope baselines of some ecosystems changed over the course of the Quaternary. For instance, the dynamic of the nitrogen cycle and the isotopic composition of the nitrogen baseline in grasslands of Yukon Territory have changed between the Late Pleistocene and the present⁴⁶. There, Mammoth Steppe plant macrofossil $\delta^{15}\text{N}$ was on average $\sim 2.8\text{‰}$ higher than that of modern grassland plants collected from same location⁴⁶. This ^{15}N -enrichment at the base of the food web was incorporated into successive trophic levels and contributed to unusually high $\delta^{15}\text{N}_{\text{col}}$ of Mammoth Steppe grassland herbivores (such as the Arctic ground squirrel and the woolly mammoth)^{38,46}. The plant macrofossil ^{15}N -enrichment was likely the result of an arid climate and a more open nitrogen cycle⁴⁷. It is less likely, however, that Pleistocene wetland ecosystems experienced such a shift in $\delta^{15}\text{N}$.

The ^{15}N -enrichment of the nitrogen baseline observed in the Mammoth Steppe grasslands was induced by factors that would not have been nearly as pronounced in Pleistocene wetland ecosystems, or wetter, forested ecosystems with lower megafauna population densities. Late Pleistocene mastodon and giant beaver populations, for example, were contemporary in Yukon Territory and in the Great Lakes region of North America^{48,49}. These species both showed preference for the same type of habitat (open mix-forest interspersed with wetlands). Mastodon were browsers and their collagen is significantly depleted of ^{15}N relative to Mammoth Steppe fauna that were graminoid and forb specialists (e.g., in North America, average $\delta^{15}\text{N}_{\text{col}}$ of mastodon is $\sim 4\text{‰}$ lower than mammoth)^{38,50,51}. In short, while plant enrichment in ^{15}N arising from aridity and associated shifts in the nitrogen cycle can profoundly affect grassland plants, we posit that pronounced changes are much less likely in plants from forested and wetland habitats. In the absence of unaltered ancient equivalents, modern plants from uncontaminated wetlands or wetter, forested habitats can therefore provide an adequate, if not ideal, proxy for incorporation into isotopic models that simulate Pleistocene dietary preferences.

In addition, the relative nitrogen isotope pattern of each plant functional group should remain the same, despite changes in nitrogen baseline $\delta^{15}\text{N}$. Aquatic plants (submerged, emergent, and floating macrophytes), for example, should have, on average higher $\delta^{15}\text{N}$ than terrestrial plants on account of their access to more ^{15}N -enriched sources of bioavailable nitrogen.

Results

Radiocarbon dates. Six new radiocarbon dates obtained from *Castoroides* bone and dentin collagen are reported in Table 1. Bone collagen from *Castoroides* that lived at high latitudes north of the Arctic Circle yielded dates that are analytically non-finite or so close to the analytical limit of radiocarbon dating ($\sim 45,000$ ^{14}C years BP) that they can be treated as non-finite ages. An additional summary of *Castoroides* radiocarbon dates from the literature is presented in Table 2.

Collagen preservation. Collagen yield (wt. %), atomic C:N ratio, and carbon and nitrogen contents (wt. %) were used to assess collagen preservation in all three rodent species (Tables 3 to 5). All *Castoroides* specimen parameters are within the accepted range for archaeological skeletal material from temperate or polar regions (collagen yield $> 1\%$; $\% \text{C} = 34.8 \pm 8.8\%$; $\% \text{N} = 11\text{--}16\%$; C:N = 3.1–3.5)⁵².

Stable isotopes of bone and dentin collagen. *Castoroides* ($n = 11$) $\delta^{13}\text{C}_{\text{col}}$ ranges from -21.2 to -10.9‰ , with a mean of -17.6‰ ; *Castoroides* $\delta^{15}\text{N}_{\text{col}}$ ranges from $+1.9$ to $+7.7\text{‰}$, with a mean of $+5.8\text{‰}$ (Table 3). Extant *O. zibethicus* (Table 4) and *C. canadensis* (Table 5) show distinctly different isotopic patterns. The carbon and nitrogen isotope compositions of muskrat bone collagen closely resembles that of *Castoroides*;

| Taxon | Specimen locality | Tissue type | ^{14}C age (BP) | \pm | AMS Lab Code | Reference |
|------------------------------|--|---------------------|--------------------------|-------|--------------|--|
| <i>Castoroides ohioensis</i> | Fairy Hole Rockshelter, Allamuchy Township, Warren Co., New Jersey | Molar (collagen) | 11,140 | 30 | UGAMS-16240 | Boulanger <i>et al.</i> ⁶⁵ |
| <i>Castoroides ohioensis</i> | Big Brook locality, Holmdel Township, Monmouth Co., New Jersey | Molar (bioapatite) | 13,210 | 30 | UGAMS-18142 | Boulanger <i>et al.</i> ⁶⁵ |
| <i>Castoroides ohioensis</i> | Dutchess Quarry Cave 8, Orange Co., New York | Molar (collagen) | 11,670 | 70 | NSRL-1513 | Steadman <i>et al.</i> ⁹⁶ |
| <i>Castoroides</i> | Clyde, Wayne Co., New York | Bone (collagen) | 10,150 | 50 | OS-73632 | Fernac and Kozlowski ⁹⁷ |
| <i>Castoroides</i> | Kansas River, Johnson Co., Kansas | Mandible (collagen) | 12,150 | 80 | CAMS-20004 | McDonald and Grotzhofer ⁶⁶ |
| <i>Castoroides</i> | Mississippi River, Ramsey Co., Minnesota | Bone (collagen) | 10,320 | 250 | not provided | Erickson ⁹⁸ |
| <i>Castoroides</i> | Clear Creek, Fairfield Co., Ohio | Mandible (collagen) | 12,040 | 35 | UCLAMS-11219 | McDonald and Grotzhofer ⁶⁶ |
| <i>Castoroides</i> | Sheriden Pit, Wyandott Co., Ohio | Bone (collagen) | 10,850 | 60 | CAMS-26783 | Tankersley and Landefeld ⁹⁹ |

Table 2. Literature summary of *Castoroides* radiocarbon dates.

| Project sample ID | Institute accession ID | Specimen locality | Skeletal element | $\delta^{13}\text{C}_{\text{col}}$ (‰, VPDB) | $\delta^{15}\text{N}_{\text{col}}$ (‰, AIR) | C (wt. %) | N (wt. %) | Collagen yield (wt. %) | Atomic C:N |
|-------------------|------------------------|--------------------------|---------------------|--|---|-----------|-----------|------------------------|------------|
| C1-TP2014 | CMN 16657 | Old Crow Basin, YT | Humerus | −21.2 | +6.3 | 41.7 | 15.6 | 1.4 | 3.1 |
| C3-TP2014 | CMN 18306 | Old Crow Basin, YT | Pelvis | −19.1 ± 0.3 | +1.9 ± 0.1 | 41.5 | 15.2 | 1.4 | 3.2 |
| C4-TP2014 | CMN 18707 | Old Crow Basin, YT | Tibia | −10.7 ± 0.2 | +5.7 ± 0.1 | 41.8 | 15.5 | 3.6 | 3.1 |
| C5-TP2014 | CMN no ID | Old Crow Basin, YT | Femur | −18.5 | +7.7 | 41.2 | 15.7 | Data not provided* | 3.1 |
| C6-TP2014 | CMN 14711 | Old Crow Basin, YT | Humerus | −16.0 | +6.0 | 33.1 | 11.9 | 1.1 | 3.2 |
| C7-TP2014 | CMN 14781 | Old Crow Basin, YT | Long bone diaphysis | −14.0 | +7.4 | 39.6 | 14.5 | Data not provided* | 3.2 |
| C8-TP2014 | CMN 33640 | Old Crow Basin, YT | Humerus | −12.4 ± 0.2 | +6.2 ± 0.0 | 39.7 | 14.4 | 2.4 | 3.2 |
| C9-TP2014 | CMN 43178 | Old Crow Basin, YT | Femur | −21.2 | +6.8 | 42.7 | 15.5 | Data not provided* | 3.2 |
| C10-TP2014 | OHS N9109 | Clear Creek Township, OH | Mandible | −20.2 ± 0.1 | +5.6 ± 0.1 | 35.5 | 12.6 | 3.1 | 3.3 |
| C11-TP2014 | OHS N9087 | Williams County, OH | Mandible | −20.6 ± 0.1 | +4.5 ± 0.1 | 40.0 | 14.3 | 11.2 | 3.3 |
| C22-TP2014 | OHS N8739 | Harmony Township, OH | Incisor (dentin) | −19.5 | +5.4 | 37.4 | 13.5 | 3.8 | 3.2 |

Table 3. *Castoroides* specimen information and stable isotope results. Values in bold indicate mean and 1 SD where duplicate analyses were completed for the same specimen. *Collagen extraction performed at the Keck Carbon Cycle AMS Facility at the University of California, Irvine for radiocarbon dating. Remaining collagen material was returned for stable isotope analysis at the University of Western Ontario.

O. zibethicus ($n = 12$) $\delta^{13}\text{C}_{\text{col}} = -22.7$ to -10.7 ‰, with a mean of -15.3 ‰, and *O. zibethicus* $\delta^{15}\text{N}_{\text{col}} = +2.2$ to $+6.6$ ‰, with a mean of $+5.1$ ‰. *C. canadensis* ($n = 8$) $\delta^{13}\text{C}_{\text{col}}$ varies very little, ranging from -24.0 to -23.2 ‰, with a mean of -23.5 ‰. *C. canadensis* $\delta^{15}\text{N}_{\text{col}}$ displays more variation, ranging from $+1.4$ to $+9.5$ ‰, with a mean of $+4.5$ ‰.

Stable isotopes of plants. Modern plant sample $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ are listed in Tables A and B in Supplementary Information File SI1. A summary of plant functional group $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ mean and range are presented in Table C in Supplementary Information File SI1. Generally, macrophytes have higher $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ than terrestrial plants. Submerged macrophytes display a bimodal distribution of $\delta^{13}\text{C}$ (ranging from -41 to -13 ‰). These isotopic patterns are demonstrated in Figure A in Supplementary Information File SI1.

Discussion

Our terrestrial and freshwater plant $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ results are consistent with those of other studies from comparable regions^{53,54}. While we do not have the perfect stable isotope dietary mixing model (*i.e.*, one comprising coeval plant macrofossil and faunal material from the same site), modern plant sampling sites were purposefully chosen that had minimal observed or documented anthropogenic impact. Anthropogenic impact on the nitrogen cycle and isotopic composition of wetland habitats in Pinery Provincial Park is minimal⁵⁵. Plant samples from London, Ontario ($n = 9$) were collected from non-urban areas. While we cannot rule out that the $\delta^{15}\text{N}$ of these nine samples may have been impacted by agriculture, chemical fertilizer ($\delta^{15}\text{N} \approx 0$ ‰) is dominantly used in this region, which constrains possible variation from that source⁵⁶. Yukon Territory has very low human population density ($<36,000$), and minimal industrial and agricultural activities⁵⁷. As such, we expect minimal influence on bioavailable sources of nitrogen (*i.e.*, septic effluent, chemical, manure) and $\delta^{15}\text{N}$ of plants collected around Whitehorse and Old Crow.

In addition, the majority of the plant taxa included in this study (see plant taxonomic information in Supplementary Information File SI1 Tables A and B) are known to have existed in North America during the late Pleistocene (refer to^{19,58–60} for examples of plant macrofossil and pollen records recovered from Pleistocene sites from Beringia and the Great Lakes region). Plants sampled also included tree and macrophyte species documented as part of the diet of extant *C. canadensis* and *O. zibethicus*^{61–64}. All this said, a great opportunity remains

| Project sample ID | Specimen locality | Skeletal element | $\delta^{13}\text{C}_{\text{col}}$ (‰, VPDB) | $\delta^{15}\text{N}_{\text{col}}$ (‰, AIR) | C (wt. %) | N > ((wt. %) | Collagen yield (wt. %) | Atomic > C:N |
|-------------------|--------------------|------------------|--|---|-----------|--------------|------------------------|--------------|
| M1-TP2013 | London, ON | Mandible | −22.7 | +5.0 | 41.1 | 15.5 | 8.6 | 3.1 |
| M3-TP2014 | Old Crow Flats, YT | Mandible | −13.6 | +5.3 | 42.6 | 14.9 | 11.8 | 3.3 |
| M4-TP2014 | Old Crow Flats, YT | Mandible | −14.2 | +2.2 | 42.1 | 16.1 | 12.2 | 3.0 |
| M5-TP2014 | Old Crow Flats, YT | Mandible | −17.5 | +3.8 | 42.5 | 14.9 | 10.8 | 3.3 |
| M6-TP2014 | Old Crow Flats, YT | Mandible | −22.0 | +5.9 | 42.5 | 16.1 | 13.3 | 3.1 |
| M7-TP2014 | Old Crow Flats, YT | Mandible | −15.1 | +6.1 | 42.6 | 16.2 | 11.3 | 3.1 |
| M8-TP2014 | Old Crow Flats, YT | Mandible | −14.6 | +3.9 | 43.2 | 15.1 | 13.4 | 3.3 |
| M9-TP2014 | Old Crow Flats, YT | Mandible | −14.3 | +6.6 | 42.8 | 14.3 | 13.3 | 3.5 |
| M10-TP2014 | Old Crow Flats, YT | Mandible | −15.0 | +5.3 | 43.2 | 16.4 | 12.7 | 3.1 |
| M11-TP2014 | Old Crow Flats, YT | Mandible | −12.6 | +5.7 | 43.6 | 16.7 | 13.5 | 3.0 |
| M13-TP2014 | Old Crow Flats, YT | Mandible | −10.7 ± 0.1 | +6.0 ± 0.0 | 42.6 | 14.9 | 13.7 | 3.3 |
| M14-TP2014 | Old Crow Flats, YT | Mandible | −11.7 | +4.7 | 42.8 | 15.0 | 12.8 | 3.3 |

Table 4. *Ondatra zibethicus* specimen information and stable isotope results. Values in bold indicate mean and 1 SD when duplicate analyses were performed for the same specimen. Specimens collected between August 2013 and August 2014. Stable carbon isotope data are not corrected for the Suess effect.

| Project sample ID | Specimen locality | Skeletal element | $\delta^{13}\text{C}_{\text{col}}$ (‰, VPDB) | $\delta^{15}\text{N}_{\text{col}}$ (‰, AIR) | C (wt. %) | N (wt. %) | Collagen yield (wt. %) | Atomic C:N |
|-------------------|-------------------|------------------|--|---|-----------|-----------|------------------------|------------|
| B1-TP2013 | Inverhuron, ON | Tibia | −23.6 | +7.1 | 43.9 | 16.9 | 12.0 | 3.0 |
| B2-TP2013-A | Dawson City, YT | Metapodial | −24.0 | +3.4 | 44.8 | 16.9 | 10.2 | 3.1 |
| B3-TP2014-1 | New Liskeard, ON | Mandible | −23.7 | +2.0 | 43.6 | 16.6 | 14.4 | 3.1 |
| B4-TP2014-1 | New Liskeard, ON | Mandible | −23.1 | +9.5 | 43.6 | 16.7 | 14.7 | 3.0 |
| B5-TP2014 | New Liskeard, ON | Mandible | −23.2 | +8.0 | 42.7 | 16.3 | 14.8 | 3.1 |
| B6-TP2014-1 | Whitehorse, YT | Mandible | −23.7 | +2.0 | 43.7 | 16.8 | 16.7 | 3.0 |
| B7-TP2014 | Whitehorse, YT | Mandible | −23.3 | +1.4 | 43.1 | 16.6 | 16.6 | 3.0 |
| B8-TP2014-1 | Whitehorse, YT | Mandible | −23.7 | +2.2 | 42.3 | 16.2 | 16.3 | 3.0 |

Table 5. *Castor canadensis* specimen information and stable isotope results. Specimens collected between August 2013 and August 2014. Stable carbon isotope data are not corrected for the Suess effect.

to further test the ideas presented here through isotopic analysis of contemporary Pleistocene plant macrofossils associated with *Castoroides*.

Overall, collagen yields from giant beaver specimens were relatively low. This was particularly surprising for *Castoroides* specimens collected from Old Crow, Yukon Territory. Pleistocene faunal remains from Beringian deposits typically have excellent organic molecule preservation. We propose that the low collagen yields are in fact linked to giant beaver habitat preference. *Castoroides* remains are most commonly recovered from active or ancient wetland environments^{19,65,66}. In addition, specimens from the Canadian Museum of Nature (CMN) collections included in this study were recovered from the banks of the Porcupine River, Yukon Territory, as reported by C.R. Harington. Given its semi-aquatic nature, most giant beavers likely died in close proximity to a wetland habitat. The composite nature of bone (bioapatite crystals interlocked with strands of collagen protein) results in a structure where the mineral component protects the protein component (particularly from detrimental micro-organism enzyme activity)⁶⁷. Exposure to water in a *post-mortem* depositional environment, however, accelerates bone diagenetic processes. The deposition of giant beaver skeletal material in an aqueous environment accelerates dissolution of the bioapatite, after which it can no longer protect the organic fraction and collagen loss is more likely to occur.

A Suess effect correction of 2.1‰ was applied to modern rodent $\delta^{13}\text{C}_{\text{col}}$ and to the $\delta^{13}\text{C}$ of plants collected in 2014. A correction of 1.7‰ was applied to the $\delta^{13}\text{C}$ of plants collected in 2000. Refer to Supplementary Information File SI1 Table A and B for plant sample collection dates. The corrections were calculated using data available through the Scripps CO₂ monitoring program and the $\delta^{13}\text{C}$ of late Pleistocene atmospheric CO₂ described in Schmitt *et al.*^{68,69}. The Suess effect correction was applied when modern $\delta^{13}\text{C}$ data were incorporated into the SIAR mixing model.

Based on existing literature, collagen-diet offsets of +4.2‰ and +3.0‰ were utilized for $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$, respectively^{33,70–72}. Collectively, the corrections for the Suess effect and dietary isotopic discrimination render the rodent $\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$ directly comparable to the plant isotopic data incorporated into the SIAR mixing model (Fig. 2).

The plant functional groupings (terrestrial trees and shrubs, emergent, floating, and submerged macrophytes) were statistically defined using scripts in SIBER in R Studio 3.1.2 (Figure B and C in Supplementary Information File SI1). The Total Area Convex Hull and Standard Ellipse plots aid in identifying the niche size and relative position of each plant functional group. The relative contribution of each plant functional group to the diet of

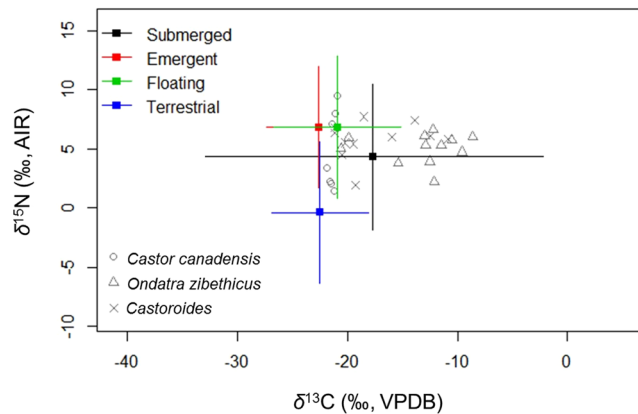


Figure 2. Stable carbon and nitrogen isotope compositions of rodent collagen and vegetation. Rodent $\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$ are corrected for trophic enrichment factors to render them comparable to the four plant functional groups (represented by their mean and a range of 2 SD) that may have comprised their diet. Carbon isotope data for modern plants and rodents are corrected for the Suess effect to enable direct comparison with results for *Castoroides*.

each rodent species was assessed using scripts from SIAR V4 in R Studio 3.1.2 (Stable Isotope Analysis in R: An Ecologist's Guide)⁴⁰.

The Proportion versus Source boxplot for *Castoroides* indicates that submerged and floating macrophytes were central to giant beaver diet, while terrestrial forage sources were utilized sparingly (Fig. 3; see Table C in Supplementary Information File S11 for plant functional group mean and range). This pattern is a striking contrast to the Proportion versus Source boxplot for *C. canadensis* (Fig. 3). This suggests that the giant beaver and its smaller cousin had complementary forage preferences and were not in direct competition for the same food resources. Overall, submerged macrophytes appear to contribute a greater proportion to the diet of the giant beaver than they do to the diet of the extant muskrat or beaver.

The Proportion versus Source boxplot for *C. canadensis* (Fig. 3) indicates a diet composed primarily of terrestrial browse, and emergent and floating macrophytes. This correlates well with what is known about modern beaver diet (extensive consumption of tree foliage and floating macrophyte rhizomes)^{62–64}.

The Proportion versus Source boxplot for *O. zibethicus* (Fig. 3) indicates a diet composed of approximately equal proportions of each macrophyte type, with a small proportion of terrestrial material. It appears that floating and submerged macrophytes are more integral to muskrat diet that previously reported^{73,74}.

The *Castoroides* $\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$ support the hypothesis that the giant beaver was a semi-aquatic rodent that grazed predominantly on aquatic macrophytes. Our $\delta^{13}\text{C}_{\text{col}}$ results are consistent with Boulanger *et al.*'s report of *Castoroides* (tooth dentin, $n = 1$) $\delta^{13}\text{C}_{\text{col}} = -21.3\text{‰}$ ⁶⁵. Additional comparisons between modern and Pleistocene *Castor* and *Ondatra* $\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$ would strengthen these conclusions. Unfortunately, there is very little published stable isotope data available from Pleistocene *Castor* and *Ondatra* specimens. There are isotopic data for a handful of samples (*Castor* $n = 2$; *Ondatra* $n = 4$) from the Aucilla River, Florida⁷⁵; however, the bone collagen carbon and nitrogen contents indicates poor preservation, and hence the isotopic results may not record original compositions. Further analyses are required to make an effective comparison.

A strong preference for submerged macrophytes rendered *Castoroides* highly dependent on wetland habitat for sustenance. The Proportion versus Source boxplots generated in SIAR strongly suggest that *C. canadensis* and *Castoroides* occupied complementary dietary niches. While they likely competed for habitat space, their forage preferences were sufficiently different to allow the two genera to co-exist across North America during the mid-to late Pleistocene. When assessing *Castoroides*' impact on the Pleistocene landscape, neither *C. canadensis* nor *O. zibethicus* constitute a perfect analogue species; however, based on $\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$ results, the muskrat is the closest extant semi-aquatic rodent that could be used to describe the diet and ecological impact of the giant beaver. These interpretations remain supported even if only $\delta^{13}\text{C}_{\text{col}}$ are compared among the three rodent species (refer to Figure D in Supplementary Information File S11).

The Proportion versus Source boxplots generated in SIAR do not lend support to the notion that *Castoroides* cut down trees or consumed foods such as tree leaves, bark or twigs. There is currently no convincing evidence of dams, lodges, or underwater food caches constructed by giant beavers from the Pleistocene sedimentological record. If the giant beaver was felling trees for food, the stable isotopic composition of its bone collagen should reflect a greater proportion of terrestrial browse (see Supplementary Information File S11 Table C for plant functional group $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ means and ranges). This conclusion is further supported by skeletal morphology, in that giant beaver incisors lack the chisel-like edge that is characteristic of *Castor* incisors⁷⁶ (Fig. 4). *C. canadensis*' ability to engineer the landscape and to create new habitat space, combined with the rise in abundance and diversity of deciduous tree species in north-eastern North America, likely gave *Castor* a distinct advantage over the giant beaver during the late Pleistocene. Hence competition for habitat space may have been a contributing factor in the extinction of the giant beaver. Testing this hypothesis, however, awaits a suitable suite of fossil remains of coeval *Castor* and *Castoroides*.

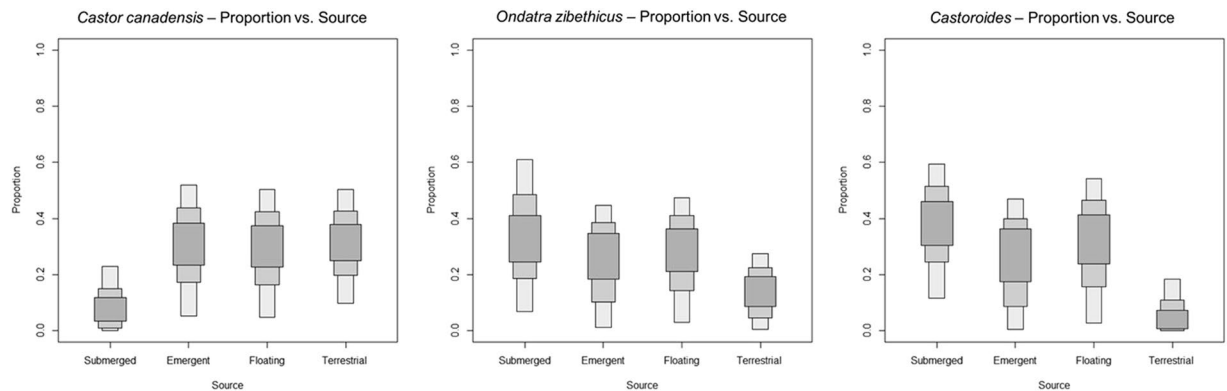


Figure 3. Proportion versus Source boxplots generated in SIAR for *C. canadensis*, *O. zibethicus*, and *Castoroides*. Each boxplot represents the relative proportion that each plant functional group (categorized here as a “Source”) contributed to the diet of each rodent species. The carbon isotope data for modern plants and rodents are corrected for the Suess effect.

Confirmation that *Castoroides* was highly dependent on wetland habitat not only for shelter from predators, but also for food allows us to assess its extinction within the context of changing environmental conditions during the late Pleistocene. The loss of both wetland habitat in lowland regions and associated open mixed-conifer forests coincide with the regional disappearance of *Castoroides* populations in many areas across North America, as is discussed next.

The first appearance of *Castoroides* in the fossil record is in Florida and dates to the Irvingtonian I NALMA (1.9 mya to 900,000 BP)^{2,77,78}. There is a more or less continuous record of *Castoroides* in the southeast, where the oldest known species of giant beaver, *Castoroides leiseyorum*, is assumed to have given rise to both *Castoroides ohioensis* and *Castoroides dilophidus*⁷⁹. Unlike *C. ohioensis*, it appears that *C. dilophidus* remained confined to the southeast (Florida, Georgia, South Carolina, and Virginia) during the Rancholabrean NALMA (240,000 to 11,000 year BP)^{79,80}. Virtually all specimens from Florida have been collected from river systems and many lack precise contextual information⁸⁰. An insecure faunal chronology, combined with scarce pollen and plant macrofossil records for the region prior to the Wisconsin Glaciation, limit our ability to discern ecological patterns over time. However, the region supported forested swamps and warm-adapted mixed forests during much of the mid-Wisconsin. Florida underwent multiple periods of increasing aridity and more seasonal precipitation patterns throughout the Last Glacial Maximum (approximately 24,000 years BP)⁸¹. Scrub and prairie conditions arose in parts of Georgia, the Coastal Plain, and in southern Florida during both Wisconsin glacial and post-glacial times⁸². Moisture conditions that favoured pine forests and swampland did not return until mid- to late Holocene, long after the extinction of the giant beaver^{81–83}.

The first records of *Castoroides* in the Great Plains region are from Kansas and date to the Irvingtonian NALMA. However, the transition from an ancestral taxon *Procastoroides* (uncrenulated incisors) to *Castoroides* (crenulated incisors) probably occurred on the Great Plains during either the late Pliocene or the early Pleistocene during the Blancan NALMA^{2,84}. Giant beaver populations became well-established across Kansas, Nebraska, and Oklahoma during the late Irvingtonian and early Rancholabrean NALMA^{1,80}. After onset of the Illinoian glaciation, giant beaver fossils disappeared from the Great Plains record and became concentrated east of the Mississippi River. The disappearance of no-analog forest communities, the regional extinction of certain conifer tree species, and the appearance of grass or herb plant communities all correlate to increasing aridity and coincide with the disappearance of the giant beaver in the region^{85–87}.

In the north, *Castoroides* and *Castor* fossils are both found in lake bed and river deposits from the unglaciated region of Old Crow, Yukon Territory⁸⁸. Giant beavers most likely inhabited Yukon Territory and Alaska during global warm periods between 200,000 to 75,000 years BP^{89,90}. This chronology is similar to that of the American mastodon (*Mammot americanum*) and western camel (*Camelops hesternus*) and are interpreted to represent northward dispersals during the relatively warm Sangamonian interglaciation^{49,91}. The presence of *Castoroides* fossils from *in situ* deposits correlative to Marine Isotope Stage 7 (~200,000 years BP) suggests there were repeated northward dispersals into the Arctic during at least two separate interglaciations⁸⁹. Giant beaver populations could only have spread northward during these periods of warm climate, where the retreat of the ice sheets left a string of meltwater wetlands, and boreal forests and wet-tundra were established regionally. The presence of *Castoroides* in these arctic latitudes suggests they were capable of surviving the cold dark winters of interglacial periods, though regional aridification and establishment of cryoxerophilous steppe tundra vegetation probably created environments that were not habitable for them during glacial periods. It is highly likely that the giant beaver was regionally extirpated with the establishment of Wisconsin glacial conditions around 70,000 years BP and subsequently all populations were restricted to areas south of the continental ice sheets until their final demise at the end of the Pleistocene⁹⁰.

Radiocarbon dates from Ohio and New York indicate that the Great Lakes Basin was home to the last known population of giant beavers⁶⁵. Fossils from the Sangamonian are also occasionally found. The population was concentrated south of the fluctuating continental ice sheet margin during the Wisconsin glacial period prior to



Figure 4. Left: A near-complete *Castoroides ohioensis* mandible and lower incisor from Clear Creek Township, Ohio, USA. Specimen OHS N9109. Right: A complete *Castoroides ohioensis* upper incisor from Old Crow, Yukon Territory, Canada. Specimen CMN 51270. Photographs by T. Plint.

the extinction of the genus^{1,17–19,65,66}. The giant beaver disappeared from Eastern North America shortly before the Pleistocene–Holocene transition⁶. With the disappearance of this population, the genus became globally extinct.

Wisconsinan age giant beaver fossils are more commonly associated with sediments from warmer interstadial times, when the retreat of continental ice sheets left the Great Lakes region flooded with meltwater. The cool, moist climate during the mid-Wisconsinan provided ideal habitat space in the form of extensive lacustrine environments surrounded by forest communities dominated by spruce and larch^{19,58}. During the onset of the Holocene warm period, much of the shallow wetland habitat in Eastern North America became in-filled with sediment and deciduous trees replaced much of the open mixed-conifer forest communities⁹².

The only known instance of direct temporal and spatial overlap between human artifacts and *Castoroides* fossils occurs in New York State. The current youngest known *Castoroides* specimen (dating to $10,150 \pm 50$ years BP) indicates that megafauna populations overlapped with Palaeoindian culture for up to a thousand years⁶. However, there is no current zooarchaeological evidence that humans butchered, hunted, or otherwise utilized the giant beaver as a resource.

Conclusions

Castoroides $\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$ indicate that giant beaver diet was composed predominantly of macrophytes (particularly submerged macrophytes), rendering the genus highly dependent on wetland habitat for survival. The presence of giant beaver fossils coincides with palaeoenvironmental conditions that supported ample swamps and shallow lakes, and such remains disappear in settings associated with the arrival of warmer, more arid conditions. Sediment infilling, loss of glacial meltwater, the onset of more seasonal precipitation patterns, and increased temperature all contributed to wetland habitat loss across North America during this time. Without apparent refugia, *Castoroides* dwindled to an isolated population located in the lowlands south of the Great Lakes, where potential competition for habitat space and ongoing profound climate change contributed to their extinction.

Methodology

Rodent sample material. *Castoroides* specimens originating from Yukon Territory, Canada, and Ohio, USA, were included to determine regional differences in diet (Fig. 1). Modern muskrat and beaver skeletal material were included to explore similarities in ecomorphology and the accuracy of the SIAR mixing model. To facilitate comparison, the modern *O. zibethicus* and *C. canadensis* samples were sought from similar locations as the *Castoroides* fossils (Fig. 1, Tables 3 to 5). Modern skeletal materials were donated by conservation authorities, trappers, and ecological research groups.

Radiocarbon dating. Radiocarbon (^{14}C) dating of *Castoroides* bone samples collected in Yukon Territory was performed at the Keck Carbon Cycle AMS Facility, University of California Irvine. The Keck laboratory employed ultrafiltration of the gelatinized collagen at 30 kDa, and a second time at 3 kDa to remove exogenous organic material⁹³. Radiocarbon dating of *Castoroides* bone samples collected in Ohio was performed by the Accelerator Mass Spectrometry Laboratory, University of Arizona using a modified Longin bulk collagen extraction method without ultrafiltration⁹⁴.

Collagen stable isotope analysis. Detailed information regarding collagen sample preparation and stable isotope analysis is provided in Supplementary Information File S11. In brief, collagen was extracted using a modified Longin method⁹⁴. Crushed sample material underwent lipid extraction, followed by slow demineralization in a weak acid, and a final soak in a weak base to remove humic acids. Gelatinized collagen was analyzed using a Costech Elemental Analyzer coupled with a Thermo Scientific Delta Plus XL isotope ratio mass spectrometer operated in continuous-flow mode with helium as the carrier gas. Stable carbon and nitrogen isotope results are reported relative to VPDB and AIR, respectively.

Stable isotope analysis of plants. An assessment of bulk plant carbon and nitrogen isotope signatures was undertaken to provide the necessary context to interpret *Castoroides*, *C. canadensis* and *O. zibethicus* $\delta^{13}\text{C}_{\text{col}}$ and $\delta^{15}\text{N}_{\text{col}}$. Supplementary Information File S11 provides detailed information regarding plant sample taxonomic

identification, preparation, and stable isotope analysis performed for this purpose. We analyzed four plant functional groups: terrestrial trees and shrubs, emergent macrophytes, floating macrophytes, and submerged macrophytes. These groupings were chosen to (1) identify plants with access to similar pools of bioavailable carbon and nitrogen, and (2) to gauge *Castoroides*' level of dependence on freshwater versus terrestrial food resources. Similar plant functional groupings are utilized in studies of modern *C. canadensis*^{54,95}.

Modern plant samples were collected from shallow lakes, marshes, and creeks located in southwestern Ontario (Pinery Provincial Park and the London area) and Yukon Territory (Whitehorse and Old Crow Basin), Canada (Fig. 1).

The Ontario sites experience a temperate climate, and with annual average precipitation of approximately 1000 mm. Pinery Provincial Park is located on the south-east shore of Lake Huron. The park consists of the northern-most extension of the Oak Savannah ecosystem and overlays a succession of ancient dune ridges. London is located approximately 70 km east of Pinery Provincial Park, and is located in an area of Quaternary sediments (glacial till interbedded with interglacial lake sediments) overlaying Paleozoic limestone bedrock. The sites in Yukon Territory experience a subarctic climate, where annual average precipitation is between 200 and 300 mm. Whitehorse is located within the Whitehorse trough (an intermontane basin) in south-central Yukon Territory. The Old Crow community is located in a region of continuous permafrost within the Arctic Circle and is situated along the southern fringe of the Old Crow Flats (a plain filled with thermokarst lakes). The region is surrounded by adjacent mountain ranges and remained unglaciated during the Quaternary.

Data Availability

The authors declare no limitations on data or standard operating protocol availability.

References

- Cahn, A. Records and distribution of the fossil beaver, *Castoroides ohioensis*. *J. Mammal.* **12**, 229–241 (1932).
- Martin, R. A. Taxonomy of the giant Pleistocene beaver *Castoroides* from Florida. *J. Paleontol.* **43**, 1033–1041 (1969).
- Reynolds, P. S. How big is a giant? The importance of method in estimating body size of extinct mammals. *J. Mammal.* **83**, 321–332 (2002).
- Barnosky, A. D., Koch, P. L., Feranec, R. S., Wing, S. L. & Shabel, A. B. Assessing the causes of late Pleistocene extinctions on the continents. *Science* **306**, 70–75 (2004).
- Faith, J. T. & Surovell, T. A. Synchronous extinction of North America's Pleistocene mammals. *Proc. Natl. Acad. Sci.* **106**, 20641–20645 (2009).
- Boulanger, M. T. & Lyman, R. L. Northeastern North American Pleistocene megafauna chronologically overlapped minimally with Paleoindians. *Quat. Sci. Rev.* **85**, 35–46 (2014).
- Cohen, T. J. *et al.* Hydrological transformation coincided with megafaunal extinction in central Australia. *Geology* **43**, 195–198 (2015).
- Cooper, A. *et al.* Abrupt warming events drove Late Pleistocene Holarctic megafaunal turnover. *Science* **349**, 602–606 (2015).
- Rabanus-Wallace, M. T. *et al.* Megafaunal isotopes reveal role of increased moisture on rangeland during late Pleistocene extinctions. *Nat. Ecol. Evol.* **1**, 1–5 (2017).
- Guthrie, R. D. New carbon dates link climatic change with human colonization and Pleistocene extinctions. *Nature* **441**, 207–209 (2006).
- Louys, J., Curnoe, D. & Tong, H. Characteristics of Pleistocene megafauna extinctions in Southeast Asia. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **243**, 152–173 (2007).
- Pushkina, D. & Raia, P. Human influence on distribution and extinctions of the late Pleistocene Eurasian megafauna. *J. Hum. Evol.* **54**, 769–782 (2008).
- Reynolds, P. S. Size, shape, and surface area of beaver, *Castor canadensis*, a semiaquatic mammal. *Can. J. Zool.* **71**, 876–882 (1993).
- Perkins, B. S. Ancient beavers did not eat trees. *Science News* **176**, 28–29 (2009).
- Stuart-Williams, H. L. Q. & Schwarcz, H. P. Oxygen isotopic determination of climatic variation using phosphate from beaver bone, tooth enamel, and dentine. *Geochim. Cosmochim. Acta* **61**, 2539–2550 (1997).
- Harrington, C. R. The giant beaver. *Neotoma* **20** (1986).
- Holman, J. A. *Ancient life of the Great Lakes Basin: Precambrian to Pleistocene*. (University of Michigan Press, 1995).
- Gregory McDonald, H. & Bryson, R. A. Modeling Pleistocene local climatic parameters using macrophysical climate modeling and the paleoecology of Pleistocene megafauna. *Quat. Int.* **217**, 131–137 (2010).
- Swinehart, A. L. & Richards, R. L. Palaeoecology of a northeast Indiana wetland harboring remains of the Pleistocene giant beaver (*Castoroides ohioensis*). *Proc. Indiana Acad. Sci.* **110**, 151–166 (2001).
- Kurtén, B. & Anderson, E. *Pleistocene Mammals of North America*. (Columbia University Press, 1980).
- Stirton, R. A. Cranial morphology of *Castoroides*. *Mining and Metallurgical Institute, Dr. D. N. Wadia Commemorative Volume*, 273–285 (1965).
- Hay, O. P. The recognition of Pleistocene faunas. *Smithson. Inst* (1912).
- McDonald, H. G. The late Pleistocene vertebrate fauna in Ohio: co-inhabitants with Ohio's Paleoindians. The first discovery of America: Archaeological evidence of the early inhabitants of the Ohio area. *Ohio Archaeol. Council* **23–39** (1994).
- Moore, J. Concerning a skeleton of the great fossil beaver. *Castoroides ohioensis*. *Cincinnati Soc. Nat. Hist.* **13**, 138–169 (1890).
- Miller, R. F., Harrington, C. R. & Welch, R. A giant beaver fossil (*Castoroides ohioensis* Foster) fossil from New Brunswick, Canada. *Atlantic Geology* **36**, 1–5 (2000).
- Lovegrove, B. G. & Mowoe, M. O. The evolution of mammal body sizes: Responses to Cenozoic climate change in North American mammals. *J. Evol. Biol.* **26**, 1317–1329 (2013).
- Anderson, E. *Who's who in the Pleistocene. A mammalian bestiary*. (University of Arizona Press, 1984).
- Hillerud, J. M. New specimens of *Castoroides ohioensis* from Ohio. In *Abstracts: Annual meeting of Society of Vertebrate Paleontology. Museum of Comparative Zoology, Cambridge, Mass* (1975).
- Rybczynski, N. Castorid phylogenetics: Implications for the evolution of swimming and tree-exploitation in beavers. *J. Mamm. Evol.* **14**, 1–35 (2007).
- Rybczynski, N. Woodcutting behavior in beavers (Castoridae, Rodentia): estimating ecological performance in a modern and a fossil taxon. *Paleobiology* **34**, 389–402 (2008).
- Tedford, R. H. & Harrington, C. R. An Arctic mammal fauna from the Early Pleistocene of North America. *Nature* **425**, 388–390 (2003).
- Kelly, J. F. Stable isotopes of carbon and nitrogen in the study of avian and mammalian trophic ecology. *Can. J. Zool.* **78**, 1–27 (2000).
- Bocherens, H. & Drucker, D. Trophic level isotopic enrichment of carbon and nitrogen in bone collagen: Case studies from recent and ancient terrestrial ecosystems. *Int. J. Osteoarchaeol.* **13**, 46–53 (2003).

34. Krajcarz, M. T., Krajcarz, M. & Bocherens, H. Collagen-to-collagen prey-predator isotopic enrichment ($\Delta^{13}\text{C}$, $\Delta^{15}\text{N}$) in terrestrial mammals - a case study of a subfossil red fox den. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **490**, 563–570 (2018).
35. Bocherens, H. Isotopic biogeochemistry and the paleoecology of the mammoth steppe fauna. *Deinsea* **9**, 57–76 (2003).
36. Coltrain, J. B. *et al.* Rancho la Brea stable isotope biogeochemistry and its implications for the paleoecology of late Pleistocene, coastal southern California. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **205**, 199–219 (2004).
37. Fox-Dobbs, K., Leonard, J. A. & Koch, P. L. Pleistocene megafauna from eastern Beringia: Paleoeological and paleoenvironmental interpretations of stable carbon and nitrogen isotope and radiocarbon records. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **261**, 30–46 (2008).
38. Metcalfe, J. Z., Longstaffe, F. J. & Hodgins, G. Proboscideans and paleoenvironments of the Pleistocene Great Lakes: Landscape, vegetation, and stable isotopes. *Quat. Sci. Rev.* **76**, 102–113 (2013).
39. Casey, M. M. & Post, D. M. The problem of isotopic baseline: Reconstructing the diet and trophic position of fossil animals. *Earth-Science Rev.* **106**, 131–148 (2011).
40. Inger, R., Jackson, A., Parnell, A. & Bearhop, S. SIAR V4: Stable Isotope Analysis in R. An Ecologist's Guide (2010).
41. Chappuis, E., Serin, V., Marti, E., Ballesteros, E. & Gacia, E. Decrypting stable-isotope ($\delta^{13}\text{C}$ and $\delta^{15}\text{N}$) variability in aquatic plants. *Freshw. Biol.* **62**, 1807–1818 (2017).
42. Keeley, J. E. & Sandquist, D. R. Carbon: freshwater plants. *Plant. Cell Environ.* **15**, 1021–1035 (1992).
43. Mendonça, R. *et al.* Bimodality in stable isotope composition facilitates the tracing of carbon transfer from macrophytes to higher trophic levels. *Hydrobiologia* **710**, 205–218 (2013).
44. Long, E. S., Sweitzer, R. A., Diefenbach, D. R. & Ben-David, M. Controlling for anthropogenically induced atmospheric variation in stable carbon isotope studies. *Oecologia* **146**, 148–156 (2005).
45. Lee, K. Y., Graham, L., Spooner, D. E. & Xenopoulos, M. A. Tracing anthropogenic inputs in stream food webs with stable carbon and nitrogen isotope systematics along an agricultural gradient. *PLoS One* **13**, 1–20 (2018).
46. Tahmasebi, F., Longstaffe, F. J. & Zazula, G. Nitrogen isotopes suggest a change in nitrogen dynamics between the late Pleistocene and modern time in Yukon, Canada. *PLoS One* **13** (2018).
47. Tahmasebi, F., Longstaffe, F. J., Zazula, G. & Bennett, B. Nitrogen and carbon isotopic dynamics of subarctic soils and plants in southern Yukon Territory and its implications for paleoecological and paleodietary studies. *PLoS One* **12** (2017).
48. Newsom, L. A. & Muhlbachler, M. C. Mastodons (*Mammot americanum*) diet foraging patterns based on analysis of dung deposits, In *First Floridians Last Mastodons Page-Ladson Site Aucilla River 263–331*. (Springer, Dordrecht, 2006).
49. Zazula, G. D. *et al.* American mastodon extirpation in the Arctic and Subarctic predates human colonization and terminal Pleistocene climate change. *Proc. Natl. Acad. Sci.* **111**, 18460–18465 (2014).
50. France, C. A. M., Zelanko, P. M., Kaufman, A. J. & Holtz, T. R. Carbon and nitrogen isotopic analysis of Pleistocene mammals from the Saltville Quarry (Virginia, USA): Implications for trophic relationships. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **249**, 271–282 (2007).
51. Schwartz-Narbonne, R., Longstaffe, F. J., Metcalfe, J. Z. & Zazula, G. Solving the woolly mammoth conundrum: Amino acid $\delta^{15}\text{N}$ -enrichment suggests a distinct forage or habitat. *Sci. Rep.* **5**, 1–6 (2015).
52. van Klinken, G. J. Bone collagen quality indicators for paleodietary and radiocarbon measurements. *J. Archaeol. Sci.* **26**, 687–695 (1999).
53. Hornibrook, E. R. C., Longstaffe, F. J., Fyfe, W. S. & Bloom, Y. Carbon-isotope ratios and carbon, nitrogen and sulfur abundances in flora and soil organic matter from a temperate-zone bog and marsh. *Geochim. J.* **34**, 237–245 (2000).
54. Milligan, H. E., Pretzlaw, T. D. & Humphries, M. M. Stable isotope differentiation of freshwater and terrestrial vascular plants in two subarctic regions. *Écoscience* **17**, 265–275 (2010).
55. Russell, S. D. J. *Nitrate sources in the Old Ausable River Channel and adjacent aquifers in Pinery Provincial Park*. PhD dissertation, The University of Western Ontario, Department of Earth Sciences (2015).
56. Bateman, A. S. & Kelly, S. D. Fertilizer nitrogen isotope signatures. *Isotopes Environ. Health Stud.* **43**, 237–247 (2007).
57. Government of Canada. *Statistics Canada*, <https://www.statcan.gc.ca> (2018).
58. Berti, A. A. Paleobotany of Wisconsin interstadials, eastern Great Lakes region, North America. *Quat. Res.* **5**, 591–619 (1975).
59. Zazula, G. D., Froese, D. G., Westgate, J. A., La Farge, C. & Mathewes, R. W. Paleocology of Beringian 'packrat' middens from central Yukon Territory, Canada. *Quat. Res.* **63**, 189–198 (2005).
60. Zazula, G. D., Froese, D. G., Elias, S. A., Kuzmina, S. & Mathewes, R. W. Arctic ground squirrels of the mammoth-steppe: paleocology of Late Pleistocene middens (~24 000–29 450 14C yr BP), Yukon Territory, Canada. *Quat. Sci. Rev.* **26**, 979–1003 (2007).
61. Busher, P. E. Food Caching Behavior of Beavers (*Castor canadensis*): Selection and use of woody species. *Am. Midl. Nat.* **135**, 343 (1996).
62. Muller-Schwarze, D. *The beaver: its life and impact*. (Cornell University Press, 2011).
63. Milligan, H. E. & Humphries, M. M. The importance of aquatic vegetation in beaver diets and the seasonal and habitat specificity of aquatic-terrestrial ecosystem linkages in a subarctic environment. *Oikos* **119**, 1877–1886 (2010).
64. Severud, W. J., Windels, S. K., Belant, J. L. & Bruggink, J. G. The role of forage availability on diet choice and body condition in American beavers (*Castor canadensis*). *Mamm. Biol.* **78**, 87–93 (2013).
65. Boulanger, M. T., Lattanzi, G. D., Parris, D. C., O'Brien, M. J. & Lyman, R. L. AMS radiocarbon dates for Pleistocene fauna from the American Northeast. *Radiocarbon* **57**, 189–192 (2015).
66. McDonald, H. G. & Glotzhober, R. C. New radiocarbon dates for the giant beaver, *Castoroides ohioensis* (Rodentia, Castoridae), from Ohio and its extinction. In *Unlocking the Unknown: Papers Honoring Dr. Richard Zakrzewski* 51–59 (2008).
67. Collins, M. J. *et al.* The survival of organic matter in bone: A review. *Archaeometry* **3**, 383–394 (2002).
68. Keeling, R. F., Walker, S. J., Piper, S. C., & Bollenbacher, A. F. Scripps CO_2 Program. Scripps Institution of Oceanography, University of California, <http://scrippsco2.ucsd.edu>. (2014).
69. Schmitt, J. *et al.* Carbon isotope constraints on the deglacial CO_2 rise from ice cores. *Science* **336**, 711–714 (2012).
70. DeNiro, M. J. & Epstein, S. Influence of diet on the distribution of carbon isotopes in animals. *Geochim. Cosmochim. Acta* **42**, 495–506 (1978).
71. DeNiro, M. J. & Epstein, S. Influence of diet on the distribution of nitrogen isotopes in animals. *Geochim. Cosmochim. Acta* **44**, 341–351 (1981).
72. Froehle, A. W., Kellner, C. M. & Schoeninger, M. J. FOCUS: Effect of diet and protein source on carbon stable isotope ratios in collagen: follow up to *J. Archaeol. Sci.* **37**, 2662–2670 (2009).
73. Campbell, K. L. & MacArthur, R. A. Seasonal changes in gut mass, forage digestibility, and nutrient selection of wild muskrats (*Ondatra zibethicus*). *Physiol. Zool.* **69**, 1215–1231 (1996).
74. Danell, K. Reduction of aquatic vegetation following the colonization of a northern Swedish lake by the muskrat, *Ondatra zibethica*. *Oecologia* **38**, 101–106 (1979).
75. France, C. A. M. *A carbon and nitrogen isotopic analysis of Pleistocene food webs in North America: implications for paleoecology and extinction*. PhD dissertation, University of Maryland, Department of Geology (2008).
76. Rinaldi, C., Martin, L., Cole, T. III. & Timm, R. Occlusal wear morphology of giant beaver (*Castoroides*) lower incisors: functional and phylogenetic implications. In *Abstracts Society of Vertebrate Palaeontology* (2009).

77. Morgan, G. S. & White, J. A. Small mammals (Insectivora, Lagomorpha, and Rodentia) from the early Pleistocene (Irvingtonian) Leisey Shell Pit Local Fauna, Hillsborough County, Florida. *Bull. Florida Museum. Nat. Hist.* **37**, 397–461 (1995).
78. Parmalee, P. W. & Graham, R. W. Additional records of the giant beaver, *Castoroides*, from the Mid-South: Alabama, Tennessee, and South Carolina. *Smithson. Contrib. to Paleobiol.* **93** (2002).
79. Hulbert, R., Kerner, A. & Morgan, G. S. Taxonomy of the Pleistocene giant beaver *Castoroides* (Rodentia: Castoridae) from the southeastern United States. *Bull. Florida Mus. Nat. Hist.* **53**, 26–43 (2014).
80. Graham, R. W. FAUNMAP: a database documenting late Quaternary distributions of mammal species in the United States. *Illinois State Museum* **25** (1994).
81. Watts, W. A. B. C. S. & Hansen, B. A. P. Environments of Florida in the late Wisconsin and Holocene. In *Wet site archaeology* 307–323 (1988).
82. Watts, W. A. The late Quaternary vegetation history of the southeastern United States. *Annu. Rev. Ecol. Syst.* **11**, 387–410 (1980).
83. Koch, P. L., Hoppe, K. A. & Webb, S. D. The isotopic ecology of late Pleistocene mammals in North America. *Chem. Geol.* **152**, 119–138 (1998).
84. Woodburne, M. Upper Pliocene geology and vertebrate paleontology of part of the Meade Basin, Kansas. *Papers of the Michigan Academy of Science, Arts, and Letters* **46** (1961).
85. Jackson, S. T. *et al.* Vegetation and environment in Eastern North America during the Last Glacial Maximum. *Quat. Sci. Rev.* **19**, 489–508 (2000).
86. Jackson, S. T. & Weng, C. Late Quaternary extinction of a tree species in eastern North America. *Proc. Natl. Acad. Sci. USA* **96**, 13847–13852 (1999).
87. Strong, W. L. & Hills, L. V. Late-glacial and Holocene palaeovegetation zonal reconstruction for central and north-central North America. *J. Biogeogr.* **32**, 1043–1062 (2005).
88. Harington, C. R. Pleistocene vertebrates of the Yukon Territory. *Quat. Sci. Rev.* **30**, 2341–2354 (2011).
89. Froese, D. *et al.* Fossil and genomic evidence constrains the timing of bison arrival in North America. *Proc. Natl. Acad. Sci.* **114**, 3457–3462 (2017).
90. Zazula, G. D., Froese, D. G., Elias, S. A., Kuzmina, S. & Mathewes, R. W. Early Wisconsinan (MIS 4) Arctic ground squirrel middens and a squirrel-eye-view of the mammoth-steppe. *Quat. Sci. Rev.* **30**, 2220–2237 (2011).
91. Zazula, G. D. *et al.* A case of early Wisconsinan “over-chill”: New radiocarbon evidence for early extirpation of western camel (*Camelops hesternus*) in eastern Beringia. *Quat. Sci. Rev.* **171**, 48–57 (2017).
92. Jacobson, G. L. Jr., Webb, T. III. & Grimm, E. C. Patterns and rates of vegetation change during the deglaciation of eastern North America. *North Am. and Adjac. Ocean. Dur. Last Deglaciation K-3*, 277–288 (1987).
93. Beaumont, W., Beverly, R., Southon, J. & Taylor, R. E. Bone preparation at the KCCAMS Laboratory. *Nucl. Instruments Methods Phys. Res.* **268**, 906–909 (2010).
94. Longin, R. New method of collagen extraction for radiocarbon dating. *Nature* **230**, 241–242 (1971).
95. Severud, W. J., Belant, J. L., Windels, S. K. & Bruggink, J. G. Seasonal Variation in Assimilated Diets of American Beavers. *Am. Midl. Nat.* **169**, 30–42 (2013).
96. Steadman, D. W., Stafford, T. W. & Funk, R. E. Nonassociation of Paleoindians with AMS-dated late Pleistocene mammals from the Dutchess Quarry Caves, New York. *Quat. Res.* **47**, 105–116 (1997).
97. Feranec, R. S. & Kozłowski, A. L. AMS radiocarbon dates from Pleistocene and Holocene mammals housed in the New York State Museum, Albany, New York, USA. *Radiocarbon* **52**, 205–208 (2010).
98. Erickson, B. R. Paleontological evidence concerning some post glacial features of the Mississippi River Valley. *Sci. Pub. Sci. Mus.* **1**, 1–4 (1967).
99. Tankersley, K. B. & Landefeld, C. S. Geochronology of the Sheriden Cave, Ohio: The 1997 Field Season. *Curr. Res. Pleist.* **15**, (136–138 (1998).

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Author Contributions

T.P. conducted the sampling and isotopic analyses, interpreted the data, and wrote the initial draft of the manuscript. F.J.L. and T.P. conceived the research design. G.Z. provided funding for some of the radiocarbon analyses. F.J.L. and G.Z. revised the manuscript and provided valuable comments and advice. F.J.L. provided funding to support the research.

Additional Information

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